

1 **The Bone Metabolic Response to Exercise and Nutrition**

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11 **Short Title:** The bone metabolic response to exercise

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21 **ABSTRACT:**

22 Bone (re)modelling markers can help determine how bone responds to different types, intensities and
23 durations of exercise. They might also help predict those at risk of bone injury. We synthesised
24 evidence on the acute and chronic bone metabolic responses to exercise, along with how nutritional
25 factors can moderate this response. Recommendations to optimise future research efforts are made.

26 **IN BRIEF:**

27 Bone (re)modelling markers elucidate the dynamic bone response to exercise and if used
28 appropriately have large potential to progress understanding.

29 **KEY WORDS:** bone remodelling, resorption, formation, exercise, turnover, loading, metabolism.

30 **KEY POINTS:**

- 31 • Bone (re)modeling markers (BMMs) are products of bone proteins or cells, and represent
32 processes involved in either the formation or resorption of bone.
- 33 • The stimuli (both mechanical and metabolic) created by an acute exercise bout, typically elicits
34 an increase in markers indicative of bone resorption (*e.g.*, β -CTX-1), whilst adaptation to
35 exercise training typically results in an increase in bone formation (*e.g.*, PINP).
- 36 • Nutritional status, and acute nutrient intake, can moderate the bone metabolic response to
37 exercise.
- 38 • Appropriate use of these biomarkers, in well-controlled settings, has the potential to progress
39 knowledge on the acute, or short-term, responses of bone to exercise and nutritional stimuli,
40 and so to contribute toward the development of strategies to protect or enhance the bone
41 health of exercising individuals.

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45 **1. INTRODUCTION**

46 The bone response to exercise is complex and influenced by multiple factors, including nutrition;
47 training status; age; genetics and the characteristics of the specific exercise stimulus. Exercise is
48 typically beneficial to bone, and sports that convey high-impact, multi-directional movement patterns,
49 and unaccustomed loads, are widely accepted as providing the optimal osteogenic stimulus (1).
50 Accordingly, exercise is considered an effective preventive or treatment strategy for those with
51 conditions characterised by bone loss, or increased fracture susceptibility (*e.g.*, osteoporosis) (2).
52 Conversely, participation in sports involving lower-impact and/or repetitive loading cycles (such as
53 endurance running) or non-weight bearing sports (such as cycling and swimming) do not typically elicit
54 skeletal benefits (3,4). Indeed some groups of athletes (*e.g.*, cyclists and jockeys) have lower BMD
55 than non-athletic controls, implying a negative influence of some types, or volumes, of exercise on
56 bone (5,6).

57 Much remains unknown about factors influencing the bone response to acute and chronic exercise,
58 or how to pre-emptively identify those at risk of bone injuries. To elucidate these factors, objective
59 and quantifiable indicators of bone strength and function are essential. BMD [assessed by dual energy
60 x-ray absorptiometry (DXA)] or bone microarchitecture [assessed by high-resolution peripheral
61 quantitative computed tomography (HR-pQCT)] may be used to predict fracture risk (7–9) or to assess
62 intervention efficacy. These outcomes are, however, chronic indicators of bone, which responds
63 slowly to stimuli. Measurable changes can take months, or even years, to occur; and so acute or
64 shorter-duration responses cannot be detected using these measures.

65 In contrast, bone (re)modelling markers (BMMs) provide information about dynamic bone activity and
66 can indicate the acute or short-term response to stimuli, and their potential to progress knowledge
67 on this topic is large. This potential cannot currently be realised, however, due to incomplete
68 understanding of their physiological relevance, however, along with large heterogeneity in study
69 design and characteristics. The aim of this review is to consolidate understanding of the acute and

70 chronic BMM response to exercise, and to make recommendations to optimise the use of appropriate
71 biomarkers in future studies. Additionally, we will describe how nutritional interventions moderate
72 the BRM response to exercise, and how this information can elucidate the mechanistic pathways
73 mediating these responses.

74 **2. THE BONE METABOLIC RESPONSE TO EXERCISE**

75 Provided the nutritional and metabolic environment is favourable, the primary stimulus for bone
76 anabolism is physical loading (10,11), with bone responding to the magnitude, rate, number and
77 direction of activity-induced loading-cycles (12). As such, different exercise modalities exert distinct
78 loading patterns and activity-specific mechanotransductive signals (13). Various metabolic signals also
79 influence the bone response to exercise, such as reactive oxygen/nitrogen species (14), altered pH
80 (15) and serum calcium availability (16). Modelling refers to the formation or resorption of bone at
81 specific sites. In contrast, remodelling is a coupled and synchronized process of bone activation,
82 resorption, reversal and formation, which is co-ordinated by teams of bone cells (*i.e.*, osteoblasts,
83 osteoclasts and osteocytes) termed the basic multicellular unit (BMU). Although some modelling
84 cannot be ruled out, it seems that remodelling is the dominant process through which bone responds
85 to the mechanical or metabolic stimuli offered by exercise (12,17). An overview of this process is
86 shown in Figure 1.

87 ***2.1. The use of bone (re)modelling markers in sport and exercise science***

88 BMMs are products of bone proteins or cells and mostly represent processes involved in either the
89 formation or resorption of bone (see Table 1 for an overview of commonly used BMMs). Their
90 potential to elucidate the mechanisms through which bone responds to exercise is large, but some
91 factors impede this interpretation, if not considered in study design and interpretation. It is important
92 to understand that many BMMs (*e.g.*, PINP, OC, OPG and PYD) are non-bone-specific, which renders
93 mechanistic interpretation challenging. For example, some biomarkers (*e.g.*, PINP, PYR, DYP and ICTP)

94 are products of collagen metabolism, which is the main structural protein of many connective tissues,
95 and not only the bone. As such, their measurement is not necessarily indicative of altered bone activity
96 only. Similarly, osteocalcin (OC) is a small non-collagenous protein synthesised by osteoblasts, which
97 is often used to estimate osteoblast activity and, therefore, bone formation. But both intact and
98 fragmented OC may also be released during bone resorption (18), suggesting that this biomarker may
99 indicate general bone remodelling rather than bone formation specifically. Additionally, OC is a non-
100 specific protein that fulfils a number of extra-skeletal roles, including functions in energy metabolism
101 and muscle activity (19). These extra-skeletal roles are particularly relevant when interpreting the OC
102 response to exercise, given that bioenergetic pathways and muscle activity are also upregulated by
103 exercise. Thus, it is difficult to attribute any changes in circulating OC to altered bone activity.

104 The repeatability of BMM measurement is another important consideration, as some show substantial
105 inter- and intra-individual variability, and/or, are difficult to accurately measure. For example, the
106 osteoprotegerin/receptor activator of NF kappaB ligand (OPG/RANKL) ratio is commonly used to
107 indicate bone resorption, but soluble RANKL is sometimes difficult to accurately measure *in vivo* (20),
108 and so results may be mis-leading. Bone biomarkers are often described as representing “bone
109 turnover”. Calculations, such as the uncoupling index, are commonly used to represent the
110 predominant state of bone metabolism, whereby resorptive activities that are “coupled” with
111 formation would represent a state of equilibrium, whereas “uncoupling” occurs when formative and
112 resorptive processes are unbalanced and favour either the loss or gain of bone. Some caution should
113 be applied when considering such calculations, because BMMs are systemic and cannot indicate bone
114 activity at any one particular site, which is an issue because the true bone response to loading is largely
115 site specific (21). A wide range of potentially confounding factors also impact BMMs and must be
116 accounted for in study design and interpretation. Bone is responsive to both acute and chronic
117 nutritional stimuli (described in Section 3) and so nutritional status must be carefully controlled in
118 exercise trials. Other factors, including sex, age (22), menstrual cycle phase (23), oral contraceptive
119 use (24), seasonal (25) and circadian (26) variations, genetics (27), various medical conditions and

120 medications (28) and injury history, particularly previous fractures (29,30), prior exercise and pre-
121 analytical storage and handling (31) may also influence BMMs.

