

Food allergy across the globe



Vanitha Sampath, PhD,^{a,b} Elissa M. Abrams, MD, MPH,^{c,d} Bahman Adlou, MS,^{a,b} Cezmi Akdis, MD,^e Mübeccel Akdis, MD, PhD,^e Helen A. Brough, PhD, MBBS, FRCPCH,^{f,g} Susan Chan, MBBS, MD,^{f,g} Pantipa Chatchatee, MD,^h R. Sharon Chinthrajah, MD,^{a,b} Renata Rodrigues Cocco, MD, PhD,ⁱ Antoine Deschildre, MD,^j Philippe Eigenmann, MD,^k Cesar Galvan, MD,^{l,m} Ruchi Gupta, MD, MPH,^{n,o} Elham Hossny, MD, PhD,^p Jennifer J. Koplin, PhD,^{q,r} Gideon Lack, MBBS, FRCPCH, MD,^{f,g} Michael Levin, MBBS,^{s,t} Lynette P. Shek, MBBS, FRCPCH,^u Mika Makela, MD, PhD,^v David Mendoza-Hernandez, MD,^w Antonella Muraro, MD, PhD,^x Nikolaos G. Papadopoulos, MD, PhD, FRCP,^{y,z} Ruby Pawankar, MD, PhD,^{aa} Kirsten P. Perrett, MBBS, FRACP, PhD,^{q,r} Graham Roberts, DM, FRCPCH,^{bb,cc,dd} Cansin Sackesen, MD,^{ee} Hugh Sampson, MD,^{ff} Mimi L. K. Tang, MBBS, PhD, FRACP, FRCPA,^{q,r} Alkis Togias, MD,^{gg} Carina Venter, PhD, RD,^{hh} Christopher Michael Warren, PhD,^{n,o} Lisa M. Wheatley, MD, MPH,^{gg} Gary W. K. Wong, MD, FRCP,ⁱⁱ Kirsten Beyer, MD,^{jj*} Kari C. Nadeau, MD, PhD,^{a,b*} and Harald Renz, MD, PhD^{kk,ll*}

Stanford, Calif; Winnipeg, Manitoba, and Vancouver, British Columbia, Canada; Davos and Geneva, Switzerland; London, Manchester, Southampton, and Isle of Wight, United Kingdom; Bangkok, Thailand; São Paulo, Brazil; Lille, France; Lima, Peru; Chicago, Ill; Cairo, Egypt; Melbourne, Australia; Cape Town, South Africa; Singapore, Singapore; Helsinki, Finland; Mexico City, Mexico; Padua, Italy; Athens, Greece; Tokyo, Japan; Istanbul, Turkey; New York, NY; Bethesda, Md; Boulder, Colo; Hong Kong, China; Berlin and Marburg, Germany; and Moscow, Russia

From ^aSean N. Parker Center for Allergy and Asthma Research at Stanford University and ^bthe Division of Pulmonary and Critical Care Medicine, Department of Medicine, Stanford University, Stanford; ^cthe Department of Paediatrics, Section of Allergy and Clinical Immunology, University of Manitoba, Winnipeg; ^dthe Department of Paediatrics, Division of Allergy and Immunology, University of British Columbia, Vancouver; ^eSwiss Institute of Allergy and Asthma Research (SIAF), University of Zurich, Davos; ^fthe Department of Women and Children's Health (Pediatric Allergy), School of Life Course Sciences, Faculty of Life Sciences and Medicine and Peter Gorer Department of Immunobiology, School of Immunology and Microbial Sciences, King's College London, London; ^gChildren's Allergy Service and Evelina Children's Hospital, Guy's and St Thomas's NHS Foundation Trust, London; ^hthe Pediatric Allergy and Clinical Immunology Research Unit, Division of Allergy and Immunology, Department of Pediatrics, Faculty of Medicine, Chulalongkorn University, King Chulalongkorn Memorial Hospital, The Thai Red Cross Society, Bangkok; ⁱAlbert Einstein Medical School, São Paulo; ^jCHU Lille, University of Lille, Pediatric Pulmonology and Allergy Unit, Hôpital Jeanne de Flandre, Lille; ^kUniversity Hospitals of Geneva and University of Geneva, Geneva; ^lthe National Institute of Children Health, National Reference Center of Allergy, Asthma and Immunology, Lima; ^mthe International Clinic, B&D Health Clinic, Lima; ⁿthe Center for Food Allergy and Asthma Research, Northwestern University Feinberg School of Medicine, Chicago; ^oAnn & Robert H. Lurie Children's Hospital of Chicago, Chicago; ^pthe Pediatric Allergy, Immunology and Rheumatology Unit, Children's Hospital, Ain Shams University, Cairo; ^qMurdoch Children's Research Institute and ^rthe Department of Paediatrics, University of Melbourne, Melbourne; ^sthe Division of Paediatric Allergy, Department of Paediatrics, University of Cape Town, Cape Town; ^tinVIVO Planetary Health Group of the World-wide Universities Network; ^uthe Department of Paediatrics, National University of Singapore, Singapore; ^vthe Skin and Allergy Hospital, Helsinki University Hospital and University of Helsinki, Helsinki; ^wthe Allergy Service, Instituto Nacional de Pediatría, Mexico City; ^xthe Food Allergy Centre, Department of Woman and Child Health, Padua University Hospital, Padua; ^ythe Allergy Department, National and Kapodistrian University of Athens, Athens; ^zthe Division of Infection, Immunity and Respiratory Medicine, University of Manchester, Manchester; ^{aa}the Department of Pediatrics, Nippon Medical School, Sendagi, Bunkyo-ku, Tokyo; ^{bb}Clinical and Experimental Sciences & Human Development in Health, Faculty of Medicine,

University of Southampton, Southampton; ^{cc}the NIHR Southampton Biomedical Research Centre, University Hospital Southampton NHS Foundation Hospital, Southampton; ^{dd}David Hide Asthma and Allergy Research Centre, St Mary's Hospital, Isle of Wight; ^{ee}the Division of Pediatric Allergy, Department of Pediatrics, Koc University School of Medicine, Istanbul; ^{ff}The Elliot and Roslyn Jaffe Food Allergy Institute, Division of Allergy and Immunology, Kravis Children's Hospital, Department of Pediatrics, Icahn School of Medicine at Mount Sinai, New York; ^{gg}the Division of Allergy, Immunology and Transplantation, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Bethesda; ^{hh}Pediatric Allergy and Immunology, University of Colorado/Childrens Hospital Colorado, Boulder; ⁱⁱthe Department of Pediatrics, Chinese University of Hong Kong, Hong Kong; ^{jj}the Department of Pediatric Respiratory Medicine, Immunology and Critical Care Medicine, Charité Universitätsmedizin Berlin, Berlin; ^{kk}the Institute of Laboratory Medicine, Philipps University Marburg, Member of the German Center for Lung Research (DZL), Member of Universities Giessen and Marburg Lung Center, Marburg; and ^{ll}the Department of Clinical Immunology and Allergology, Laboratory of Immunopathology, Sechenov University, Moscow.

*Co-senior authors.

L. M. Wheatley and A. Togias' authorship of this report does not constitute endorsement by the US National Institute of Allergy and Infectious Disease or by any other US government agency. E. M. Abrams is an employee of the Public Health Agency of Canada (PHAC). The views expressed in the article are her views and not those of the PHAC.

Received for publication August 10, 2021; revised October 20, 2021; accepted for publication October 22, 2021.

Corresponding author: Kari C. Nadeau, MD, PhD, Sean N. Parker Center for Allergy and Asthma Research at Stanford University, 240 Pasteur Dr, BMI Rm 1755, Palo Alto, CA 94304. E-mail: knadeau@stanford.edu.

The CrossMark symbol notifies online readers when updates have been made to the article such as errata or minor corrections

0091-6749/\$36.00

© 2021 American Academy of Allergy, Asthma & Immunology

<https://doi.org/10.1016/j.jaci.2021.10.018>

INFORMATION FOR CATEGORY 1 CME CREDIT

Credit can now be obtained, free for a limited time, by reading the review articles in this issue. Please note the following instructions.

Method of Physician Participation in Learning Process: The core material for these activities can be read in this issue of the Journal or online at the JACI Web site: www.jacionline.org. The accompanying tests may only be submitted online at www.jacionline.org. Fax or other copies will not be accepted.

Date of Original Release: December 2021. Credit may be obtained for these courses until November 30, 2022.

Copyright Statement: Copyright © 2021-2022. All rights reserved.

Overall Purpose/Goal: To provide excellent reviews on key aspects of allergic disease to those who research, treat, or manage allergic disease.

Target Audience: Physicians and researchers within the field of allergic disease.

Accreditation/Provider Statements and Credit Designation: The American Academy of Allergy, Asthma & Immunology (AAAAI) is accredited by the Accreditation Council for Continuing Medical Education (ACCME) to provide continuing medical education for physicians. The AAAAI designates this journal-based CME activity for a maximum of 1.00 *AMA PRA Category 1 Credit*[™]. Physicians should claim only the credit commensurate with the extent of their participation in the activity.

List of Design Committee Members: Vanitha Sampath, PhD, Elissa M. Abrams, MD, MPH, Bahman Adlou, MS, Cezmi Akdis, MD, Mübecce Akdis, MD, PhD, Helen A. Brough, PhD, MBBS, FRCPC, Susan Chan, MBBS, MD, Pantipa Chatchatee, MD, R. Sharon Chinthrajah, MD, Renata Rodrigues Cocco, MD, PhD, Antoine Deschildre, MD, Philippe Eigenmann, MD, Cesar Galvan, MD, Ruchi Gupta, MD, MPH, Elham Hossny, MD, PhD, Jennifer J. Koplin, PhD, Gideon Lack, MBBS, FRCPC, MD, Michael Levin, MBBS, Lynette P. Shek, MBBS, FRCPC, Miika Makela, MD, PhD, David Mendoza-Hernandez, MD, Antonella Muraro, MD, PhD, Nikolaos G. Papadopoulos, MD, PhD, FRCP, Ruby Pawankar, MD, PhD, Kirsten P. Perrett, MBBS, FRACP, PhD, Graham Roberts, DM, FRCPC, Cansin Sackesen, MD, Hugh Sampson, MD, Mimi L.K. Tang, MBBS, PhD, FRACP, FRCPA, Alkis Togias, MD, Carina Venter, PhD, RD, Christopher Michael Warren, PhD, Lisa M. Wheatley, MD, MPH, Gary W.K. Wong, MD, FRCPC, Kirsten Beyer, MD, Kari C. Nadeau, MD, PhD,^a and Harald Renz, MD, PhD (authors); Zuhair K. Ballas, MD (editor)