122 Notwithstanding these considerations, the clinical and mechanistic relevance of these biomarkers is
123 well recognised, and in an attempt to optimise their clinical utility, the National Bone Health Alliance
124 (NBHA) advised that all studies should include, as a minimum, measurements of N-terminal
125 propeptide of type 1 procollagen (PINP) and the C-terminal telopeptide of type 1 collagen (β -CTX-I),
126 as indicators of bone formation and resorption (32–34). The decision to focus on two biomarkers was
127 made to allow for greater harmonisation, and therefore comparability, of ongoing research efforts.
128 These particular biomarkers were selected based upon the recommendations of an expert working
129 group of the International Osteoporosis Foundation and the International Federation of Clinical
130 Chemistry and Laboratory Medicine, who deemed them to have a relatively smaller biological
131 variability, higher specificity to bone metabolism and to be more responsive to anti-resorptive or
132 anabolic treatments, than other available BRMs (reviewed in detail in 33,35). Considering the currently
133 available information, we concur with this recommendation, and support the use of PINP and β -CTX-
134 I as core components of biomarker panels used to investigate the bone metabolic response to exercise
135 and nutrition interventions.

136

137 ***2.2. The bone metabolic response to an acute bout of exercise***

138 Increased bone resorption is the initial response to an acute bout of exercise, and increased β -CTX-I
139 has been reported in a number of trials (36–40). This finding is consistent with what we assume about
140 bone remodelling, whereby osteoclast activation, induced by mechanical or metabolic signals,
141 activates the BMU, causing an acute increase in bone resorption. This was shown in response to
142 different exercise types, including treadmill running (39), intense cycling (36) and a fatiguing bilateral
143 jump protocol (40). In contrast, bone formation markers seem to be largely non-responsive to acute

144 exercise, with most studies reporting no change to serum PINP, even when high-intensity exercise
145 protocols were used (40–43). This finding aligns with the bone remodelling process shown in Figure 2,
146 whereby the BMU is thought to be activated by an initial stimulation of bone resorption, meaning that
147 any change in bone formation would be expected to lag behind that of bone resorption. Despite this,
148 increased PINP has been reported following 60 minutes of continuous running at intensities ranging
149 from 55 – 75% of VO_{2max} (39,44), or an unaccustomed football session (45), demonstrating that
150 indicators of bone formation do, sometimes, respond to acute exercise, although this response is less
151 common than that observed in markers indicative of bone resorption. The reason for this
152 inconsistency in response is not entirely clear and further research to better characterise the BMM
153 response to acute bouts of exercise under varying conditions and with longer follow-ups post exercise
154 bout will be of interest.

155 Exercise intensity and duration seem to be instrumental in determining the BMM response to acute
156 exercise (38), with higher, but not lower, intensity protocols typically eliciting a response. Those
157 studies that observed no response to an acute bout of exercise generally used lower intensity and/or
158 shorter duration protocols, including 30 minutes of walking or jogging (46,47), a 30 second Wingate
159 cycling test (48) or water aerobics (46). Bone is commonly thought to respond only to high-impact or
160 unusual impact loads, but the available studies show that these are not essential to elicit an acute
161 BMM response. For example, cycling tests consistently increase β -CTX-I (36,37,49), despite conveying
162 little to no impact loads. Recently, intensity-matched interval sessions conducted either on a bike or
163 treadmill induced comparable sclerostin responses in men (50) and women (51). This demonstrates
164 that impact was not necessary to elicit this BMR response, which instead must have been stimulated
165 by other, potentially metabolic, factors, such as increased reactive oxygen or nitrogen species (14),
166 acidosis (15) or reduced serum calcium availability (16).

167 Despite strong evidence that bone resorptive markers, such as β -CTX-I, are responsive to acute
168 exercise, some well-controlled investigations reported no change to any BMM, even though they used

169 high-intensity protocols (43). It is important to keep in mind, that the BMM response to exercise is
170 time-specific and transient (38,39,52). For example, some studies reported changes to BMM
171 concentrations as the area under the curve of multiple sampling points (53), but the response was not
172 apparent when based on single sampling points. Similarly, both a sustained (39) and a transient (44)
173 β -CTX-I response to treadmill running in the days following an acute exercise bout were reported, with
174 exercise intensity the apparent differentiator between these two responses. Thus, studies that use
175 single sampling points may well miss transient or time-specific changes, consequently impacting
176 mechanistic interpretation of findings. Future studies should seek to use multiple sampling points to
177 characterise the BRM responses to acute exercise, ideally taken over several days, post-exercise. The
178 nature of the temporal BRM response to exercise is, however, incompletely understood, rendering it
179 difficult to identify the most appropriate timing and number of sampling points. Additionally,
180 hemoconcentration should be assessed and accounted for in order to control for the potentially
181 confounding influence of plasma volume changes on the bone biomarker response to exercise.

182 ***2.3. The bone metabolic response to longitudinal exercise interventions***

183 Prolonged exposure to exercise training typically elicits an increase in resting levels of bone formation
184 markers (either PINP, bone alkaline phosphatase (BALP) or both) (54–67), indicating that training
185 might stimulate chronic upregulation of bone formative processes. This aligns with the model shown
186 in Figure 2, whereby increased bone resorption in response to acute exercise (described in Section
187 2.1) activates the BMU, ultimately leading to an increase in bone formation. Unlike the largely
188 consistent finding of increased bone formation in response to exercise interventions, markers of bone
189 resorption are less responsive to training, with most studies reporting no change. A few studies have
190 reported a reduction in bone resorption markers following a training program (55,57,68,69), and this
191 was typically accompanied by an increase in bone formation, suggesting a metabolic bone profile
192 favouring formation. Some studies also reported a concomitant increase in BMD alongside increases

193 in bone formation markers (59,68,70,71), indicating that training can be osteogenic, and that this can
194 be monitored by BMMs.

195 In common with the studies investigating the BMM response to acute bouts of exercise, efforts have
196 been made to identify how various exercise characteristics, including type, intensity and total work
197 done, influence their response to training. Studies that matched training volume, but varied intensity,
198 reported either a larger (59) or similar (72) response in bone formation markers (serum OC and B-ALP)
199 when a higher intensity protocol was employed. These inconsistent results suggest that, although
200 exercise intensity may well influence the BRM response to exercise, it is unlikely to be the
201 predominant factor. Instead the total amount of work done (to which intensity will certainly
202 contribute) may be a more important factor.

203 A wide range of exercise types have been investigated, but no one type stands out as being more or
204 less effective at eliciting a BMM response. It is generally accepted that high-intensity exercise that
205 conveys large and unaccustomed gravitational or muscular loads is necessary to elicit an osteogenic
206 response (73). It follows that this type of exercise would induce the largest and most consistent
207 increase in markers of bone formation following a period of training, but this does not seem to be the
208 case. Similar to evidence from acute studies (described in Section 2.1), exercise types with lower
209 impact and repetitive loading cycles (such as treadmill walking/running, step aerobics and yoga
210 (54,55,57,60,66)) were equally likely to elicit a response in bone formation markers, as those
211 modalities that exert large muscle or gravitational forces, (such as football training (63), high-impact
212 jump activities (61,68) resistance training (58,59,62,64) and multi-modal activities (67), including
213 military combat training (56)). This raises an important question about the relationship between BMM
214 and chronic bone outcomes, such as mass and strength. It is widely accepted that exercise type is an
215 important determinant of the bone response to exercise, but this view is not supported by the
216 available BMM data. Is it possible that exercise type is less important to bone than previously
217 believed? Or perhaps BMM changes are not necessarily predictive of changes to bone mass, strength

218 or microarchitecture? The available evidence does not allow this question to be answered, but in order
219 to optimise the use of BMMs in sport and exercise science, it should be addressed in future
220 investigations.

221 Individual participant characteristics, such as age, health and training status, are also important when
222 considering the bone metabolic response to exercise. Many of the investigations that reported no
223 response to exercise training were conducted on older adults (74–76), children with type 2 diabetes
224 (77) and breast cancer survivors (78). It seems plausible to suggest, therefore, that age, or health-
225 related factors, may have influenced these results. Indeed “anabolic resistance” has been reported
226 to be a consequence of senescence, and refers to a blunted response to anabolic stimuli, such as
227 exercise or protein (79,80). Similarly “osteogenic resistance” to bone loading programs in older adults
228 has been proposed (81), which may be due to various age-associated physiological changes, such as
229 reduced sex hormone content, although direct evidence to support or refute this hypothesis does not
230 currently exist. Having said that, exercise interventions were osteogenic in postmenopausal (2) and
231 older populations (82), which would necessitate an upregulation in bone remodelling, suggesting that
232 while age and hormonal changes may attenuate exercise-induced osteogenesis, they do not block it.