Disclosure of Significant Relationships with Relevant Commercial Companies/Organizations: E. M. Abrams is an employee of the Public Health Agency of Canada (PHAC). C. Akdis reports grants from the Swiss National Science Foundation, EU CURE, and Novartis Research Institutes (Basel Switzerland); is Chair of the European Academy of Allergy and Clinical Immunology (EAACI) Guidelines on Environmental Science in Allergic diseases and Asthma; and is Editor-in-Chief of *Allergy*. M. Akdis reports grants from the Swiss National Science Foundation, EU CURE, and Novartis Research Institutes (Basel Switzerland). H. A. Brough reports receiving research grants from the National Institutes of Health (NIH), Aimmune, and DBV Technologies and speaker fees from DBV Technologies and Sanofi; is Pediatric Section Chair, the EAACI; and is President Elect Royal Society of Medicine. S. Chan reports grants from the NIH and Aimmune and has received a medical device from Primus. R. S. Chinthrajah reports grants from the National Institute of Allergy and Infectious Diseases (NIAID), CoFAR, Aimmune, DBV Technologies, Astellas, Regeneron, Stanford Maternal and Child Health Research Institute, and Food Allergy Research & Education (FARE) and is an Advisory Board Member at Alladapt Therapeutics, Novartis, Genentech, Sanofi, Allergen, and Nutricia. A. Deschildre receives consulting fees, honoraria, or support for attending meetings/travel from Aimmune Therapeutics, DBV Technologies, Nestlé Health Science, Nutricia, Novartis, and AstraZeneca; participated on a Data Safety Monitoring Board or Advisory Board at Aimmune and Novartis; has a leadership role at SFA (société Française d'Allergologie) (ongoing), and SP2A (Société Pédiatrique de Pneumologie et Allergologie) (stopped in 2019); and is

past president of the Cercle d'investigations Cliniques et Biologiques en Allergie Alimentaire. P. Eigenmann receives grants, consulting fees, speaking fees, honorarium, royalties, stock options, or travel support from Ulrich Muller Gierock Foundation, Thermo Fisher Scientific, DBV Technologies, Nestlé, Danone, Novartis, Abbott, Alexia, University Hospitals of Geneva, ALK, UpToDate, Elsevier, and EAACI. C. Galvan reports payment from Reckitt Benckiser Mead Johnson. D. Mendoza-Hernandez reports payment or honoraria from Nestlé and Thermo Fisher Scientific. J. J. Koplin reports grants from the National Health and Medical Research Council of Australia. G. Lack receives grants from NIAID, NIH, FARE, MRC & Asthma UK Centre, UK Department of Health through the *National Institute for Health Research*, National Peanut Board, The Davis Foundation, UK Food Standards Agency, and Medical Research Council; consulting fees, payment, honorarium, or travel support from Novartis, Sanofi-Genzyme, Regeneron, ALK-Abelló, Lurie Children's Hospital, ALK-Abelló, Northwestern University Feinberg School of Medicine, DBV Technologies, the American Academy of Allergy, Asthma & Immunology (AAAAI), and Nestlé; is on the Scientific Advisory Board in DBV Technologies, Lurie Children's Hospital, ALK-Abelló, and Novartis; and holds stock or stock options for DBV Technologies and Mighty Mission Me. A. Muraro discloses speaker's fee from Mylan, Viatrix, Aimmune, DVB Technologies, Nestlé Health Institute, Nestlé Purina, and Nutricia. N. G. Papadopoulos reports grants from Gerolymatos International SA and Capricare; payment or honoraria from HAL, Menarini/Faes Farma, MSD, Biomay, Novartis, Nutricia, Sanofi, Boehringer Ingelheim, Mylan/Meda, and Asit Biotech; and participating on the Advisory Board at HAL, Menarini/Faes Farma, Novartis, Nutricia, GlaxoSmithKline (GSK), AstraZeneca, and Mylan/Meda. K. P. Perrett reports grants from Melbourne Children's Clinician-Scientist Fellowship; research grants from the National Health and Medical Research Council, the Immune Tolerance Network, DBV Technologies, and GSK; consulting fees from Aravax; and serving as Chair of the Scientific Advisory Board for Allergy-Pal. G. Roberts reports grants from United Kingdom Food Standards Agency to assess the epidemiology of food allergy and is a member of the EAACI Food Allergy Guideline Group. C. Sackesen reports payment, honoraria, or travel support from Abbott Company and Nutricia; is the head of a food allergy group at the Turkish Society of Allergy and Clinical Immunology; and has patents planned for a twice-baked cow's milk product as a hypoallergenic milk food. H. Sampson reports grants, consulting fees, stock options, and travel support from NIAID, NIH, Elsevier, DBV Technologies, N-Fold LLC, Siolta Therapeutics, and DBV Technologies. M.L.K. Tang reports grants, drugs received, contracts, travel support, leadership or fiduciary role, stocks or stock options, or consulting fees from the National Health and Medical Research Council (Australia), Prota Therapeutics Pty Ltd, Murdoch Children's Research Institute, Pfizer, Prota Therapeutics, the EAAACI, the Asia Pacific Association of Allergy, Asthma and Clinical Immunology, the AAAAI, the International Union of Immunological Societies, the Australasian Society of Clinical Immunology and Allergy, Anaphylaxis Australia Incorporated, the Melbourne Academic Centre for Health, and the World Allergy Organization; is member of the International Expert Panel on Guidelines for Food Allergy in Schools, Health World Ltd; has patents including "A method of including tolerance to an antigen and allergy treatment"; and is an employee of Prota Therapeutics. A. Togias is a paid employee of the US Federal Government, which funds food allergy research in the United States and across the globe. C. Venter reports grants, payment, or honoraria from Danone, Nestlé Nutrition Institute, Reckitt Benckiser, Abbott, Before Brands, Sifter, and Else Nutrition and leadership or fiduciary role at the American Academy of Allergy, Asthma and Immunology, the European Academy of Allergy, Asthma and Immunology, and Food Allergy Research and Education. C. M. Warren reports grants from the NIAID and FARE. K. Beyer received support (grants, consulting or speaker fees) from Aimmune Therapeutics, ALK, DBV, Bencard, Nestlé, Novartis, and Allergopharma; was on the Advisory Board or the Data Safety Monitoring Board of Aimmune Therapeutics, Bencard, DBV, Nestlé, and Novartis; has a leadership or fiduciary role at AGATE (Anaphylaxis Training and Education), GPA (German

Society for Pediatric Allergology & Environmental Medicine), DGKJ (German Society for Allergology & Clinical Immunology), and DAAB (German Allergy and Asthma Association). K. C. Nadeau reports grants from the NIAID, the National Heart, Lung, and Blood Institute, the National Institute of Environmental Health Sciences, and FARE; is Director of the World Allergy Organization; Advisor at Cour Pharma; consultant for Excellergy, Red tree ventures, and Phylaxis; cofounder of Before Brands, Alladapt, Latitude, and IgGenix; and National Scientific Committee member at the Immune Tolerance Network, and NIH clinical research centers, outside the submitted work; patents include “Mixed allergen composition and methods for using the same,” “Granulocyte-based methods for detecting and monitoring immune system disorders,” “Methods and assays for detecting and quantifying pure subpopulations of white blood cells in immune system disorders,” and “Methods of isolating allergen-specific antibodies from humans and uses thereof.” H. Renz reports grants from Deutsche Zentrum für Lungenforschung (DZL, German Lung Center, no. 82DZL00502) and Deutsche Forschungsgemeinschaft (DFG)-funded SFB 1021. The rest

of the authors declare that they have no relevant conflicts of interest. Z. K. Ballas (editor) disclosed no relevant financial relationships.

Activity Objectives:

1. To recognize emerging guidelines regarding prevention of food allergy in children.
2. To understand important trends in food allergy management.
3. To identify challenges to implementing emerging therapies for food allergy.

Recognition of Commercial Support: This CME activity has not received external commercial support.

List of CME Exam Authors: Kelsey Lecerf, MD, Christopher Brooks, MD, Kasey Lanier, MD, Christian Coletta, MD, and Rebecca Scherzer, MD

Disclosure of Significant Relationships with Relevant Commercial Companies/Organizations: The examination authors declare no relevant conflicts of interest.

The prevalence of food allergy (FA) is increasing in some areas of the globe, highlighting the need for better strategies for prevention, diagnosis, and therapy. In the last few decades, we have made great strides in understanding the causes and mechanisms underlying FAs, prompting guideline updates. Earlier guidelines recommended avoidance of common food allergens during pregnancy and lactation and delaying the introduction of allergenic foods in children aged between 1 and 3 years. Recent guidelines for allergy prevention recommend consumption of a healthy and diverse diet without eliminating or increasing the consumption of allergenic foods during pregnancy or breast-feeding. Early introduction of allergenic foods is recommended by most guidelines for allergy prevention after a period of exclusive breast-feeding (6 months [World Health Organization] or 4 months [European Academy of Allergy and Clinical Immunology]). New diagnostics for FA have been developed with varied availability of these tests in different countries. Finally, the first oral immunotherapy drug for FA was approved by the US Food and Drug Administration and European Medicines Agency in 2020. In this review, we will address the global prevalence of FA, our current understanding of the causes of FA, and the latest guidelines for preventing, diagnosing, and treating FA. We will also discuss similarities and differences between FA guidelines. (J Allergy Clin Immunol 2021;148:1347-64.)

Key words: Food allergy, guidelines, prevention, treatment, epidemiology

Food allergy (FA) prevalence is increasing in some regions of the world.^{1,2} However, geographical variability in the incidence, type, and clinical presentation of FA as well as variations in symptoms and clinical phenotypes due to race, ethnicity, age, and coexisting allergic diseases exist.^{3,4}

The increasing incidence of FA in certain regions of the world has spurred efforts to understand the causes and mechanisms underlying FA and tolerance to optimize diagnostics and find ways to prevent or treat FA. Early guidelines recommended dietary avoidance or delayed introduction of allergenic foods in infants to prevent FAs.⁵ However, later studies either did not see a

benefit of delayed introduction or indicated that early introduction may potentially be beneficial in preventing FA. Findings from recent studies of early-life dietary interventions for FA prevention have led to revised guidelines, moving away from an avoidance approach of allergenic foods to actively recommending introduction of allergenic foods in the first 4 to 6 months of life.⁶⁻⁸ A number of novel diagnostics have been developed, but these are still mainly performed in research laboratories and not readily available. In 2020, Palforzia, an oral immunotherapy drug for peanut allergy, obtained US Food and Drug Administration approval. This was an important milestone for FA therapy because it was the first-ever drug approved for the treatment of FA. Palforzia, a biological oral immunotherapy drug for FA, is composed of peanut allergen powder for the treatment of peanut allergy.⁹

We have gained valuable insights into FA over the last few years regarding the causes and the mechanisms of FA as well as new developments into diagnostics, prevention strategies, and treatments. The aim of this article was to provide an overview of the incidence of FA, causes, prevention strategies, diagnostic methods, and recommendations for therapies in FA, and report on global similarities and differences in FA guidelines.

EPIDEMIOLOGY OF FA

More than 160 foods are known to cause FAs, with varying prevalence rates by specific food and population affected.¹⁰⁻¹² Large population-based studies using double-blind placebo-controlled food challenges (DBPCFCs), the criterion standard for the diagnosis of FA, hold promise for accurate FA prevalence assessment; however, DBPCFCs and oral food challenges (OFCs) are resource-intensive and pose risk of severe allergic reactions, thereby raising concerns about low participation rates among participants and selection bias. Therefore, prevalence data using OFCs are very limited.¹³ A 2013 survey of 89 countries found that only 10% of countries had prevalence data based on OFC.¹⁴ Instead of OFCs, surrogate markers are often used for determination of FA. These include self-reported clinical history of FA, clinical or hospital visits for FA, or determination of allergen-specific IgE (sIgE) either by skin prick test (SPT) or by serum sIgE. sIgE tests are associated

Abbreviations used

AAAAI:	American Academy of Allergy, Asthma & Immunology
APT:	Atopy patch test
ASCIA:	Australasian Society of Clinical Immunology and Allergy
DBPCFC:	Double-blind placebo-controlled food challenge
EAACI:	European Academy of Allergy and Clinical Immunology
FA:	Food allergy
OFC:	Oral food challenge
OIT:	Oral immunotherapy
QOL:	Quality of life
sIgE:	Allergen-specific IgE
SPT:	Skin prick test

with high rates of false positives, leading to overestimation. A false-positive diagnosis carries the risk of nutritional deficiency and significantly impacts quality of life (QOL). Misdiagnosis also leads to an increased economic burden on the health care system, with increased costs associated with specialist referral, additional testing, and unnecessary medication prescriptions. Self-reporting of FAs also leads to overestimation because these may also include food intolerances or toxicities. For example, a study found that 14% of families reported a milk allergy in their infant, but milk allergy could be confirmed in only 1.4%.¹⁵

The Melbourne HealthNuts and SchoolNuts studies are large population-based studies with challenge-confirmed FA. These studies provide the highest quality of prevalence data and show rates of more than 10% in infants¹⁶ and 4% to 5% in older children and young adolescents.^{17,18} However, a limitation of the HealthNuts study is that it clinically evaluated only a few of the food allergens—egg, peanut, sesame, cow's milk, and shrimp in infants. The SchoolNuts study evaluated only 15 food allergens.

In the United States, 2 large (N > 38,000) cross-sectional well-designed population-based surveys have been conducted. Reported FAs were considered as convincingly IgE-mediated if reported symptoms to specific allergens met well-defined criteria consistent with IgE-mediated reactions. The studies found that 7.6% of children¹⁹ and 10.8% of adults had probable FA.²⁰ In children with FA, 40% were affected by more than 1 FA. The study has the potential of overdiagnosing FA because vomiting is one of the self-reporting symptoms, which is not exclusive for FA. It is also a symptom of food protein–induced enterocolitis syndrome, early-onset eosinophilic esophagitis, and other food intolerances.

Initial reports on the prevalence of FA in Europe did not consider the wide variety of eating habits of the various geographical areas and cultures. Data were available for specific countries or regions, with extrapolations for other areas. The EuroPrevall research project addressed this diversity by applying the same methodology in various centers across the continent.^{21,22} In the study, children with suspected FA symptoms were diagnosed via OFCs and sIgE (SPT or serum measurements). Birth cohorts with more than 12,000 participants revealed a mean incidence at age 2 years of 1.23% for hen's egg allergy, with country-specific incidence from 0.07% in Greece to 2.18% in the United Kingdom²³ and 0.54% for cow's milk allergy (ranging from <0.3% in Lithuania, Germany, and Greece to 1% in the Netherlands and the United Kingdom).²⁴ The types of

FAs differed substantially between countries, with fish and shrimp allergy being more prevalent in the Mediterranean area and in Iceland, and nuts, fruits, and vegetable allergies being more prevalent in Central Europe.²⁵ Among the children in the EuroPrevall studies, 23.6% had non-IgE-mediated cow's milk allergy, with most children in the United Kingdom reporting non-IgE-mediated cow's milk allergy, whereas the Netherlands reported no child with non-IgE-mediated cow's milk allergy. In this study, non-IgE-associated cow's milk allergy was defined as cow's milk allergy diagnosed by DBPCFC with sIgE to milk less than 0.35 kU/L and SPT wheal diameter less than 3 mm.²⁴

In many Asian countries, South and Central America, and Africa,²⁶ FA is thought to be uncommon; however, reliable epidemiological data are limited.^{27,28} An epidemiological investigation of FA in an urban area of Wenzhou, China, found FA prevalence to be at least 0.84% among children aged 3 to 6 years based on OFC and sIgE or SPT.²⁹ Using a definition of probable FA as reporting allergic symptoms within 2 hours of ingestion of a specific food plus the presence of allergic sensitization to the specific food (positive sIgE and/or positive SPT result), the EuroPrevall-INCO surveys found that the prevalence of FA was 1.50% (Hong Kong), 0.21% (Guangzhou, China), 0.69% (Shaoguan, China), and 0.14% (India).²⁷

In Africa, most studies use sensitization as a surrogate marker for allergy²⁶ or are performed in high-risk populations. The South African Food Allergy study is the sole study using challenge-proven FA as an outcome in an unselected population.³⁰ The study showed marked urban-rural differences, with the prevalence of FA of 2.5% in children aged 1 to 3 years in Cape Town, but only 0.5% in the rural Eastern Cape.³¹ Unusual allergens may occur in various parts of Africa, including Mopane worms,^{32,33} and there are areas in Africa with high rates of both sensitization to galactose- α -1,3-galactose and allergy to mammalian meat (galactose- α -1,3-galactose syndrome).³⁴

The studies of prevalence in South and Central America mostly use parent-proxy or self-reported allergic reaction symptoms, with few studies measuring SPT wheal diameter. Prevalence rates reported in these studies range between 0.9% and 52%.³⁵⁻⁴³ Foods that trigger these reactions are similar to those in other parts of the world although in some countries such as Mexico, Costa Rica, and Colombia, sensitizations to tropical vegetables and fruits have been found.^{40,42,44} In tropical regions of South America, oral mite syndrome (due to mite-contaminated wheat flour) has been described and in a case series study in Venezuela represented the third most reported cause of anaphylaxis.⁴⁵ Recently, more studies on oral allergy syndrome and galactose- α -1,3-galactose allergy are also being reported in Mexico and Colombia, respectively.^{46,47}

It should be noted that the data relied on to estimate global FA prevalence are subject to substantial limitations, most notably the nonspecificity of proxy measures of allergic sensitization (eg, SPT and sIgE) and lack of concordance between survey-reported and food challenge–confirmed prevalence estimates. However, the overall consensus is that FA has significantly increased in developed countries, potentially due to changes in environmental exposures and lifestyle. The study by Botha et al³¹ using OFCs to confirm a diagnosis of FA found a significant increase in the prevalence of FA between children born in urban and rural areas. This suggests that urbanization is leading to increases in FA.

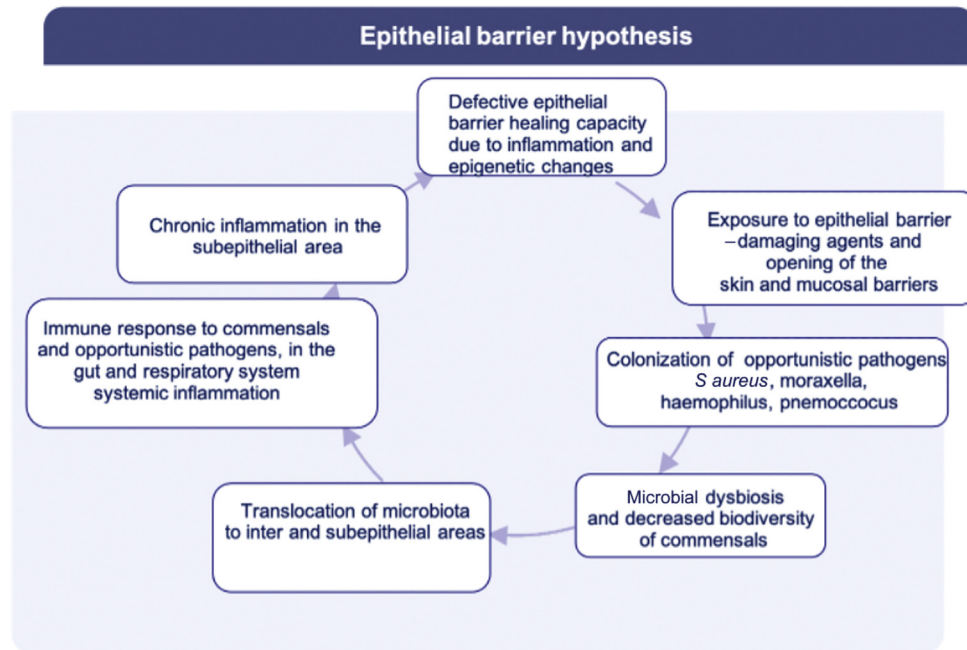


FIG 1. The physiopathology of epithelial barrier hypothesis: The vicious cycle of chronic epithelial barrier leakiness. Genetic defects in barrier-related molecules or exposure to epithelial barrier-damaging agents cause an opening of the skin and mucosal tight junction barriers. This is followed by translocation of microbiota to inter and subepithelial areas and colonization of opportunistic pathogens, such as *Staphylococcus aureus*, moraxella, haemophilus, and pneumococcus. An immune response develops toward commensals and opportunistic pathogens in the gut and respiratory system and a systemic inflammation takes place. In most cases of allergic diseases, a systemic type 2 inflammation predominates, and is directed against not only allergens but also commensals and opportunistic pathogens. For example, anti-*S aureus* antibodies show a very high prevalence in asthma chronic rhinosinusitis and atopic dermatitis. This is associated with microbial dysbiosis and decreased biodiversity of commensals. Chronic inflammation in the subepithelial area prevails, as one of the main reasons for the development of chronic diseases in the affected tissues. Defective epithelial barrier healing capacity due to inflammation and epigenetic changes take place, instigating a vicious circle of leaky barriers, microbial dysbiosis, and chronic inflammation.

Although high-quality prevalence data for FA are lacking for many geographical regions and age groups, increases in FA prevalence are supported by hospitalization rates for FA. A nationwide survey in the United States of hospitalization due to pediatric food-induced anaphylaxis found a significantly increasing trend from 1.2 per 100,000 children in 2006 to 1.5 per 100,000 children in 2012. The leading causes of hospitalizations due to food-induced anaphylaxis were peanut, followed by tree nuts and seeds, and milk products.⁴⁸ In Australia, a 4-fold increase in hospitalizations for FA-related anaphylaxis was observed between 1998/1999 and 2011/2012 (2.0-8.2 per 100,000).⁴⁹ However, additional studies using standardized methodologies are necessary for accurate detection of FA to better understand the true extent of the problem and its impact on health services.

CAUSES OF FA

FA is a complex immune disorder caused by specific genetic variants in combination with environmental and nutritional exposures. Genome-wide association studies have found certain loci for FA including genes involving barrier integrity (filaggrin and serine protease inhibitor), immune function, and

others.⁵⁰⁻⁵³ However, the increase in FA is too rapid to be due to genetics alone, and migration studies show us that these increases can occur in a single generation.⁵⁴ Epigenetics provides a framework for understanding the mechanisms by which environmental and nutritional factors interact with genetic factors to mediate FA.

Innate lymphoid cells, which contribute to type 2 immune responses, have also been implicated in mediating FA. Innate lymphoid cells are localized at barrier surfaces of the airways, gut, and skin and form a link between innate and adaptive immunity.⁵⁵

A number of studies have evaluated the role of nutrition and diet in the development of FA.⁵⁶ There is strong evidence that early introduction of allergens in infants, such as the introduction of peanut or egg beginning at age 4 to 6 months, prevents the development of FA.⁵⁷ For milk, the window of opportunity is probably much earlier—while infants fed cow's milk formula from birth rarely develop cow's milk allergy, allergy is seen in infants who are temporarily supplemented during the first week of life with avoidance thereafter.^{56,58} Less is known about the development of FA in later life although anecdotally, novel FAs are seen in populations when new foods are introduced into the national diet.⁵⁹ Besides allergens, other dietary factors that have

Types of FA	Representative patient	Mechanism	Risk Factor	Incidence Global variability
IgE-mediated,	Peanut anaphylaxis		Atopy, acute reactions, infections, exercise, alcohol etc,	1%-10%, depending on age, large variability by allergens globally
Non-IgE-mediated,	Acute FPIES with milk allergy		Family history, short duration of breast-feeding, ethnicity?	0.15%-0.7%, by 2 years of age
Mixed (IgE non-IgE-mediated),	Eosinophilic esophagitis		Atopy, early-life antibiotic exposure, family history	10-57/100,000

FIG 2. Example of IgE-mediated, non-IgE-mediated, and mixed IgE- and cell-mediated FAs along with their prevalence, mechanisms, and examples of causal allergens. *FPIES*, Food protein-induced enterocolitis syndrome. Figures for IgE-mediated, non-IgE-mediated, and mixed adapted from Otsu and Dreskin,⁷⁸ Brown-Whitehorn and Cianferoni,⁷⁹ and Mulder and Justinich,⁸⁰ respectively.

been associated with FA are prebiotics, probiotics, vitamin D, and omega-3 polyunsaturated fatty acids. However, the evidence for these associations is weak. Although specific microbiome patterns are associated with FA or tolerance,⁶⁰ the role of prebiotics, probiotics, or symbiotics in mediating these effects is not well understood.