233 Most of the investigations described in this review indicate that exercise training triggers an increase
234 in bone formation activities, and so should be osteogenic. But circulating OC and B-ALP decreased
235 following a period of intensive training in two groups of military recruits (83,84), showing that training
236 can, in certain situations, suppress bone formation. This finding likely relates to the amount of energy
237 available to support bone remodelling (85) (described in Section 3.1). It is also plausible that
238 inadequate recovery during periods of particularly arduous and unaccustomed training may impede
239 the reversal phase of the bone remodelling cycle, thus attenuating its osteogenic potential. These
240 findings highlight the many factors, independent to the actual exercise itself, that may moderate the
241 bone metabolic response to exercise training. This complexity makes it difficult to isolate, and so to
242 investigate, any one individual factor. Recognition of this challenge is essential to the design and

243 interpretation of appropriately-controlled studies that are capable of really enhancing understanding
244 of this important research area.

245 **2.4. The bone metabolic profile of different athletic populations**

246 Cross-sectional studies of different athletic groups provide insight into the influence of habitual
247 training practices on bone metabolism. As expected, increased BMMs (both formation and
248 resorption), alongside increased BMD and/or enhanced geometry, have been reported in athletes
249 participating in sports involving high mechanical loading, including gymnasts (86), decathletes (87) and
250 football players (88). Altered bone metabolism was also reported in athletes involved in lower-impact
251 sports (*e.g.*, swimming (89), cycling (90,91) and horse-racing (92)), but these typically presented as
252 either decreased bone formation (89,91) or increased bone resorption (90,92), suggesting overall
253 bone loss. Low-impact sports such as these are considered to provide a sub-optimal stimulus to bone,
254 although it is not clear if this is due to the lower mechanotransductive signals provided by low-impact
255 and repetitive loading cycles, or whether other factors, such as an insufficient energy availability (EA;
256 described in Section 3.1), may also influence this response.

257 The finding of altered bone metabolism in athletic groups is not consistent across the literature; no
258 BMM differences were shown between controls and female athletes involved in high-impact sports
259 (93), rhythmic gymnasts (94) and male master runners and speed/power athletes (95). Adapted BMD
260 and/or bone microarchitecture was, however, reported in these studies, suggesting that bone was
261 impacted by sports participation. This might suggest that BMMs are not necessarily indicative of
262 altered bone mass or microarchitecture. On the other hand, many of the studies described herein
263 were based upon single sampling points and given the temporal BMM responses to exercise and
264 training it is possible that upregulated metabolism was not detected.

265 **3. THE INFLUENCE OF NUTRIENT INTAKE ON THE BONE METABOLIC RESPONSE TO EXERCISE**

266 When considering the BMM response to exercise, it is essential to also consider the nutritional
267 environment within which that response took place. Bone is acutely responsive to nutrient intake (96–
268 98) and studies investigating the impact of nutritional interventions on the bone metabolic response
269 to exercise can be used to identify the mechanistic pathways underpinning this response, and to
270 inform the development of nutritional interventions to improve bone health.

271

272 **3.1. Energy availability**

273 EA refers to the amount of energy available for physiological processes, after the demands of training
274 are met (99), and is an important determinant of bone health in athletes. Extensive research shows
275 that insufficient EA negatively impacts a variety of bone parameters (85), including BMMs. In a
276 parallel-group study, Ihle & Loucks. (2004) examined the dose-response relation between EA (10, 20,
277 30 or 45 kcal·kgLBM·day⁻¹) and bone metabolism in sedentary, but otherwise healthy, eumenorrheic
278 young women. Bone formation (OC and carboxyterminal propeptide of type 1 procollagen (PICP)) was
279 suppressed at all levels of low EA (30, 20 and 10 kcal·kgLBM·day⁻¹). This was accompanied by a
280 significant increase of bone resorption (aminoterminal telopeptide of type 1 collagen (NTX)) during
281 the lowest EA condition (10 kcal·kg LBM·day⁻¹ (100)). More recently, Papageougiou et al. (101)
282 conducted two, independent repeated-measure investigations (reported in the same paper), on the
283 influence of 5 days low EA (15 Kcal·kg LBM·day⁻¹) on bone metabolism in physically active men (study
284 1) and women (study 2). Low EA increased bone resorption (β -CTX-I) and reduced bone formation
285 (PINP) in women, but not in men (101). Each of the studies described herein induced low EA through
286 a combination of exercise and dietary restriction, and so could not distinguish whether restricted
287 energy intake, or increased energy expenditure, was the dominant cause of altered bone metabolism.
288 This important point was subsequently investigated, and it seems that low EA (15 kcal·kg LBM·day⁻¹)
289 induced through dietary restriction, but not by exercise-induced energy expenditure, reduces bone
290 formation (PINP) (102). It is not clear whether this occurred because the benefits of exercise masked,
291 or over-rode, the negative effects of restricted energy intake, and further mechanistic elucidation is

292 important. Irrespective of the mechanisms, however, it seems that exercise may protect bone during
293 periods of energy restriction, which has implications for interventions designed to protect bone during
294 such periods, suggesting that the focus should be on increasing dietary intake, rather than on reducing
295 exercise. The benefits of this strategy may extend beyond the bone alone, although the efficacy of this
296 approach for bone, and for other tissues should be confirmed using randomised controlled
297 investigations.

298 Reduced bone formation was also reported in cross-sectional studies conducted on energy deficient
299 athletes (103–106), and in clinical populations characterised by extreme energy deficiency (*e.g.*,
300 anorexia nervosa) (107–109). This likely occurs in an attempt to preserve energy for more immediately
301 essential physiological processes (99). Reduced bone formation was accompanied by reduced
302 resorption in energy and oestrogen-deficient exercising women (103), adolescent boys with anorexia
303 (108), fasted lightweight male rowers (104) and energy-deficient amenorrhic and oligomenorrhic
304 women (105). In contrast, extremely low EA simultaneously increased bone resorption, and decreased
305 formation, in severe restriction trials (10 kcal·kg LBM·day⁻¹, (100)) and in some studies of patients with
306 anorexia nervosa (107,109). Such a bone profile has particularly negative consequences for bone,
307 should it persist for prolonged periods of time. Evidence of disrupted bone metabolism in response to
308 low EA has implications for research in this area and likely accounts, at least in part, for findings
309 described earlier in this review, including reduced bone formation following periods of arduous
310 training (83,84,110) (Section 2.2) or as identified in cross-sectional investigations of some athletic
311 groups (89,90,92) (Section 2.3).

312 **3.2. Macronutrient ingestion pre, during and post exercise**

313 Nutrient ingestion pre, during and immediately post acute exercise can alter the BMM response to
314 that exercise bout. Scott et al. (39) investigated pre-exercise ingestion of a mixed meal, versus fasting,
315 on response to a 60-minute treadmill run conducted at 65% of VO_{2max}. Meal ingestion reduced pre-
316 exercise β-CTX-I, but did not influence its exercise induced increase, and the authors concluded that

317 pre-exercise feeding did not meaningfully impact the BMM response to the subsequent exercise bout.
318 However, this study suggested that the stress of the exercise bout over-rode the pre-exercise effect
319 of feeding on β -CTX-I, which, in turn, raised the question of whether or not more continuous nutrient
320 provision throughout the exercise bout would exert a more noticeable effect. This was investigated
321 by Sale et al. (53), who provided CHO before, during, and after a 120 minute treadmill run and reported
322 modestly reduced PINP and β -CTX-I post-exercise.

323 Studies investigating nutrient ingestion pre- and during exercise are limited by practical considerations
324 related to the type and volume of nutrients that can be ingested without impacting exercise
325 performance. The post-exercise period is thus more amenable to feeding interventions. Townsend et
326 al. (111) investigated the influence of a combined CHO/protein supplement following a fatiguing
327 treadmill run and reported a suppression of the β -CTX-I response when compared to the control trial,
328 along with a smaller, but statistically significant, increase in PINP concentrations at 3 and 4 hours post-
329 exercise (111).