Allergens, certain bacteria, fungus, viruses, laundry and dishwasher detergents, household cleaners, surfactants, enzymes and emulsifiers in processed food, cigarette smoke, particulate matter, diesel exhaust, ozone, nanoparticles, and microplastics all

disrupt the epithelial barrier.⁶¹⁻⁷² According to the epithelial barrier hypothesis (Fig 1), exposure to many of these substances damages and initiates inflammation around the epithelium that covers the surface of the skin, and respiratory, urogenital, and gastrointestinal tracts.⁷³ Epithelial cell activation and release of epithelial cell cytokines, such as IL-25, IL-33, and thymic stromal lymphopoietin, play a major role in the development and exacerbation of allergic diseases.^{74,75}

The importance of a diverse microbiome in reducing the risk of FA is now recognized. A greater number of siblings and

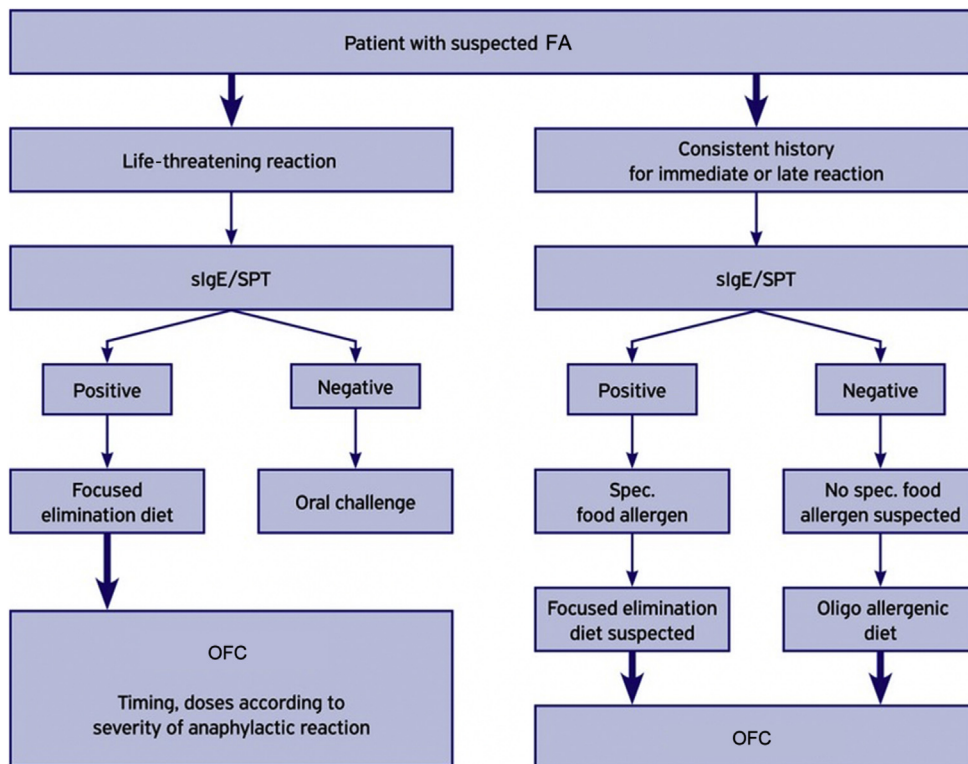


FIG 3. EAACI Diagnostic guidelines in 2014.

dog ownership, both of which can increase microbial diversity, have been associated with a reduced risk of developing FA.^{76,77}

Clearly, the causes of FA are multifactorial and are likely to result from a complex interaction of genetic, dietary, and environmental factors. Arguably, at present, the 2 most important risk factors for FA development early in life are skin barrier dysfunction and delayed introduction of allergenic solids. However, remarkably little remains known about the determinants and mechanisms of adult-onset FA, wherein oral tolerance is lost among patients who previously tolerated the offending food without incident. In many cases of adult-onset FA to previously tolerated foods, there is a period of abstinence to the food (either instructed by physician often due to atopic dermatitis or the initiation of a new exclusionary diet, before onset of adult-onset FA). Furthermore, many of the putative causal mechanisms outlined above have not been extensively tested in human subjects via well-designed randomized controlled trials and therefore are of limited utility, both clinically and for informing policies aiming to reduce the public health burden of FA.

DIAGNOSTICS

FAs are primarily IgE-mediated; however, mixed IgE- and cell-mediated, and non-IgE-mediated FAs also exist. Fig 2 list examples of IgE-mediated, non-IgE-mediated, and mixed IgE- and cell-mediated FAs along with their prevalence, and mechanisms.

The international criterion standard for FA diagnosis is the DBPCFCs. However, because these are time and resource-intensive and pose risk of severe allergic reactions, other surrogate diagnostic tests are often used.

In Europe, the 2014 European Academy of Allergy and Clinical Immunology (EAACI) Guidelines for FA Diagnosis were developed according to the Institute of Medicine/Guidelines International Network reference, involving all the relevant stakeholders and combining the level of evidence with the experts' opinion, when evidence was lacking. Grades A to D recommendations were created on the basis of level of evidence available, with D indicating when experts' opinion had to complement the existing data. The EAACI algorithm for FA diagnosis (Fig 3) includes 5 essential steps: (1) the patient's clinical history with the use of structured questions (Grade D), (2) determination of sensitization with standardized SPT and/or sIgE directed by case history as well as the use of molecular allergology with component-resolved diagnostics to better profile the patient (Grades A-C) (3) elimination diet for diagnostic purposes, that is, short-term avoidance 2 to 4 weeks (Grade D), (4) an OFC to definitely confirm or exclude the diagnosis (Grade D), and (5) evaluation for non-IgE-mediated FA when the history is convincing and SPT result and sIgE are negative (Grade D).

In the United States, the 2010 Guidelines for the Diagnosis and Management of Food Allergy recommends the use of SPT or sIgE to evaluate FA.⁸¹ They do not recommend the use of intradermal testing, total serum IgE, or the atopy patch test (APT). The guidelines stated that a combination of 2 or more of SPT plus

TABLE I. Global FA documents on prevention of FA since 2002

Year	North America	Australia and New Zealand	Europe	Asia
2004			EAACI ⁹⁵	
2005		ASCIA ⁹⁶		
2006	ACAAI ⁵			
2007			ESPGHAN ⁹⁷	
2008	AAP ⁹⁸		EAACI ⁹⁹	
2009			DGAKI and DGKJ ¹⁰⁰	
2010	NIAID ¹⁰¹	ASCIA ¹⁰²		AMS-MOH ^{84,103}
2012			Finnish Allergy Program 2008-2018 ¹⁰⁴	
2013	CPS and CSACI ¹⁰⁵			
2014			DGAKI, DGKJ, ¹⁰⁶ and EAACI ¹⁰⁷	
2015	AAP ¹⁰⁸			GLAD-P ¹⁰⁹
2016			ISPAI and ISP ¹¹⁰	HKIA ^{111,112} and GLAD-P ¹¹³
2017	NIAID ¹¹⁴		BSACI ¹¹⁵ and ESPGHAN ¹¹⁶	JSPACI, ¹¹⁷ PSAIL, and PSPGHN ¹¹⁸
2018			BSACI, ¹¹⁹ SACN, and COT ¹²⁰	APAPARI ⁸⁷
2019	AAP, ⁹⁸ CPS, and CSACI ¹²¹	ASCIA ¹²²		AMSMOH ¹²³
2020			EAACI ⁵⁷	JSPACI, ⁸⁶ Chinese Expert Consensus, ¹²⁴ ISPGHAN ¹²⁵
2021	AAAAI, ACAAI, and CSACI ⁶			MAP ¹²⁶

AAP, American Academy of Pediatrics; ACAAI, American College of Asthma, Allergy and Clinical Immunology; AMSMOH, Academy of Medicine, Singapore Ministry of Health; APAPARI, Asia Pacific Association of Pediatric Allergy, Respiratory & Immunology; BSACI, British Society of Allergy and Clinical Immunology; COT UK, Committee on Toxicity United Kingdom; CPS, Canadian Pediatric Society; CSACI, Canadian Society of Allergy and Clinical Immunology; DGAKI, German Society for Allergy and Clinical Immunology; DGKJ, German Society for Pediatric and Adolescent Medicine; ESPGHAN, European Society for Paediatric Gastroenterology, Hepatology and Nutrition; GLAD-P, World Allergy Organization-McMaster University Guidelines for Allergic Disease Prevention; HKIA, Hong Kong Institute of Allergy; ISP, Italian Society of Pediatrics; ISPAI, Italian Society of Paediatric Allergy and Immunology; ISPGHAN, Indian Society of Pediatric Gastroenterology, Hepatology and Nutrition; JSPACI, Japanese Society of Pediatric Allergy and Clinical Immunology; MAP, Malaysia Allergy Prevention; NIAID, National Institute of Allergy and Infectious Diseases; PSAIL, Philippine Society of Allergy, Asthma and Immunology; PSPGHN, Philippine Society for Paediatric Gastroenterology, Hepatology and Nutrition; SACN, Scientific Advisory Committee on Nutrition.

serum allergen-specific sIgE and/or APTs marginally improved positive and negative predictive values, but did not obviate the need for DBPCFC. The guideline, however, did not recommend the use of a combination of tests over the use of sIgE or SPT alone. Food elimination diets may be useful particularly in the diagnosis of non-IgE-mediated and mixed IgE/non-IgE-mediated FA, where no diagnostic test can otherwise identify the causative food. However, for definitive diagnosis, DBPCFCs were cited as the criterion standard for diagnosing FA, with allowance for single-blind and open challenges to be used in the clinical setting. The 2010 guideline states that further studies are necessary to determine the efficacy of food allergen epitope specificity and component protein-based assays. The diagnosis of eosinophilic gastrointestinal diseases is supported by dietary elimination, OFC, endoscopy, and esophageal biopsy. SPTs, sIgE tests, and APTs are not diagnostic but can be used to support the diagnosis. Food protein-induced enterocolitis syndrome, food protein-induced allergic proctocolitis, and food protein-induced enteropathy syndrome can be diagnosed by medical history in combination with an elimination diet, and an OFC. Allergic contact dermatitis and systemic contact dermatitis can be diagnosed by a combination of medical history, including resolution of symptoms when the causative food is avoided, and positive patch test results, whereas the diagnosis of IgE-mediated contact urticaria is supported by history, including the absence of symptoms while the causative food is avoided, positive sIgE test results or

SPT results, and positive immediate epicutaneous skin test results (eg, positive immediate responses to APTs).

Similar to international approaches, a thorough clinical history that considers the symptoms indicative of IgE-mediated allergic reactions to food is the first-line approach in diagnosing FA in Australia. Second-line, evidence-based *in vivo* (SPT) and *in vitro* (sIgE) investigations of sensitization are essential adjunct tools, which the Australasian Society of Clinical Immunology and Allergy (ASCIA) specifically advises should be used only in conjunction with clinical history. In carefully selected patients to confirm or exclude FA, medically supervised OFCs are performed. Recently, because of the increased use of widely available online allergy testing services, ASCIA has written a position paper that strongly recommends against using online allergy tests,⁸² due to potential harm (even if evidenced-based tests are ordered because advice is given in the absence of personal consultation), resulting in misdiagnosis, ineffective treatments, increased costs for the patient or caregiver, and a greater burden on the health care system. In Australia, most pediatric allergy clinics use SPT in preference to sIgE testing because results are immediately available. Consistently, SPTs have been shown to have a high sensitivity, but low specificity, and so more accurate diagnostic testing is being actively researched.

National guidelines of South and Central America are represented by a minority of the total of 36 countries. Clinical

history and physical examination are considered the most important step to proceed with further laboratory investigation; OFC is the criterion standard for diagnosis. Mexico and Chile local guidelines seem to follow the same general recommendations as Brazil.

The few Asian guidelines that discuss diagnosis of FA mention taking a good history, a careful physical examination, the use of food diaries, and elimination diets (where appropriate) as the first steps. This is followed by performing SPTs, measurement of specific IgE, and conducting OFC as needed.⁸³⁻⁸⁶ Predictive threshold values for SPTs and food-specific IgE are lacking in Asian populations, and this is an important unmet need because it is unlikely that data accrued from other ethnicities can be used in Asia.⁸⁷ In fact, one of the challenges in the diagnosis of FA in Asian countries is that many countries are resource-limited, both in the number of trained allergy specialists and in access to SPT reagents, laboratory facilities, and food challenge set-ups.^{26,88} Countries that do not currently have a national allergy specialty training and accreditation program should see this as a priority for their health care needs.