330 These studies demonstrate that feeding around exercise can modulate the bone resorptive response
331 to that exercise bout, with the post-exercise period perhaps the most practical and influential
332 opportunity for nutrient provision. Theoretically, this reduction in bone resorption may protect bone
333 during periods of high intensity and/or volume training. On the other hand, and as described in Section
334 2, exercise-induced increases in bone resorption are necessary for BMU activation, and it is plausible
335 that attenuated bone resorption during or post-exercise, could, theoretically, blunt the bone adaptive
336 response. To date, longitudinal studies investigating how acute BMM alterations translate in the long-
337 term are not available, meaning that these potential long-term consequences are hypothetical and
338 require investigation.

339 The studies described above were not designed to investigate the independent influence of the three
340 macronutrients (carbohydrates, fats and proteins) on the BMM response to exercise, and limited data
341 on this topic exist. Protein is a particularly interesting macronutrient in this context, given it's

342 relevance to athletic adaptation and performance, along with the many, and potentially conflicting,
343 pathways through which it influences bone. The available evidence indicates that protein is a bone-
344 protective nutrient (112) and largely positive, albeit somewhat inconsistent, results have been
345 reported in studies investigating the influence of protein supplementation in conjunction with
346 exercise-training on bone metabolism in healthy men and women (113), overweight and obese
347 individuals (114,115) and postmenopausal women (116). No change (114), a trend toward increased
348 formation and resorption (113) and increased bone formation only (115,116) were reported.
349 However, the latter two studies provided a combined protein/calcium supplement (115) or
350 protein/CHO/calcium/Vit D (116) and so the influence of protein supplementation *per se* could not be
351 isolated. The independent and combined influence of protein, carbohydrates and fats on the bone
352 metabolic response to exercise represents another exciting avenue for on-going research.

353 **3.3. Micronutrient ingestion**

354 Many micronutrients influence bone (117), but only the impact of calcium and vitamin D ingestion in
355 conjunction with exercise has been investigated. Vigorous exercise increases PTH secretion, which in
356 turn activates bone resorption (36,37,118–121). This increase in PTH secretion may be induced, at
357 least in part, by a reduction of serum ionized calcium (iCa). Therefore, strategies to protect serum
358 calcium availability during exercise may influence the bone metabolic response to that exercise bout.
359 This hypothesis is supported by studies that showed suppressed PTH and β -CTX-I (37,122), or
360 suppressed PTH but no change to β -CTX-I (36) when a calcium supplement was provided during and/or
361 pre-exercise. Recently, Kohrt and colleagues (16) conducted an elegant study, investigating the
362 influence of serum iCa availability on the PTH and bone resorptive response to a 60 minute, vigorous
363 cycling protocol. A clamp was used to provide a variable iCa infusion throughout the exercise test, thus
364 preventing an exercise-induced decline in serum iCa. This maintenance of serum iCa availability
365 attenuated, but did not fully prevent, exercise-induced increases in PTH and β -CTX-I (16),
366 demonstrating that calcium disruption, at least partially, regulates the bone resorptive response to

367 exercise. The underlying causes of exercise induced calcium disruptions are not entirely clear. Dermal
368 calcium losses due to sweating may contribute, but these losses are small (apart from during very
369 prolonged and/or intense exercise perhaps), and are unlikely to largely impact either serum calcium
370 availability or the β -CTX response to exercise (123). Further research is certainly required to elucidate
371 the underlying mechanism, particularly given that calcium supplementation may be protective to
372 athletic bone health. In further support of this, reduced β -CTX-I levels, along with enhanced tibial bone
373 properties, were reported following 6 months of combined calcium and vitamin D supplementation in
374 a group of young horse-racing jockeys (124); of note, the study was not designed to investigate the
375 independent influence of calcium or vitamin D.

376 **4. FUTURE PERSPECTIVES**

377 It is clear that both acute and chronic exercise can induce a BMM response (summarised in Figure 2)
378 and these biomarkers have exciting potential to increase our understanding of the complex relation
379 between exercise and bone. Currently, important gaps in our understanding of the different factors
380 that regulate the bone response to exercise, and a lack of data on BMMs predictive ability exists. These
381 knowledge gaps should be filled to progress understanding, and thus practical application, of these
382 exciting biomarkers.

383 The scientific triad of standardisation, harmonisation and population specific reference ranges were
384 identified as vital steps toward the optimisation of BMMs in clinical practice (33,125), and the same is
385 true for their use within sport and exercise science and medicine. Elevated bone metabolism within
386 the clinical setting is indicative of increased fracture risk (34). But BMMs were unable to differentiate,
387 or to predict, stress fracture occurrence in athletes or military recruits (126–129). In order to move
388 toward the practical use of BMMs in sports medicine, validated, population specific, reference ranges
389 are essential. This will allow differentiation between those for whom altered BRM simply reflects the
390 demands of training, and those for whom changes may be pathological. The specific conditions
391 required to standardise and optimise selected bone biomarkers should be investigated in the design
392 and planning stages of all projects, to ensure that conditions are optimised and that valid information

393 is obtained. For example, β -CTX-I is known to be more significantly influenced by circadian rhythms
394 and nutritional intake than PINP, which is relatively stable in response to these factors (130). As such,
395 the control and standardisation approaches adopted for both may differ, depending on the primary
396 objective of the study. Harmonisation of future research efforts through including, at a minimum, the
397 reference markers of PINP and β -CTX-I, will allow for greater comparability of future findings, while
398 rigorous standardisation and control of research design and protocols will allow for a greater isolation
399 of moderating factors.

400 Most investigations on this topic have relied upon simple dichotomous interpretations of
401 increases/decreases in various BMMs as being either positive or negative. Some care must be taken
402 with this approach, as it does not recognise the complexity of these processes, and the context and
403 magnitude of change must be considered when interpreting BMM results. For example, strategies that
404 attenuate the bone resorptive response to acute exercise are generally considered to be positive, and
405 this may well be the case for highly-active individuals at risk of bone loss. But could these same
406 strategies also blunt subsequent anabolic adaptations? Our current understanding of the BMM
407 response to exercise is insufficient to answer this question. Pending a more complete understanding
408 of the physiological relevance of the BRM response to exercise, results should not be extrapolated
409 beyond the delimitations of the study, unless accompanied by appropriate clinical or functional
410 outcomes. The length and context of exposure to stimuli, and the temporal nature of BRM responses
411 to exercise is also very important. Transient exposure to various exercise-induced stimuli, including
412 reactive species, acidosis, or glucocorticoids, may well be essential for BMU activation and subsequent
413 remodelling and adaptation. Conversely, prolonged exposure to these same stimuli, as occurs in many
414 situations (*e.g.*, clinical conditions characterised by oxidative stress, low grade metabolic acidosis or
415 the sustained use of glucocorticoid therapies) are detrimental to bone.

416 We do not know how transient changes to individual BRMs translate in the long-term toward changes
417 to BMD and microarchitecture and, ultimately, to bone strength and fracture susceptibility. Where
418 possible, longitudinal studies should correlate changes in individual BRMs with these chronic

419 indicators in order to estimate their predictive ability. Careful consideration of these, and other factors
420 described in this review, may enhance the use of these biomarkers in ongoing investigations, thus
421 providing a platform upon which evidence-based practical and clinical recommendations may be
422 made to enhance the bone health of athletes, as well as those undergoing exercise-based therapeutic
423 interventions.

424 **References**

- 425 1. Lima F, De Falco V, Baima J, Carazzato J, Pereira R. Effect of impact load and active load on
426 bone metabolism and body composition of adolescent athletes. *Med Sci Sport Exerc.*
427 2001;33(8):1318–23.
- 428 2. Howe T, Shea B, Dawson L, Downie F, Murry A, Ross C, et al. Exercise for preventing and
429 treating osteoporosis in postmenopausal women. *Cochrane Database Syst Rev.*
430 2011;6(7):CD000333.
- 431 3. Olmedillas H, Gonzalez-Aquero A, Moreno L, Casajus J, Vicente-Rodríguez G. Cycling and bone
432 health: A systematic review. *BMC Med.* 2012;10:168.
- 433 4. Gómez-Bruton A, González-Agüero A, Gómez-Cabello A, Casajús JA, Vicente-Rodríguez G. Is
434 bone tissue really affected by swimming? A systematic review. *PLoS One.* 2013;8(8):e70119.
- 435 5. Ackerman K, Nazem T, Chapko D, Russell M, Mendes N, Taylor A, et al. Bone
436 microarchitecture is impaired in adolescent amenorrheic athletes compared with
437 eumenorrheic athletes and nonathletic controls. *J Clin Endocrinol Metab.* 2011;96(10):3123–
438 33.
- 439 6. Scofield K, Hecht S. Bone health in endurance athletes: runners, cyclists, and swimmers. *Curr*
440 *Sports Med Rep.* 2012;11(6):328–34.
- 441 7. Kanis JA, Borgstrom F, De Laet C, Johansson H, Johnell O, Jonsson B, et al. Assessment of

- 442 fracture risk. *Osteoporos Int.* 2005;16(6):581–9.
- 443 8. Sornay-Rendu E, Boutroy S, Duboeuf F, Chapurlat R. Bone microarchitecture assessed by HR-
444 pQCT as predictor of fracture risk in postmenopausal women: The OFELY study. *J Bone Miner
445 Res.* 2017;32(6):1243–51.
- 446 9. Nishiyama K, Shane E. Clinical imaging of bone microarchitecture with HR-pQCT. *Curr
447 Osteoporos Rep.* 2013;11(2):147–55.
- 448 10. Bass S, Eser P, Daly R. The effect of exercise and nutrition on the mechanostat. *J
449 Musculoskelet Neuronal Interact.* 2005;5(3):239–54.
- 450 11. Frost H. A 2003 update of bone physiology and Wolff’s Law for clinicians. *Angle Orthod.*
451 2004;74(1):3–15.
- 452 12. Robling AG, Castillo AB, Turner CH. Biomechanical and molecular regulation of bone
453 remodeling. *Annu Rev Biomed Eng.* 2006;8(1):455–98.
- 454 13. Orr A, Helmke B, Blackman B, Schwartz M. Mechanisms of mechanotransduction. *Dev Cell.*
455 2006;10(1):11–20.
- 456 14. Ha H, Bok Kwak H, Woong Lee S, Mi Jin H, Kim HM, Kim HH, et al. Reactive oxygen species
457 mediate RANK signaling in osteoclasts. *Exp Cell Res.* 2004;301(2):119–27.
- 458 15. Krieger NS, Frick KK, Bushinsky DA. Mechanism of acid-induced bone resorption. *Curr Opin
459 Nephrol Hypertens.* 2004;13(4):423–36.
- 460 16. Kohrt W, Wherry S, Wolfe P, Sherk D, Wellington T, Swanson C, et al. Maintenance of serum
461 ionized calcium during exercise attenuates parathyroid hormone and bone resorption
462 responses. *J Bone Miner Res.* 2018;33(7):1326–34.
- 463 17. Hadjidakis D, Androulakis I. Bone remodeling. *Ann N Y Acad Sci.* 2006;1092:385–96.

- 464 18. Ivaska K, Hentunen T, Vaaraniemi J, Ylipahkala H, Pettersoon K, Vaananen H. Release of intact
465 and fragmented osteocalcin molecules from bone matrix during bone resorption in vitro. *J*
466 *Biol Chem.* 2004;279(18):18361–9.
- 467 19. Lombardi G, Perego S, Luzi L, Banfi G. A four-season molecule: osteocalcin. Updates in its
468 physiological roles. *Endocrine.* 2015;48(2):394–404.
- 469 20. Hegedus D, Ferencz V, Lakatos P, Meszaros S, Lakatos P, Horvath C, et al. Decreased bone
470 density, elevated serum osteoprotegerin, and beta-cross-laps in Wilson disease. *J Bone Miner*
471 *Res.* 2002;17(11):1961–7.
- 472 21. Kannus P, Haapsasalo H, Sievanen H, Oja P, Vuori I. The site-specific effects of long-term
473 unilateral activity on bone mineral density and content. *Bone.* 1994;15(3):279–84.
- 474 22. Wishart J, Need A, Horowitz M, Morris H, Nordin B. Effect of age on bone density and bone
475 turnover in men. *Clin Endocrinol (Oxf).* 1995;42(2):141–6.
- 476 23. Gass M, Kagan R, Kohles J, Martens M. Bone turnover marker profile in relation to the
477 menstrual cycle of premenopausal healthy women. *Menopause.* 2008;15(4):667–75.
- 478 24. Garnero P, Sornay-Rendu E, Delmas P. Decreased bone turnover in oral contraceptive users.
479 *Bone.* 1995;16(5):499–503.
- 480 25. Woitge H, Scheidt-Nave C, Kissling C, Leidig-Bruckner G, Meyer K, Grauer A, et al. Seasonal
481 variation of biochemical indexes of bone turnover: results of a population-based study. *J Clin*
482 *Endocrinol Metab.* 1998;83(1):68–75.
- 483 26. Qvist P, Christgau S, Pedersen B, Schlemmer A, Christiansen C. Circadian variation in the
484 serum concentration of C-terminal telopeptide of type I collagen (serum CTx): effects of
485 gender, age, menopausal status, posture, daylight, serum cortisol, and fasting. *Bone.*
486 2002;31(1):57–61.

- 487 27. Kemp J, Sayers A, Paternoster L, Evans D, Deere K, St Pourcain B, et al. Does bone resorption
488 stimulate periosteal expansion? A cross sectional analysis of β -C-telopeptides of type I
489 collagen (CTX), genetic markers of the RANKL pathway, and periosteal circumference as
490 measured by pQCT. *J Bone Miner Res.* 2014;29(4):1015–24.
- 491 28. Hlaing TT, Compston JE. Biochemical markers of bone turnover - uses and limitations. *Ann*
492 *Clin Biochem.* 2014;51(2):189–202.
- 493 29. Veitch S, Findlay S, Hamer A, Blumsohn A, Eastell R, Ingle B. Changes in bone mass and bone
494 turnover following tibial shaft fracture. *Osteoporos Int.* 2006;17(3):364–72.
- 495 30. Ingle B, Hay S, Bottier H, Eastell R. Changes in bone mass and bone turnover following distal
496 forearm fracture. *Osteoporos Int.* 1999;10(5):399–407.
- 497 31. Christensen G, Halgreen J, Milenkovski M, Kose A, Quardon N, Jorgensen N. Bone turnover
498 markers are differentially affected by pre-analytical handling. *Osteoporos Int.* 2019;
- 499 32. Vasikaran S, Eastell R, Bruyère O, Foldes AJ, Garnero P, Griesmacher A, et al. Markers of bone
500 turnover for the prediction of fracture risk and monitoring of osteoporosis treatment: A need
501 for international reference standards. *Osteoporos Int.* 2011;22(2):391–420.
- 502 33. Bauer D, Krege J, Lane N, Leary E, Libanati C, Miller P, et al. National Bone Health Alliance
503 Bone Turnover Marker Project: Current practices and the need for US harmonization,
504 standardization, and common reference ranges. *Osteoporos Int.* 2012;23(10):2425–33.
- 505 34. Vasikaran SD, Paul Chubb SA. The use of biochemical markers of bone turnover in the clinical
506 management of primary and secondary osteoporosis. *Endocrine.* 2016;52(2):222–5.
- 507 35. Chubb SAP, Byrnes E, Manning L, Golledge J, Ebeling PR, Flicker L, et al. Bone turnover
508 markers: Defining a therapeutic target. *Clin Biochem.* 2017;50(3):162–3.
- 509 36. Barry DW, Hansen KC, Van Pelt RE, Witten M, Wolfe P, Kohrt WM. Acute calcium ingestion