There are many diagnostics tests that are used to support diagnosis, although not recommended by current guidelines. However, some of these tests are available only in specialized centers and not available in many countries.⁸⁶ Component-resolved diagnostics (measuring IgE to specific food allergen components) is becoming increasingly used for confirming peanut allergy when tests of sensitization are in the middle range (SPT wheal diameter 3-8 mm or sIgE 0.35-15 kUA/L).⁸⁹ Ara h 2-specific antibody levels used following SPT or whole peanut sIgE in a 2-step algorithm were shown to successfully reduce the need for OFCs by almost two-thirds.⁹⁰ Other examples of component-resolved diagnostics include Ana o 3 for cashew, Gal d 1, 2, 3, and 5 for egg,⁹¹ and Cor a 9 and Cor a 14 for hazelnut allergy.^{92,93} Other specialized and research-based tests include allergen-specific IgG₄ determination and basophil activation tests. Specialized tests such as component-resolved testing, basophil activation measurements, and endoscopy for non-IgE-mediated FA are available only in select countries.⁸⁶

Although recent diagnostic advances (eg, component-resolved diagnostics, basophil activation testing, and allergenic epitope-specific IgE⁹⁴) hold considerable promise for improving accuracy and reliability of FA diagnosis in settings where OFC is impractical, their limited global availability and technical laboratory requirements render them of limited utility in many clinical and epidemiological contexts. Furthermore, the diagnostic validity of these emerging methods remains unknown within many subpopulations, and reference values (eg, sensitivity, specificity, and 95% positive predictive value) are unavailable for many key allergens. Therefore, further work—much of which is ongoing—is still needed to refine these approaches before their more widespread utilization.

PREVENTION OF FA

Guidelines regarding timing of introduction of allergenic foods have undergone dramatic changes as new data have emerged in the last few years. [Table I](#) lists FA prevention guidelines from around the world. [Table II](#) highlights key recommendations from a few international guidelines. Although some earlier studies¹²⁹⁻¹³¹ and guidelines recommended allergen avoidance

during pregnancy and lactation, these were not supported by later studies.¹³² The 2019 American Academy of Pediatrics Clinical Report specifically states that there is a lack of evidence to support deliberate maternal exclusion of high-risk allergens during pregnancy and while breast-feeding for the purposes of preventing allergic diseases, including FA.⁹⁸ There is now consensus among current guidelines, which predominantly recommend that women should consume a healthy diet in accordance with dietary recommendations for the general population and do not recommend eliminating or increasing the consumption of potentially allergenic foods during pregnancy or breast-feeding as a strategy for preventing the development or clinical course of FA.

Similarly, early guidelines recommended dietary avoidance or delayed introduction of allergenic foods in infants to prevent FAs. It was hypothesized that increased permeability of the immature infant gut would increase sensitization on allergen ingestion.¹³³ However, a number of subsequent studies showed either no benefits of allergen avoidance or benefits of early allergen consumption.¹³⁴ The 2015 Learning Early About Peanut Allergy¹³⁵ was a large trial in infants at high risk of allergy, and it demonstrated that early introduction of peanut was significantly associated with reduced risk of peanut allergy, and that peanut allergy was 5 times more likely in children who avoided peanuts. The 2016 Enquiring About Tolerance¹³⁶ study compared the effect of early introduction of the 6 most common childhood food allergens (cow's milk, hen's egg, peanut, sesame, cod fish, and wheat) after exclusive breast-feeding and showed the benefits with cooked egg. Subsequent to the publication of these 2 pivotal studies that reported that early introduction of egg and peanut was associated with reduced risk of egg and peanut allergy, FA prevention guidelines were reevaluated, which resulted in many guidelines reversing previous recommendations of allergen avoidance, and instead recommending early introduction of allergenic foods. A number of guidelines now recommend introduction of common food allergens between age 4 and 6 months^{57,98,136}; however, some guidelines recommend not delaying the introduction⁸⁶ of allergen or introduction during the first year of life.¹²² A few guidelines recommend screening before allergen ingestion in high-risk infants.

In 2017, the National Institute of Allergy and Infectious Diseases-sponsored expert panel reversed guidelines and recommended early introduction of peanut for infants who are deemed at risk of developing peanut allergy by virtue of their early-onset hen's egg allergy and/or eczema.¹¹⁴ In 2019, the American Academy of Pediatrics issued a clinical report concluding that there is no evidence that delaying the introduction of allergenic foods, including peanuts, eggs, and fish, beyond age 4 to 6 months prevents FA and indicated that there is now evidence that early introduction of peanuts may prevent peanut allergy.⁹⁸ In 2021, a consensus approach to the primary prevention of FA was published, endorsed by the American Academy of Allergy, Asthma & Immunology (AAAAI), the American College of Allergy, Asthma and Immunology, and the Canadian Society for Allergy and Clinical Immunology.⁶ Among all infants irrespective of risk, it recommends introduction of cooked egg and peanut at around age 6 months, but not before 4 months of life, at home when the infant is developmentally ready. For other allergens, it recommends not deliberately delaying introduction because there are no data showing harm with introduction of other allergens in the first year of life (but also

TABLE II. Comparison of international FA prevention guidelines

Topic	International FA guidelines			
	ASCIA 2017 ¹²⁷	NIAID 2017 ¹¹⁴	Commission on Toxicity (COT) UK 2018 ^{120,128}	APAPARI 2018 ⁸⁷
Foods of relevance	All foods	PN	PN	All foods
BF	BF: at least 6 mo and for as long as mother and infant wish to continue		EBF for around the first 6 mo of life	Continue BF up to 2 y
High-risk definition	Infants with severe eczema and/or egg allergy	Infants with AD and/or HE FA	Infants with a history of early-onset AD or suspected FA	Infants with severe eczema
Pregnant/BF mother	Healthy diet Excluding any foods (including allergenic) not recommended Up to 3 serves of oily fish p/w No recommendation on probiotics			
Introduction of solid foods	All infants: when infant is ready: around 6 mo, but not before 4 mo		CF: introduce in an age-appropriate form from around 6 mo	Healthy infants: complementary foods at 6 mo
Introduction of allergens	All infants should be given allergenic solid foods including PN, cooked HE, dairy, and wheat in first year—includes HR HR: Good evidence: regular PN intake <12 mo can reduce PN allergy. Mod evidence: cooked HE <8 mo (family history of allergy), reduce developing HE allergy	Different PN introduction schedules depending on risk: between 4 and 6 mo in infants with severe AD and/or HE allergy; around 6 mo in infants with mild to moderate AD; family and cultural feeding practices should be followed in infants with no AD or FA	GR: PN and HE need NOT be differentiated from other CF Exclusion of PN and HE beyond 6-12 mo may increase risk of FA to these foods HR may wish to seek medical advice before introducing PN and HE	High-risk infants with family history of allergy: introduction of allergenic foods should not be delayed. High risk with severe eczema: SPT and/or OFC to PN and egg may be required. Introduction of allergenic foods should not be delayed
Continued intake		PN protein to be regularly consumed per week should be approximately 6-7 g over 3 or more feedings	Once introduced, PN and HE should be part of the infant's usual diet. If initial exposure is not continued, this may increase the risk of sensitization and FA	
Formula	Hydrolyzed (partially and extensively) infant formula is not recommended for prevention of allergic disease. If BF is not possible, use std cow's milk formula		Does not support the use of hydrolyzed cow's milk formulas, either exclusively or partially hydrolyzed, to influence the risk of developing allergic or autoimmune disease	
Other				

ACAAI, American College of Asthma, Allergy and Immunology; AD, atopic dermatitis; APAPARI, Asia Pacific Association of Pediatric Allergy, Respiriology & Immunology; BF, breast-feeding; BSACI, British Society of Allergy and Clinical Immunology; CF, complementary foods; COT UK, Committee on Toxicity United Kingdom; CSACI, Canadian Society of Allergy and Clinical Immunology; EBF, exclusive breast-feeding; F&V, fruit and vegetables; GR, general risk; HE, hen's egg; HR, high risk; JPGFA, Japanese Pediatric Guideline for Food Allergy; NIAID, National Institute of Allergy and Infectious Diseases; PN, peanut.

International FA guidelines				
BSACI 2018 ¹¹⁹	American Academy of Pediatrics 2019 ⁹⁸	Consensus statement: AAAAI, ACAAI, Canadian Society of Allergy and Clinical Immunology 2020 ⁶	European Academy of Allergy and Clinical Immunology (2004, ⁹⁵ 2008, ⁹⁹ 2014, ¹⁰⁷ 2020 ⁵⁷)	JPGFA 2020 ⁸⁶
PN and egg	All foods	All foods	All foods	All foods
EBF for around the first 6 mo of life and continue BF for first year	No conclusions can be made about the role of BF in either preventing or delaying the onset of specific FAs	EBF: recommended for all mothers, no association between EBF and prevention of FA	EBF for around the first 6 mo of life and continue BF for first year	No evidence that BF prevents FA
Infants with a history of early-onset AD or suspected FA		Consider infants: - infants with severe AD (highest risk) - mild to moderate AD, family history of atopy in either/both parents, or infants with 1 known FA potentially at some increased risk of developing FA (or an additional FA). FA often develops in infants who have no identifiable risk factors. No evidence to clearly support the younger sibling of a PN-allergic child is at increased risk of developing PN allergy, though such infants may be at risk of developing PN allergy secondary to delayed introduction of PN	(2004, ⁹⁵ 2008, ⁹⁹ 2014, ¹⁰⁷ 2020 ⁵⁷) Infants with 1 or 2 parents and/or older siblings with a history of atopic diseases 2020 Limited to PN Populations with high prevalence of PN allergy	Infants with eczema
Omega-3 fatty acids may help reduce the risk of atopic dermatitis in early life	Lack of evidence to support maternal dietary restrictions either during pregnancy or during lactation	Do not recommend maternal exclusion of common allergens We do support any food or supplement	Against: avoiding food allergens (2004, ⁹⁵ 2008, ⁹⁹ 2014, ¹⁰⁷ 2020 ⁵⁷)	Avoidance of allergens not recommended
From 4 mo on (HR) From around 6 mo when developmentally ready, not before 4 mo (GR)			All infants: some families choose to start complementary feeding between 4 and 6 mo (2004, ⁹⁵ 2008, ⁹⁹ 2014, ¹⁰⁷ 2020 ⁵⁷)	All infants: around 5-6 mo (for all)
GR: PN and HE as part of the family diet HR: introduce HE and PN when ready; from 4 mo on; HE before PN	All infants: no evidence that delaying the introduction of allergenic foods, including PN, HE, and fish, beyond 4-6 mo prevents atopic disease. There is now evidence that early introduction of PN may prevent PN allergy (based on NIAID guidelines)	Introduce PN and cooked HE to all infants starting around 6 mo, not before 4 mo Do not delay introduction of other allergenic CF (cow's milk, soy, wheat, tree nuts, sesame, fish, shellfish) around 6 mo, not before 4 mo	All infants: introduce PN and well-cooked HE as part of CF from 4 to 6 mo of life (2020 ⁵⁷)	All infants/high risk: delayed introduction of FA not recommended
Once successfully introduced, continue to give the allergen food to baby regularly as part of their usual diet (eg, at least once per week)				
Soya-based or hydrolyzed infant formula is not recommended for prevention of allergic disease. If BF is not possible, use std cow's milk formula	Lack of evidence that partially or extensively hydrolyzed formula prevents atopic disease even HR	Do not use any HFs for prevention of FA or sensitization Infants should be fed a diverse diet, because this may prevent FA. No recommendation on prebiotics and probiotics	There is no recommendation for or against hydrolyzed infant formulas Against: soy protein formula in the first 6 mo of life No recommendation for or against: vitamin supplements, fish oil, prebiotics, probiotics, or synbiotics in pregnancy when BF or in infancy; altering the duration of EBF; hydrolyzed infant formulas	Insufficient evidence on the usefulness of hydrolyzed milk in preventing the onset of FAs No recommendation on probiotics during pregnancy

no data showing specific benefit). Before allergen introduction, such as peanut, preemptive screening is not required, but the guidance notes that the decision to screen is preference-sensitive. If screening is done, it is recommended that an OFC follows any positive result. Once allergens are introduced, ongoing regular ingestion for maintenance of tolerance is recommended (although noting insufficient evidence to support a precise dose and frequency of ingestion).