- 510 attenuates exercise-induced disruption of calcium homeostasis. *Med Sci Sports Exerc.*
511 2011;43(4):617–23.
- 512 37. Guillemant J, Accarie C, Peres G, Guillemant S. Acute effects of an oral calcium load on
513 markers of bone metabolism during endurance cycling exercise in male athletes. *Calcif Tissue*
514 *Int.* 2004;74(5):407–14.
- 515 38. Maïmoun L, Manetta J, Couret I, Dupuy AM, Mariano-Goulart D, Micallef JP, et al. The
516 intensity level of physical exercise and the bone metabolism response. *Int J Sports Med.*
517 2006;27(2):105–11.
- 518 39. Scott JPR, Sale C, Greeves JP, Casey A, Dutton J, Fraser WD. Effect of fasting versus feeding on
519 the bone metabolic response to running. *Bone.* 2012;51(6):990–9.
- 520 40. Rantalainen T, Heinonen A, Linnamo V, Komi P, Takala T, Kainulainen H. Short-term bone
521 biochemical response to a single bout of high-impact exercise. *J Sport Sci Med.*
522 2009;8(4):553–9.
- 523 41. Pomerants T, Tillmann V, Karelson K, Jurimae J, Jurimae T. Impact of acute exercise on bone
524 turnover and growth hormone/insulin-like growth factor axis in boys. *J Sports Med Phys*
525 *Fitness.* 2008;48(2):266–71.
- 526 42. Scott JPR, Sale C, Greeves JP, Casey A, Dutton J, Fraser WD. The effect of training status on
527 the metabolic response of bone to an acute bout of exhaustive treadmill running. *J Clin*
528 *Endocrinol Metab.* 2010;95(8):3918–25.
- 529 43. Scott JPR, Sale C, Greeves JP, Casey A, Dutton J, Fraser WD. Effect of recovery duration
530 between two bouts of running on bone metabolism. *Med Sci Sports Exerc.* 2013;45(3):429–
531 38.
- 532 44. Scott JPR, Sale C, Greeves JP, Casey A, Dutton J, Fraser WD, et al. The role of exercise intensity

- 533 in the bone metabolic response to an acute bout of weight-bearing exercise. 2011;(52):423–
534 32.
- 535 45. Bowtell JL, Jackman SR, Scott S, Connolly LJ, Mohr M, Ermidis G, et al. Short duration small
536 sided football and to a lesser extent whole body vibration exercise induce acute changes in
537 markers of bone turnover. *Biomed Res Int*. 2016;2016.
- 538 46. Morgan AL, Weiss J, Kelley ET. Bone turnover response to acute exercise with varying impact
539 levels: A preliminary investigation. *Int J Exerc Sci*. 2015;8(2):154–63.
- 540 47. Tosun A, Bölükbaşı N, Çıngı E, Beyazova M, Ünlü M. Acute effects of a single session of
541 aerobic exercise with or without weight-lifting on bone turnover in healthy young women.
542 *Mod Rheumatol*. 2006;16(5):300–4.
- 543 48. Kristoffersson A, Hultdin J, Holmlund I, Thorsen K, Lorentzon R. Effects of short-term maximal
544 work on plasma calcium, parathyroid hormone, osteocalcin and biochemical markers of
545 collagen metabolism. *Int J Sports Med*. 1995;16(3):145-9.
- 546 49. Herrmann M, Muller M, Scharhag J, Sand-Hill M, Kindermann W, Herrmann W. The effect of
547 endurance exercise-induced lactacidosis on biochemical markers of bone turnover. *Clin Chem
548 Lab Med*. 2007;45(10):1381–9.
- 549 50. Kouvelioti R, LeBlanc P, Falk B, Ward W, Josse A, Klentrou P. Effects of high-intensity interval
550 running versus cycling on sclerostin, and markers of bone turnover and oxidative stress in
551 young men. *Calcif Tissue Int*. 2019;1(1):1–9.
- 552 51. Kouvelioti R, Kurgan N, Falk B, Ward W, Josse A, Klentrou P. Response of sclerostin and bone
553 turnover markers to high intensity interval exercise in young women: Does impact matter?
554 *Biomed Res Int*. 2018;4864952:1–8.
- 555 52. Hinton PS, Rolleston A, Rehrer NJ, Hellemans IJ, Miller BF. Bone formation is increased to a

- 556 greater extent than bone resorption during a cycling stage race. *Appl Physiol Nutr Metab.*
557 2010;35(3):344–9.
- 558 53. Sale C, Varley I, Jones T, James R, Tang J, Fraser W, et al. Effect of carbohydrate feeding on
559 the bone metabolic response to running. *J Appl Physiol.* 2015;119(7):824–30.
- 560 54. Adami S, Gatti D, Viapiana O, Fiore CE, Nuti R, Luisetto G, et al. Physical activity and bone
561 turnover markers: A cross-sectional and a longitudinal study. *Calcif Tissue Int.*
562 2008;83(6):388–92.
- 563 55. Alp A. Effects of aerobic exercise on bone-specific alkaline phosphatase and urinary CTX levels
564 in premenopausal women. *Turkish J Phys Med Rehabil.* 2013;59(4):310–3.
- 565 56. Lutz LJ, Karl JP, Rood JC, Cable SJ, Williams KW, Young AJ, et al. Vitamin D status, dietary
566 intake, and bone turnover in female Soldiers during military training. *J Int Soc Sports Nutr.*
567 2012;9:1–7.
- 568 57. Roghani T, Torkaman G, Movassegh S, Hedayati M, Goosheh B, Bayat N. Effects of short-
569 term aerobic exercise with and without external loading on bone metabolism and balance in
570 postmenopausal women with osteoporosis. *Rheumatol Int.* 2013;33(2):291–8.
- 571 58. Tajima O, Ashizawa N, Ishii T, Amagai H, Mashimo T, Liu LJ, et al. Interaction of the effects
572 between vitamin D receptor polymorphism and exercise training on bone metabolism. *J Appl*
573 *Physiol.* 2000;88(8750–7587):1271–6.
- 574 59. Vincent KR, Braith RW. Resistance exercise and bone turnover in elderly men and women.
575 *Med Sci Sports Exerc.* 2002;34(1):17–23.
- 576 60. Ardawi M-SM, Rouzi AA, Qari MH. Physical activity in relation to serum sclerostin, insulin-like
577 growth factor-1, and bone turnover markers in healthy premenopausal women: A cross-
578 sectional and a longitudinal study. *J Clin Endocrinol Metab.* 2012;97(10):3691–9.

- 579 61. Erickson CR, Vukovich MD. Osteogenic index and changes in bone markers during a jump
580 training program: A pilot study. *Med Sci Sports Exerc.* 2010;42(8):1485–92.
- 581 62. Fujimura R, Ashizawa N, Watanabe M, Mukai N, Amagai H, Fukubayashi T, et al. Effect of
582 resistance exercise training on bone formation and resorption in young male subjects
583 assessed by biomarkers of bone metabolism. *J Bone Miner Res.* 1997;12(4):656–62.
- 584 63. Helge EW, Andersen TR, Schmidt JF, Jørgensen NR, Hornstrup T, Krstrup P, et al.
585 Recreational football improves bone mineral density and bone turnover marker profile in
586 elderly men. *Scand J Med Sci Sport.* 2014;24(SUPPL.1):98–104.
- 587 64. Hu M, Finni T, Xu L, Zou L, Cheng S. Effects of resistance training on biomarkers of bone
588 formation and association with red blood cell variables. *J Physiol Biochem.* 2011;67(3):351–8.
- 589 65. Kim S, Sherk VD, Bembem MG, Debra A. Effects of short term low intensity resistance training
590 with blood flow restriction on bone markers and muscle cross- sectional area in young men.
591 *Int J Exerc Sci.* 2012;5(27):136–47.
- 592 66. Kim SJ, Bembem MG, Knehans AW, Bembem DA. Effects of an 8-month ashtanga-based yoga
593 intervention on bone metabolism in middle-aged premenopausal women: A randomized
594 controlled study. *J Sport Sci Med.* 2015;14(4):756–68.
- 595 67. Lester ME, Urso ML, Evans RK, Pierce JR, Spiering BA, Maresh CM, et al. Influence of exercise
596 mode and osteogenic index on bone biomarker responses during short-term physical
597 training. *Bone.* 2009;45(4):768–76.
- 598 68. Basat H, Esmailzadeh S, Eskiyurt N. The effects of strengthening and high-impact exercises
599 on bone metabolism and quality of life in postmenopausal women: A randomized controlled
600 trial. *J Back Musculoskelet Rehabil.* 2013;26(4):427–35.
- 601 69. Bezerra L, Bottaro M, Machado Reis V, Abdhala L, Lima R, Soares S, et al. Effects of yoga on

- 602 bone metabolism in postmenopausal women. *J Exerc Physiol online*. 2010;13(4):58–65.
- 603 70. Menkes A, Mazel S, Redmond R, Koffler K, Libanati C, Gundberg C, et al. Strength training
604 increases regional bone mineral density and bone remodeling in middle-aged and older men.
605 *J Appl Physiol*. 1993;74(5):2478–84.
- 606 71. Schroeder ET, Hawkins SA, Jaque SV. Musculoskeletal adaptations to 16 weeks of eccentric
607 progressive resistance training in young women. *J Strength Cond Res*. 2004;18(2):227–35.
- 608 72. Bemben D, Fethers N, Bemben M, Nabavi N, Koh T. Musculoskeletal responses to high- and
609 low-intensity resistance training in early postmenopausal women. *Med Sci Sport Exerc*.
610 2000;32(11):1949–57.
- 611 73. Kohrt WM, Barry DW, Schwartz RS. Muscle forces or gravity: What predominates mechanical
612 loading on bone? *Med Sci Sport Exerc*. 2009;41(11):2050–5.
- 613 74. Pruitt LA, Taaffe DR, Marcus R. Effects of a one-year high-intensity versus low-intensity
614 resistance training program on bone mineral density in older women. *J Bone Miner Res*.
615 1995;10(11):1788–95.
- 616 75. Rantalainen T, Hoffrén M, Linnamo V, Heinonen A, Komi P V., Avela J, et al. Three-month
617 bilateral hopping intervention is ineffective in initiating bone biomarker response in healthy
618 elderly men. *Eur J Appl Physiol*. 2011;111(9):2155–62.
- 619 76. Ryan A, Treuth M, Rubin M, Miller J, Nicklas B, Landis D, et al. Effects of strength training on
620 bone mineral density: Hormonal and bone turnover relationships. *J Appl Physiol*.
621 1994;77(4):1678–84.
- 622 77. Maggio A, Rizzoli R, Marchand L, Ferrari S, Beghetti M, Farpour-Lambert N. Physical activity
623 increases bone mineral density in children with Type 1 diabetes. *Med Sci Sport Exerc*.
624 2012;44(7):1206–11.

- 625 78. Simonavice E, Liu P-Y, Ilich JZ, Kim J-S, Arjmandi B, Panton LB. The effects of a 6-month
626 resistance training and dried plum consumption intervention on strength, body composition,
627 blood markers of bone turnover, and inflammation in breast cancer survivors 1. *Appl Physiol*
628 *Nutr Metab.* 2014;39(6):730–9.
- 629 79. Deutz N, Bauer J, Barazzoni R, Biolo G, Boirie Y, Bosy-Westphal A, et al. Protein intake and
630 exercise for optimal muscle function with aging: recommendations from the ESPEN Expert
631 Group. *Clin Nutr.* 2014;33(6):929–36.
- 632 80. Morton R, Traylor D, Weijs P, Phillips S. Defining anabolic resistance: implications for delivery
633 of clinical care nutrition. *Curr Opin Crit Care.* 2018;24(2):124–30.
- 634 81. Santos L, Elliott-Sale KJ, Sale C. Exercise and bone health across the lifespan. *Biogerontology.*
635 2017;18(6):931–46.
- 636 82. Gómez-Cabello A, Ara I, González-Agüero A, Casajús J, Vicente-Rodríguez G. Effects of training
637 on bone mass in older adults: A systematic review. *Sport Med.* 2012;42(4):301–25.
- 638 83. Etherington J, Keeling J, Bramley R, Swaminathan R, McCurdie I, Spector TD. The effects of 10
639 weeks military training on heel ultrasound and bone turnover. *Calcif Tissue Int.*
640 1999;64(5):389–93.
- 641 84. Hughes JM, Smith MA, Henning PC, Scofield DE, Spiering BA, Staab JS, et al. Bone formation is
642 suppressed with multi-stressor military training. *Eur J Appl Physiol.* 2014;114(11):2251–9.
- 643 85. Papageorgiou M, Dolan E, Elliott KJ, Craig S. Reduced energy availability: Implications for
644 bone health in physically active populations. *Eur J Nutr.* 2018;57(3):847–59.
- 645 86. Courteix D, Rieth N, Thomas T, Van Praagh E, Benhamou CL, Collomp K, et al. Preserved bone
646 health in adolescent elite rhythmic gymnasts despite hypoleptinemia. *Horm Res.*
647 2007;68(1):20–7.

- 648 87. Maïmoun L, Coste O, Puech AM, Peruchon E, Jaussent A, Paris F, et al. No negative impact of
649 reduced leptin secretion on bone metabolism in male decathletes. *Eur J Appl Physiol.*
650 2008;102(3):343–51.
- 651 88. Karlsson KM, Karlsson C, Ahlborg HG, Valdimarsson Ö, Ljunghall S. The duration of exercise as
652 a regulator of bone turnover. *Calcif Tissue Int.* 2003;73(4):350–5.
- 653 89. Creighton DL, Morgan AL, Boardley D, Brolinson PG. Weight-bearing exercise and markers of
654 bone turnover in female athletes. *J Appl Physiol.* 2001;90(2):565–70.
- 655 90. McVeigh JA, Meiring R, Cimato A, Micklesfield LK, Oosthuysen T. Radial bone size and strength
656 indices in male road cyclists, mountain bikers and controls. *Eur J Sport Sci.* 2015;15(4):332–
657 40.
- 658 91. Maïmoun L, Mariano-Goulart D, Couret I, Manetta J, Peruchon E, Micallef JP, et al. Effects of
659 physical activities that induce moderate external loading on bone metabolism in male
660 athletes. *J Sports Sci.* 2004;22(9):875–83.
- 661 92. Dolan E, McGoldrick A, Davenport C, Kelleher G, Byrne B, Tormey W, et al. An altered
662 hormonal profile and elevated rate of bone loss are associated with low bone mass in
663 professional horse-racing jockeys. *J Bone Miner Metab.* 2012;30(5):534–42.
- 664 93. Maïmoun L, Coste O, Philibert P, Briot K, Mura T, Galtier F, et al. Peripubertal female athletes
665 in high-impact sports show improved bone mass acquisition and bone geometry.
666 *Metabolism.* 2013;62(8):1088–98.
- 667 94. Tournis S, Michopoulou E, Fatouros IG, Paspatis I, Michalopoulou M, Raptou P, et al. Effect of
668 rhythmic gymnastics on volumetric bone mineral density and bone geometry in
669 premenarcheal female athletes and controls. *J Clin Endocrinol Metab.* 2010;95(6):2755–62.
- 670 95. Nowak A, Straburzyńska-Lupa A, Kusy K, Zieliski J, Felsenberg D, Rittweger J, et al. Bone

- 671 mineral density and bone turnover in male masters athletes aged 40-64. *Aging Male*.
672 2010;13(2):133–41.
- 673 96. Clowes JA, Hannon RA, Yap TS, Hoyle NR, Blumsohn A, Eastell R. Effect of feeding on bone
674 turnover markers and its impact on biological variability of measurements. *Bone*.
675 2002;30(6):886–90.
- 676 97. Babraj JA, Smith K, Cuthbertson DJR, Rickhuss P, Dorling JS, Rennie MJ. Human bone collagen
677 synthesis is a rapid, nutritionally modulated process. *J Bone Miner Res*. 2005;20(6):930–7.
- 678 98. Bjarnason N., Henriksen EE., Alexandersen P, Christgau S, Henriksen D., Christiansen C.
679 Mechanism of circadian variation in bone resorption. *Bone*. 2002;30(1):307–13.
- 680 99. Mountjoy M, Burke L, Ackerman KE, Blauwet C, Lebrun C, Melin A, et al. International
681 Olympic Committee (IOC) consensus statement on relative energy deficiency in sport (RED-S):
682 2018 Update. *Int J Sport Nutr*. 2018;28(4):316–31.
- 683 100. Ihle R, Loucks AB. Dose-response relationships between energy availability and bone turnover
684 in young exercising women. *J Bone Miner Res*. 2004;19(8):1231–40.
- 685 101. Papageorgiou M, Elliott-Sale KJ, Parsons A, Tang JCY, Greeves JP, Fraser WD, et al. Effects of
686 reduced energy availability on bone metabolism in women and men. *Bone*. 2017;105:191–9.
- 687 102. Papageorgiou M, Martin D, Colgan H, Cooper S, Greeves J, Tang J, et al. Bone metabolic
688 responses to low energy availability achieved by diet or exercise in active eumenorrheic
689 women. *Bone*. 2018;114:181–8.
- 690 103. De Souza MJ, West SL, Jamal SA, Hawker GA, Gundberg CM, Williams NI. The presence of
691 both an energy deficiency and estrogen deficiency exacerbate alterations of bone
692 metabolism in exercising women. *Bone*. 2008;43(1):140–8.
- 693 104. Talbott S, SHapses S. Fasting and energy intake influence bone turnover in lightweight male