The Australian guidelines were updated in 2017 to actively recommend the introduction of allergenic foods including cooked egg and peanut in the first year of life.¹²⁷ Studies have shown high levels of adoption of this advice, with more than 80% of infants now introduced to peanut in the first year of life, with a median age of introduction of 6 months.¹³⁷ Whether the successful adoption of earlier introduction of allergenic foods has had an impact on the prevalence of FA and anaphylaxis is currently being investigated.

The EAACI updated its guidelines on the prevention of onset of development of FA in young children in 2020.⁵⁷ Previous guidelines were published in 2004,⁹⁵ 2008,⁹⁹ and 2014.¹⁰⁷ In 2019, the European Food Safety Authority Panel on Nutrition, Novel Foods and Food Allergens revised its Scientific Opinion of 2009 on the appropriate age for introduction of complementary feeding of infants.¹³⁸ A systematic review was carried out using the Grading of Recommendations, Assessment, Development, and Evaluation (GRADE) approach⁵⁶ and based on this, guidelines were written using the Appraisal Guidelines for Research and Evaluation (AGREE II) framework. The key changes from the 2014 guidelines were for recommendation of introduction of peanut and well-cooked egg (between age 4 and 6 months) into the infant's diet as part of complementary feeding. The recommendations for egg and peanut were made for all infants regardless of risk status for the development of FAs, although it was noted that the high-risk infants would likely benefit more.

There is no consistent evidence that breast-feeding is effective for the prevention of allergic disease. However, for optimal health of the infant, the World Health Organization⁵⁷ and the EAACI¹³⁹ recommend exclusive breast-feeding for a minimum of 6 months and 4 months, respectively. Some guidelines recommend continuing breast-feeding alongside solid food introduction for up to 1^{57,115} or 2 years⁸⁷ or as long as possible.¹²² There is lack of strong evidence that partially or extensively hydrolyzed formula prevents atopic disease in infants and children, even in those at high risk for allergic disease, and discrepancy among guidelines exist, with major guidelines either not recommending their use,^{57,115} or having no recommendations,⁸⁶ or recommending their use (for high-risk infants).¹²³ The EAACI guidelines also recommend against the use of regular cow's milk formula in the first week of life.⁵⁷

Recommendations for the use of dietary supplements vary with guidelines. Currently available evidence does not indicate that probiotic supplementation reduces the risk of developing allergy in children. However, considering all critical outcomes in this context, the World Allergy Organization Guidelines for Allergic Disease Prevention guideline panel determined that there is a likely net benefit from using probiotics resulting primarily from prevention of eczema, but not FA.¹¹³ Other guidelines that recommend their use include Singapore^{84,103} and Hong Kong.^{111,112} Current EAACI and ASCIA guidelines do not recommend their use.^{57,122}

Diet diversity during infancy has been hypothesized to prevent FA, likely by exposing the gut microbiota to diverse foods, increased intake of fiber and nutrients, and promoting development of immune tolerance.¹⁴⁰ In 2014, the Protection Against Allergy Study in Rural Environments was the first to investigate the association between the introduction of several foods during the first year of life and the development of asthma, allergic rhinitis, FA, or atopic sensitization. Their hypothesis was that exposure in early life to diverse food antigens could increase maturation of the mucosal immune system and induce tolerance. The study evaluated diet diversity using 4 methods: minimum diet diversity (World Health Organization classification), food diversity, fruit and vegetable diversity, and food allergen diversity. Children were assessed for FA at 1, 2, 3, and 10 years. The study found that increased infant diet diversity, as measured by all 4 different methods, decreased the likelihood of developing FA. The study showed that the introduction of each additional food at age 6 and 12 months reduced by 10.8% and 33.2%, respectively, the odds of developing FA over the first 10 years of life.¹⁴¹ A systematic review of diet diversity in infancy and childhood suggested that diet diversity in infancy may be associated with reduced allergy outcomes (including FA), but additional studies are required to define more clearly the role of diet diversity and diet patterns, while clearly adjusting for appropriate confounders.¹⁴²

Current evidence suggests that allergic sensitization occurs through an impaired skin barrier, while consumption of these foods at an early age may result in tolerance (dual allergen exposure hypothesis). The loss of skin integrity is thought to enable penetration of allergens, pollutants, and microbes, leading to immune dysfunction and initiation of the allergic cascade and eventual formation of IgE. The immune dysfunction is thought to further exacerbate the impaired skin barrier, forming a vicious cycle. Research into skin emollient to protect the skin and prevent development of a proinflammatory atopic state, which could lead to the development of AD and subsequently, FA, is an active area of research.^{143,144}

TREATMENTS

The most recent 2014 Practice Parameter from the Joint Task Force of the American Academy of Allergy, Asthma and Immunology and the American College of Allergy, Asthma and Immunology recommends absolute avoidance of the allergenic food(s) and preparedness for treatment with an intramuscular injection of epinephrine in case of reactions after inadvertent exposures in those with FA.¹⁴⁵ Nevertheless, oral immunotherapy (OIT) using off-the-shelf food products and allowing ingestion of the allergenic food is widespread in private practices in the United States. In January of 2020, the United States Food and Drug Administration approved Palforzia, which is the first drug ever approved for FA. Palforzia is an OIT drug for peanut allergy, in children aged 4 to 17 years, which mitigates the risk of allergic reactions, including anaphylaxis, due to accidental peanut ingestion. The drug consists of a characterized peanut powder. Treatment includes ingestion of the contents of a series of capsules with increasing doses of peanut protein, leading to a daily maintenance dose sachet of 300 mg of peanut protein. Patients on Palforzia must continue with strict peanut avoidance and continue daily dosing to maintain protection. The Canadian Society for Allergy and Clinical Immunology has taken a notably

different approach and recommends OIT with off-the-shelf foods as a treatment to achieve desensitization to allergenic foods in toddlers through adolescents, and possibly in adults.¹⁴⁶ Uncontrolled asthma is an absolute contraindication to OIT, as is pregnancy in the Canadian Society for Allergy and Clinical Immunology guidelines.

The EAACI guidelines on IgE-mediated FA management, published in 2014, differentiates acute management from long-term strategies.¹³⁹ Appropriate dietary avoidance remains the cornerstone. Education is highlighted as a key point, including diet and emergency kit/management plan utilizations. In 2018, following a systematic review on food immunotherapy, the EAACI concluded that the major benefit of OIT is to increase the threshold of reaction, particularly for cow's milk, hen's egg, and peanut in children.^{147,148} Concerns about safety were addressed, and careful monitoring for local and systemic anaphylactic reactions was recommended. As a consequence, the European guidelines restrict immunotherapy to research centers or clinical centers with substantial experience. Up to now, they do not support biologicals, such as omalizumab, alone or in association with immunotherapy. In December 2020, the European Medicines Agency also approved Palforzia for treating children aged between 4 and 17 years with peanut allergy.¹⁴⁹

The current ASCIA guidelines on the treatment of FA are to adhere to strict allergen avoidance. At this time, there are no Therapeutic Goods Administration–approved FA treatments in Australasia. Few allergists in Australasia currently perform OIT due to concerns regarding high rates of reaction including anaphylaxis. However, it has now been observed that over the long-term, rates decrease.^{150,151} The ASCIA Position Paper on OIT for FA¹⁵² currently recommends against the use of OIT for the treatment of FAs, and highlights the need for additional studies to establish safety, tolerability, cost-effectiveness, QOL, and long-term outcomes. Several phase 2 studies are underway in Australia to evaluate whether the use of an adjuvant alongside OIT can improve efficacy and/or safety of OIT for treatment of peanut, egg, and milk allergies.¹⁵³⁻¹⁵⁶

Guidelines regarding the treatment of FA in Latin America are scarce, and all of them focused on the pediatric population.¹⁵⁷⁻¹⁶¹ Mexico and Chile have developed government-issued recommendations, the former for all groups of food allergens, while the latter specifically for cow's milk protein allergy. Other groups of experts from Argentina, Brazil, Colombia, and Latin America in general have published consensus regarding different allergens. Most of these documents are based on international guidelines and do not necessarily reflect the characteristics of the Latin American population.¹⁵⁹⁻¹⁶¹ The common ground between most of these guidelines is the relevance attributed to the restriction of FAs both in the patient and in the lactating mother, the strategies that practitioners might use to avoid malnutrition, and the importance of treating anaphylactic episodes with intramuscular epinephrine.¹⁵⁷⁻¹⁶¹ The use of OIT and mAbs is supported by the Colombian, Brazilian, and Argentinian guidelines¹⁵⁹⁻¹⁶¹; however, the latter also mentions the usefulness of some other types of immunotherapy, including sublingual and epicutaneous routes.¹⁵⁹ In addition, the Brazilian consensus recommends the induction of oral tolerance through baked foods, due to the good response that most patients present to baked allergens, especially milk and egg.¹⁶⁰ Because of the

lack of regional references, it is imperative for Latin American associations to develop local studies to deliver focused recommendations for this specific population.

In Asia, the standard approach for the management of FA recommended by all guidelines is the avoidance of causative foods.^{86,103,162-165} The more current guidelines mention OIT.^{86,163,164} The Japanese guideline advocates minimum avoidance of causative foods, with recommendation for patients to take lower amounts of foods or hypoallergenic forms such as heated or cooked.⁸⁶ In Malaysia and Hong Kong, cow's milk allergy guidelines while recognizing the promise of OIT do not recommend it in routine clinical practice.^{163,164} In Japan, the FA guideline describes OIT as an investigational intervention in patients with immediate-type FA in whom natural early acquisition of tolerance is not expected.⁸⁶ Furthermore, OIT must be approved by the relevant ethics committee and administered only with informed consent. Protocols for OIT vary between institutes and countries including the practice of low-dose OIT.¹⁶⁶

Although an OIT drug for peanut allergy has been approved and OIT for other allergens show promise in clinical trials, there are practical considerations that still need to be addressed. OIT generally requires multiple clinic visits and treatment over many months to years to reach desensitization. The fear of adverse reactions from OFCs during screening is an additional barrier to initiation of therapy. For these reasons, there is a push toward "real-life" OIT studies where patients are enrolled without OFCs. In these patients, enrollment is based on history and positive serum sIgE or SPT results. To reduce the risk of allergic reaction, the maintenance dose reached after stepwise incremental increases in allergen was lowered. A study by Vickery et al¹⁶⁷ demonstrated that low-dose peanut OIT (300 mg/d) achieved similar sustained desensitization to those achieved by high-dose (3000 mg/d) treatment. For those with multiple FAs (about 45% of individuals with FA), OIT is even more burdensome, both in study duration and in frequency of clinical visits. To address this, research into simultaneous introduction of multiple foods during OIT is being investigated. Some multi-OIT protocols used in clinical trials pretreat patients with a short course of anti-IgE antibody, omalizumab, before the start of OIT. In a randomized placebo-controlled study by Andorf et al,¹⁶⁸ allergens used in multi-OIT were 1 or more of the following: cashew, walnut, hazelnut, almond, sesame, cow's milk, hen's egg, peanut, soy, or wheat. Multifood OIT with adjunctive omalizumab has been found not only to be safe and effective but also has been shown to rapidly decrease time to desensitization to multiple foods. Further research on safety and efficacy as well as optimization of omalizumab and multifood OIT dose and frequency is ongoing (NCT03881696). Research into combination of multi-OIT with a novel biologic, dupilumab, approved for atopic dermatitis is also ongoing (NCT03679676).

A major gap is whether OIT provides real-world benefit in terms of reducing reactions and improving QOL. A recent meta-analysis showed that in patients with FA, OIT is associated with an improvement in health-related QOL.¹⁶⁹ However, well-designed and long-term health-related QOL studies are necessary to ascertain sustained benefits of OIT.¹⁶⁹ Although tremendous strides in FA therapeutics have been taken in the past decade, much ground remains to be covered to meaningfully reduce the population-level burden of FA. For example, awareness of and

access to OIT remains low among the affected patient population—at least in the United States where these data were recently obtained from a nationally representative sample of patients with FA and caregivers.¹⁷⁰ Furthermore, concerns about the treatment burden of OIT have led to numerous innovations designed to reduce the risk of anaphylaxis and more rapidly induce desensitization, including the use of biologic, probiotic, and Chinese herbal adjunct therapies.¹⁷¹ However, despite their promise, the effectiveness of these novel approaches in improving patient outcomes remains largely unknown. Finally, despite the current lack of Food and Drug Administration/European Medicines Agency–approved therapeutic options for patients with allergies besides peanut, numerous other immunotherapeutic approaches are under investigation—including sublingual and epicutaneous immunotherapies, vaccines, and biologic monotherapies.¹⁷² The ultimate goal of prevention and treatment strategies should be to create a personalized approach of shared decision making, taking into account not only the individual's FA characteristics (severity, number, and type of allergenic foods) but also their comorbidities and impact on QOL.

FUTURE RESEARCH/CONCLUSIONS

FA guidelines across the globe have differences, some of which reflect regional, cultural, and societal preferences, whereas others are associated with organizational aspects of local health delivery systems. However, a common theme behind guidelines is the lack of clear scientific evidence for some important matters and, consequently, reliance on expert opinion; this underlines the need for future clinical research, particularly in the diagnosis, management, and prevention of FA.

Although clinical research in the field needs to reflect local needs, coordination with regard to methodology and analytic approaches will help overcome some of the differences that currently exist. This can be achieved by the formation of a global consortium of FA researchers who can design common clinical research protocols that could then be applied in various parts of the globe with modifications based on local reality. Specific major research areas requiring intensive and coordinated efforts in the next few years include the following: (1) Efforts to replace OFC as the criterion standard for diagnosis using simple algorithms that combine standardized clinical tools (questionnaires and laboratory testing); these will most likely need to be specific for each of the major allergenic food. Such efforts will also contribute to improving our ability to conduct accurate, reliable epidemiologic studies and track the incidence and prevalence of FA around the world, (2) standardization of OIT or development of other allergen immunotherapy approaches (eg, other forms of allergen exposure and allergen plus an immunomodulator) aiming at improved safety and ease of use and conferring the ability to switch from immunotherapy to natural food consumption, and (3) better identification of risk factors for development of FA such that women who are pregnant, planning to become pregnant, or lactating can be provided clear information about their and their infants' diet, including optimal timing and quantities of specific food introduction before switching to *ad lib* food consumption. In the long run, efforts should be directed toward development of improved methods (including genetic tests) for determining infants at high risk, development of nonallergen treatments, and improvements in dietary and environmental approaches to improve barrier function and microbiome structure and function

across all epithelial barriers (gut, skin, nose, ear, lung) to prevent FA.

What do we know?

- The incidence/prevalence of FA is rising in certain regions of the world.
- The presentations are getting more complex, more severe, stretching across many different immune mechanisms, and development of tolerance is delayed.
- Food allergen identification is often based on surrogate markers of sensitization rather than food challenge.
- Eliminating allergenic food consumption during pregnancy or breast-feeding for preventing sensitization is not recommended.
- Early intervention with active food allergen introduction and increased diet diversity might prevent FA.
- Early intervention with proper emollient care might prevent sensitization to foods.

What is still unknown?

- The spectrum of food allergens is not identified in some geographic locations.
- The impact of immigration, ethnicity, and genetic variability on the clinical expression of FA needs to be evaluated.
- There are still many dietary and environmental factors and their specific role in epithelial integrity and microbiome structure and function that need further clarification.
- The level of specific IgE that positively predicts clinical reactivity is not identified for many food allergens.
- In addition to sIgE/SPT, new biomarkers predicting FA phenotype are needed.
- Evidence-based diagnostic criteria for non-IgE-mediated FA are needed.
- Global acceptability of OIT and multi-OIT needs to be further assessed.
- Long-term efficacy of OIT needs to be determined.
- New therapies to try to treat food allergies are under investigation.
- Early biomarkers of treatment response are needed.
- Molecular mechanisms of food allergen tolerance and desensitization to be efficiently used in the clinical setting.

REFERENCES

1. Sigurdardottir ST, Jonasson K, Clausen M, Lilja Bjornsdottir K, Sigurdardottir SE, Roberts G, et al. Prevalence and early-life risk factors of school-age allergic multimorbidity: the EuroPrevall-IFAAM birth cohort. *Allergy* 2021;76:2855-65.
2. Burks AW, Tang M, Sicherer S, Muraro A, Eigenmann PA, Ebisawa M, et al. ICON: food allergy. *J Allergy Clin Immunol* 2012;129:906-20.
3. Warren CM, Brewer AG, Grobman B, Jiang J, Gupta RS. Racial/ethnic differences in food allergy. *Immunol Allergy Clin North Am* 2021;41:189-203.
4. Tang R, Wang ZX, Ji CM, Leung PSC, Woo E, Chang C, et al. Regional differences in food allergies. *Clin Rev Allergy Immunol* 2019;57:98-110.
5. Fiocchi A, Assa'ad A, Bahna S. Adverse Reactions to Foods Committee, American College of Allergy, Asthma and Immunology. Food allergy and the introduction of solid foods to infants: a consensus document. *Adverse Reactions to Foods*

- Committee, American College of Allergy, Asthma and Immunology. *Ann Allergy Asthma Immunol* 2006;97:10-20, quiz 1, 77.
6. Fleischer DM, Chan ES, Venter C, Spergel JM, Abrams EM, Stukus D, et al. A consensus approach to the primary prevention of food allergy through nutrition: guidance from the American Academy of Allergy, Asthma, and Immunology; American College of Allergy, Asthma, and Immunology; and the Canadian Society for Allergy and Clinical Immunology. *J Allergy Clin Immunol Pract* 2021;9:22-43.e4.
 7. Krawiec M, Fisher HR, Du Toit G, Bahnsen HT, Lack G. Overview of oral tolerance induction for prevention of food allergy—where are we now? *Allergy* 2021;76:2684-98.
 8. Allen JW, Edwards N, Koplin JJ, Netting MJ, Allen KJ. International compliance with WHO infant feeding guidelines—is the confusion cause for concern? *Allergy* 2020;75:673-4.
 9. Dougherty JA, Wagner JD, Stanton MC. Peanut allergen powder-dnfp: a novel oral immunotherapy to mitigate peanut allergy. *Ann Pharmacother* 2021;55:344-53.
 10. Ballmer-Weber BK. Allergic reactions to food proteins. *Int J Vitam Nutr Res* 2011;81:173-80.
 11. Warren CM, Jiang J, Gupta RS. Epidemiology and burden of food allergy. *Curr Allergy Asthma Rep* 2020;20:6.
 12. Sudharson S, Kalic T, Hafner C, Breiteneder H. Newly defined allergens in the WHO/IUIS Allergen Nomenclature Database during 01/2019-03/2021 [published online ahead of print July 26, 2021]. *Allergy*. <https://doi.org/10.1111/all.15021>.
 13. Loh W, Tang MLK. The epidemiology of food allergy in the global context. *Int J Environ Res Public Health* 2018;15:2043.
 14. Prescott SL, Pawankar R, Allen KJ, Campbell DE, Sinn J, Fiocchi A, et al. A global survey of changing patterns of food allergy burden in children. *World Allergy Organ J* 2013;6:21.
 15. Hilvo M. Maternal elimination diet and symptoms of cow's milk allergy in breastfed infants. *JAMA Pediatr* 2021;175:425-6.
 16. Osborne NJ, Koplin JJ, Martin PE, Gurrin LC, Lowe AJ, Matheson MC, et al. Prevalence of challenge-proven IgE-mediated food allergy using population-based sampling and predetermined challenge criteria in infants. *J Allergy Clin Immunol* 2011;127:668-76.e1-2.
 17. Peters RL, Koplin JJ, Gurrin LC, Dharmage SC, Wake M, Ponsonby AL, et al. The prevalence of food allergy and other allergic diseases in early childhood in a population-based study: HealthNuts age 4-year follow-up. *J Allergy Clin Immunol* 2017;140:145-53.e8.
 18. Sasaki M, Koplin JJ, Dharmage SC, Field MJ, Sawyer SM, McWilliam V, et al. Prevalence of clinic-defined food allergy in early adolescence: the SchoolNuts study. *J Allergy Clin Immunol* 2018;141:391-8.e4.
 19. Gupta RS, Warren CM, Smith BM, Blumenstock JA, Jiang J, Davis MM, et al. The public health impact of parent-reported childhood food allergies in the United States. *Pediatrics* 2018;142:e20181235.
 20. Gupta RS, Warren CM, Smith BM, Jiang J, Blumenstock JA, Davis MM, et al. Prevalence and severity of food allergies among US adults. *JAMA Netw Open* 2019;2:e185630.
 21. Keil T, McBride D, Grimshaw K, Niggemann B, Xepapadaki P, Zannikos K, et al. The multinational birth cohort of EuroPrevall: background, aims and methods. *Allergy* 2010;65:482-90.
 22. Kummeling I, Mills EN, Clausen M, Dubakiene R, Perez CF, Fernandez-Rivas M, et al. The EuroPrevall surveys on the prevalence of food allergies in children and adults: background and study methodology. *Allergy* 2009;64:1493-7.
 23. Xepapadaki P, Fiocchi A, Grabenhenrich L, Roberts G, Grimshaw KE, Fiandor A, et al. Incidence and natural history of hen's egg allergy in the first 2 years of life—the EuroPrevall birth cohort study. *Allergy* 2016;71:350-7.
 24. Schoemaker AA, Sprickelman AB, Grimshaw KE, Roberts G, Grabenhenrich L, Rosenfeld L, et al. Incidence and natural history of challenge-proven cow's milk allergy in European children—EuroPrevall birth cohort. *Allergy* 2015;70:963-72.
 25. Baseggio Conrado A, Patel N, Turner PJ. Global patterns in anaphylaxis due to specific foods: a systematic review [published online ahead of print May 1, 2021]. *J Allergy Clin Immunol*. <https://doi.org/10.1016/j.jaci.2021.03.048>.
 26. Hossny E, Ebisawa M, El-Gamal Y, Arasi S, Dahdah L, El-Owaidy R, et al. Challenges of managing food allergy in the developing world. *World Allergy Organ J* 2019;12:100089.
 27. Li J, Ogorodova LM, Mahesh PA, Wang MH, Fedorova OS, Leung TF, et al. Comparative study of food allergies in children from China, India, and Russia: the EuroPrevall-INCO surveys. *J Allergy Clin Immunol Pract* 2020;8:1349-58.e16.
 28. Tham EH, Leung ASY, Pacharn P, Lee S, Ebisawa M, Lee BW, et al. Anaphylaxis—lessons learnt when East meets West. *Pediatr Allergy Immunol* 2019;30:681-8.
 29. Dai H, Wang F, Wang L, Wan J, Xiang Q, Zhang H, et al. An epidemiological investigation of food allergy among children aged 3 to 6 in an urban area of Wenzhou, China. *BMC Pediatr* 2020;20:220.
 30. Basera W, Botha M, Gray CL, Lunjani N, Watkins AS, Venter C, et al. The South African Food Sensitisation and Food Allergy population-based study of IgE-mediated food allergy: validity, safety, and acceptability. *Ann Allergy Asthma Immunol* 2015;115:113-9.
 31. Botha M, Basera W, Facey-Thomas HE, Gaunt B, Gray CL, Ramjith J, et al. Rural and urban food allergy prevalence from the South African Food Allergy (SAFFA) study. *J Allergy Clin Immunol* 2019;143:662-8.e2.
 32. Kung SJ, Fenemore B, Potter PC. Anaphylaxis to Mopane worms (*Imbrasia belina*). *Ann Allergy Asthma Immunol* 2011;106:538-40.
 33. Kung SJ, Steenhoff AP, Gray C. Food allergy in Africa: myth or reality? *Clin Rev Allergy Immunol* 2014;46:241-9.
 34. Mabelane T, Basera W, Botha M, Thomas HF, Ramjith J, Levin ME. Predictive values of alpha-gal IgE levels and alpha-gal IgE: total IgE ratio and oral food challenge-proven meat allergy in a population with a high prevalence of reported red meat allergy. *Pediatr Allergy Immunol* 2018;29:841-9.
 35. Mendoza-Quispe D, Alvarez L, Galván CA. [Overestimation of food allergies reported by parents in a Peruvian allergy, asthma, and immunology center]. *Rev Peru Med Exp Salud Publica* 2018;35:708-10.
 36. Naspič CK, Sole D, Jacob CA, Sarinho E, Soares FJ, Dantas V, et al. Sensitization to inhaled and food allergens in Brazilian atopic children by in vitro total and specific IgE assay. Allergy Project—PROAL [in Portuguese]. *J Pediatr (Rio J)* 2004;80:203-10.
 37. Martínez J, Mendez C, Talesnik E, Campos E, Viviani P, Sanchez I. Skin prick test of immediate hypersensitivity in a selected Chilean pediatric population sample [in Spanish]. *Rev Med Chil* 2005;133:195-201.
 38. Guimarães TC, Gonçalves LC, Silva RM, Segundo GR. Prevalence of parent-reported food allergy in infants and preschoolers in Brazil. *Allergol Immunopathol (Madr)* 2015;43:424-5.
 39. Marrugo J, Hernández L, Villalba V. Prevalence of self-reported food allergy in Cartagena (Colombia) population. *Allergol Immunopathol (Madr)* 2008;36:320-4.
 40. Soto-Quiros M, Gutierrez I, Calvo N, Araya C, Karlberg J, Hanson LA, et al. Allergen sensitization of asthmatic and nonasthmatic schoolchildren in Costa Rica. *Allergy* 1998;53:1141-7.
 41. Gonzales-González VA, Díaz AM, Fernández K, Rivera MF. Prevalence of food allergens sensitization and food allergies in a group of allergic Honduran children. *Allergy Asthma Clin Immunol* 2018;14:23.
 42. Avila Castañón L, Pérez López J, del Río Navarro BE, Rosas Vargas MA, Lerma Ortiz L, Sierra Monge JJ. Hypersensitivity detected by skin tests to food in allergic patients in the Hospital Infantil de México Federico Gómez [in Spanish]. *Rev Alerg Mex* 2002;49:74-9.
 43. Navarro D, López C, Villalobos D, Flores L, Rodríguez R, González L, et al. Gastrointestinal symptoms in children with atopic dermatitis and its association with food allergy. *Arch Venez Pueric Pediatr* 2004;67:181-6.
 44. Leal F, García E, Fiorentino S, Gomez E. Sensibilización alérgica a los alimentos tropicales usados en la ablatación. *Actualizaciones Pediatr* 1991;1:3-5, 14.
 45. Sánchez-Borges M. Etiology and clinical picture of anaphylaxis in ambulatory patients from Caracas, Venezuela. *J Investig Allergol Clin Immunol* 2010;20:623-4.
 46. Mariscal-Castro J, Bedolla-Barajas M, Bedolla-Pulido TR, Domínguez-García MV, Bedolla-Pulido TI, Morales-Romero J, et al. The prevalence of oral allergy syndrome: regarding a new classification [in Spanish]. *Rev Alerg Mex* 2020;67:214-23.
 47. García MB, Gomez-Samper AF, García E, Peñaranda A. Delayed urticaria or anaphylaxis after consumption of red meat with evidence of alpha-gal sensitization. *BMJ Case Rep* 2020;13:e236923.
 48. Okubo Y, Nochioka K, Testa MA. Nationwide survey of hospitalization due to pediatric food-induced anaphylaxis in the United States. *Pediatr Emerg Care* 2019;35:769-73.
 49. Mullins RJ, Dear KB, Tang ML. Time trends in Australian hospital anaphylaxis admissions in 1998-1999 to 2011-2012. *J Allergy Clin Immunol* 2015;136:367-75.
 50. Marenholz I, Grosche S, Kalb B, Rüschenhoff F, Blümchen K, Schlags R, et al. Genome-wide association study identifies the SERPINB gene cluster as a susceptibility locus for food allergy. *Nat Commun* 2017;8:1056.
 51. Winters A, Bahnsen HT, Ruczinski I, Boorgula MP, Malley C, Keramati AR, et al. The MALT1 locus and peanut avoidance in the risk for peanut allergy. *J Allergy Clin Immunol* 2019;143:2326-9.