- 694 rowers. *Int J Sport Nutr Exerc Metab.* 1998;8(4):377–87.
- 695 105. Zanker CL, Swaine IL. Relation between bone turnover, oestradiol, and energy balance in
696 women distance runners. *Br J Sports Med.* 1998;32(2):167–71.
- 697 106. Zanker CL, Swaine IL. Responses of bone turnover markers to repeated endurance running in
698 humans under conditions of energy balance or energy restriction. *Eur J Appl Physiol.*
699 2000;83(4–5):434–40.
- 700 107. Bolton JGF, Patel S, Lacey JH, White S. A prospective study of changes in bone turnover and
701 bone density associated with regaining weight in women with anorexia nervosa. *Osteoporos*
702 *Int.* 2005;16(12):1955–62.
- 703 108. Misra M, Katzman DK, Cord J, Manning SJ, Mendes N, Herzog DB, et al. Bone metabolism in
704 adolescent boys with anorexia nervosa. *J Clin Endocrinol Metab.* 2008;93(8):3029–36.
- 705 109. Soyka LA, Grinspoon S, Levitsky LL, Herzog DB, Klibanski A. The effects of anorexia nervosa on
706 bone metabolism in female adolescents. *J Clin Endocrinol Metab.* 1999;84(12):4489–96.
- 707 110. Maïmoun L, Galy O, Manetta J, Coste O, Peruchon E, Micallef JP, et al. Competitive season of
708 triathlon does not alter bone metabolism and bone mineral status in male triathletes. *Int J*
709 *Sports Med.* 2004;25(3):230–4.
- 710 111. Townsend R, Elliott-Sale KJ, Currell K, Tang J, Fraser WD, Sale C. The effect of postexercise
711 carbohydrate and protein ingestion on bone metabolism. *Med Sci Sports Exerc.*
712 2017;49(6):1209–18.
- 713 112. Dolan E, Sale C. Protein and bone health across the lifespan. *Proc Nutr Soc.* 2019;78(1):45–55.
- 714 113. Ballard TLP, Clapper JA, Specker BL, Binkley TL, Vukovich MD. Effect of protein
715 supplementation during a 6-mo strength and conditioning program on insulin-like growth
716 factor I and markers of. 2005;(1).

- 717 114. Cao JJ, Pasiakos SM, Margolis LM, Sauter ER, Whigham LD, McClung JP, et al. Calcium
718 homeostasis and bone metabolic responses to high-protein diets during energy deficit in
719 healthy young adults. *Am J Clin Nutr.* 2014;99(2):400–7.
- 720 115. Josse AR, Atkinson SA, Tarnopolsky MA, Phillips SM. Diets higher in dairy foods and dietary
721 protein support bone health during diet- and exercise-induced weight loss in overweight and
722 obese premenopausal women. *J Clin Endocrinol Metab.* 2012;97(1):251–60.
- 723 116. Holm L, Olesen JL, Matsumoto K, Doi T, Mizuno M, Alsted TJ, et al. Protein-containing
724 nutrient supplementation following strength training enhances the effect on muscle mass,
725 strength, and bone formation in postmenopausal women. *J Appl Physiol.* 2008;105(1):274–
726 81.
- 727 117. Palacios C. The role of nutrients in bone health, from A to Z. *Crit Rev Food Sci Nutr.*
728 2006;46(8):621-8.
- 729 118. Barry D, Kohrt W. Acute effects of 2 hours of moderate-intensity cycling on serum
730 parathyroid hormone and calcium. *Calcif Tissue Int.* 2007;80(6):359–65.
- 731 119. Shea K, Barry D, Sherk V, Hansen K, Wolfe P, Kohrt W. Calcium supplementation and
732 parathyroid hormone response to vigorous walking in postmenopausal women. *Med Sci*
733 *Sport Exerc.* 2014;46(10):2007–13.
- 734 120. Sherk V, Wherry S, Barry D, Shea K, Wolfe P, Kohrt W. Calcium supplementation attenuates
735 disruptions in calcium homeostasis during exercise. *Med Sci Sport Exerc.* 2017;49(7):1437–42.
- 736 121. Townsend R, Elliott-Sale K, Pinto A, Thomas C, Scott J, Currell K, et al. Parathyroid hormone
737 secretion is controlled by both ionized calcium and phosphate during exercise and recovery in
738 men. *J Clin Endocrinol Metab.* 2016;101(8):3231–9.
- 739 122. Haakonssen E, Ross M, Knight E, Cato L, Nana A, Wluka A, et al. The effects of a calcium-rich

- 740 pre-exercise meal on biomarkers of calcium homeostasis in competitive female cyclists: a
741 randomised crossover trial. *PLoS One*. 2015;10(5):e0123302.
- 742 123. Wherry S, Swanson C, Wolfe P, Wellington T, Boxer R, Schwartz R, et al. Bone biomarker
743 response to walking under different thermal conditions in older adults. *Med Sci Sport Exerc*.
744 2019;51(8):1599–605.
- 745 124. Silk LN, Greene DA, Baker MK, Jander CB. Tibial bone responses to 6-month calcium and
746 vitamin D supplementation in young male jockeys: A randomised controlled trial. *Bone*.
747 2015;81:554–61.
- 748 125. Morris HA, Eastell R, Jorgensen NR, Cavalier E, Vasikaran S, Chubb SAP, et al. Clinical
749 usefulness of bone turnover marker concentrations in osteoporosis. *Clin Chim Acta*.
750 2017;467:34–41.
- 751 126. Bennell KL, Malcolm SA, Brukner PD, Green RM, Hopper JL, Wark JD, et al. A 12-month
752 prospective study of the relationship between stress fractures and bone turnover in athletes.
753 *Calcif Tissue Int*. 1998;63(1):80–5.
- 754 127. Välimäki VV, Alfthan H, Lehmuskallio E, Löyttyniemi E, Sahi T, Suominen H, et al. Risk factors
755 for clinical stress fractures in male military recruits: A prospective cohort study. *Bone*.
756 2005;37(2):267–73.
- 757 128. Wakamatsu K, Sakuraba K, Suzuki Y, Maruyama A, Tsuchiya Y, Shikakura J, et al. Association
758 between the stress fracture and bone metabolism/quality markers in lacrosse players. *Open*
759 *Access J Sport Med*. 2012;3:67–71.
- 760 129. Yanovich R, Evans RK, Friedman E, Moran DS. Bone turnover markers do not predict stress
761 fracture in elite combat recruits basic research. *Clin Orthop Relat Res*. 2013;471(4):1365–72.
- 762 130. Szulc P, Naylor K, Hoyle N, Eastell R, Leary E. Use of CTX-I and PINP as bone turnover markers:

763 National Bone Health Alliance recommendations to standardize sample handling and patient
764 preparation to reduce pre analytical variability. Osteoporos Int. 2017;28:2541–56.

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768 **FIGURES:**

769 **Figure 1:** The bone remodelling cycle

770 **Figure 2:** Bone remodelling in response to exercise

771 Legend: **Panel A** describes how the mechanical and metabolic signals generated by an acute bout of
772 exercise activate the basic multicellular unit (BMU), thus mainly upregulating osteoclast activity,
773 represented by an increased blood biomarkers of resorptive activity, *e.g.*, β -CTX-1 (section 2.1).
774 Through the process of reversal, this increased bone resorptive activity induces a coupled elevation in
775 osteoblast activity, as is evident by increased resting blood bone formation markers following a period
776 of exercise training (**Panel B**; section 2.2).

777