52. Alfano DN, Klei LR, Klei HB, Trotta M, Gough PJ, Foley KP, et al. MALT1 protease plays a dual role in the allergic response by acting in both mast cells and endothelial cells. *J Immunol* 2020;204:2337-48.
53. van Ginkel CD, Pettersson ME, Dubois AEJ, Koppelman GH. Association of STAT6 gene variants with food allergy diagnosed by double-blind placebo-controlled food challenges. *Allergy* 2018;73:1337-41.
54. Koplin JJ, Peters RL, Ponsonby AL, Gurrin LC, Hill D, Tang ML, et al. Increased risk of peanut allergy in infants of Asian-born parents compared to those of Australian-born parents. *Allergy* 2014;69:1639-47.
55. Sahiner UM, Layhadi JA, Golebski K, Istvan Komlosi Z, Peng Y, Sekerel B, et al. Innate lymphoid cells: the missing part of a puzzle in food allergy. *Allergy* 2021;76:2002-16.
56. de Silva D, Halken S, Singh C, Muraro A, Angier E, Arasi S, et al. Preventing food allergy in infancy and childhood: systematic review of randomised controlled trials. *Pediatr Allergy Immunol* 2020;31:813-26.
57. Halken S, Muraro A, de Silva D, Khaleva E, Angier E, Arasi S, et al. EAAACI guideline: preventing the development of food allergy in infants and young children (2020 update). *Pediatr Allergy Immunol* 2021;32:843-58.
58. Katz Y, Rajuan N, Goldberg MR, Eisenberg E, Heyman E, Cohen A, et al. Early exposure to cow's milk protein is protective against IgE-mediated cow's milk protein allergy. *J Allergy Clin Immunol* 2010;126:77-82.e1.
59. Lucas JS, Grimshaw KE, Collins K, Warner JO, Hourihane JO. Kiwi fruit is a significant allergen and is associated with differing patterns of reactivity in children and adults. *Clin Exp Allergy* 2004;34:1115-21.
60. Feehley T, Plunkett CH, Bao R, Choi Hong SM, Cullen E, Belda-Ferre P, et al. Healthy infants harbor intestinal bacteria that protect against food allergy. *Nat Med* 2019;25:448-53.
61. Folletti I, Siracusa A, Paolucci G. Update on asthma and cleaning agents. *Curr Opin Allergy Clin Immunol* 2017;17:90-5.
62. Xian M, Wawrzyniak P, Ruckert B, Duan S, Meng Y, Sokolowska M, et al. Anionic surfactants and commercial detergents decrease tight junction barrier integrity in human keratinocytes. *J Allergy Clin Immunol* 2016;138:890-3.e9.
63. Wang M, Tan G, Eljaszewicz A, Meng Y, Wawrzyniak P, Acharya S, et al. Laundry detergents and detergent residue after rinsing directly disrupt tight junction barrier integrity in human bronchial epithelial cells. *J Allergy Clin Immunol* 2019;143:1892-903.
64. Xavier RJ, Podolsky DK. Unravelling the pathogenesis of inflammatory bowel disease. *Nature* 2007;448:427-34.
65. Caraballo JC, Yshii C, Westphal W, Moninger T, Comellas AP. Ambient particulate matter affects occludin distribution and increases alveolar transepithelial electrical conductance. *Respirology* 2011;16:340-9.
66. Vita AA, Roysse EA, Pullen NA. Nanoparticles and danger signals: oral delivery vehicles as potential disruptors of intestinal barrier homeostasis. *J Leukoc Biol* 2019;106:95-103.
67. Altunbulakli C, Reiger M, Neumann AU, Garzorz-Stark N, Fleming M, Huel-puesch C, et al. Relations between epidermal barrier dysregulation and *Staphylococcus* species-dominated microbiome dysbiosis in patients with atopic dermatitis. *J Allergy Clin Immunol* 2018;142:1643-7.e12.
68. Xian M, Wang K, Lou H, Wang Y, Zhang L, Wang C, et al. Particulate matter 2.5 causes deficiency in barrier integrity in human nasal epithelial cells. *Allergy Asthma Immunol Res* 2020;12:56-71.
69. Michaudel C, Mackowiak C, Maillat I, Fauconnier L, Akdis CA, Sokolowska M, et al. Ozone exposure induces respiratory barrier biphasic injury and inflammation controlled by IL-33. *J Allergy Clin Immunol* 2018;142:942-58.
70. Jin Y, Lu L, Tu W, Luo T, Fu Z. Impacts of polystyrene microplastic on the gut barrier, microbiota and metabolism of mice. *Sci Total Environ* 2019;649:308-17.
71. Leino MS, Loxham M, Blume C, Swindle EJ, Jayasekera NP, Dennison PW, et al. Barrier disrupting effects of *Alternaria alternata* extract on bronchial epithelium from asthmatic donors. *PLoS One* 2013;8:e71278.
72. Aghapour M, Raae P, Moghaddam SJ, Hiemstra PS, Heijink IH. Airway epithelial barrier dysfunction in chronic obstructive pulmonary disease: role of cigarette smoke exposure. *Am J Respir Cell Mol Biol* 2018;58:157-69.
73. Akdis CA. Does the epithelial barrier hypothesis explain the rise in allergy, autoimmunity and other chronic conditions? [published online ahead of print April 12, 2021]. *Nat Rev Immunol*. <https://doi.org/10.1038/s41577-021-00538-7>.
74. Barnes PJ. Targeting cytokines to treat asthma and chronic obstructive pulmonary disease. *Nat Rev Immunol* 2018;18:454-66.
75. Akdis M, Aab A, Altunbulakli C, Azkur K, Costa RA, Cramer R, et al. Interleukins (from IL-1 to IL-38), interferons, transforming growth factor beta, and TNF-alpha: receptors, functions, and roles in diseases. *J Allergy Clin Immunol* 2016;138:984-1010.
76. Koplin JJ, Dharmage SC, Ponsonby AL, Tang ML, Lowe AJ, Gurrin LC, et al. Environmental and demographic risk factors for egg allergy in a population-based study of infants. *Allergy* 2012;67:1415-22.
77. Marrs T, Logan K, Craven J, Radulovic S, McLean W, Lack G, et al. Dog ownership at three months of age is associated with protection against food allergy. *Allergy* 2019;74:2212-9.
78. Otsu K, Dreskin SC. Peanut allergy: an evolving clinical challenge. *Discov Med* 2011;12:319-28.
79. Brown-Whitehorn T, Cianferoni A, editors. Food protein induced enterocolitis (FPIES): diagnosis and management. Springer Nature Switzerland AG; 2020.
80. Mulder D, Justinich C. Understanding eosinophilic esophagitis: the cellular and molecular mechanisms of an emerging disease. *Mucosal Immunol* 2021;4:139-47.
81. NIAID-Sponsored Expert Panel, Boyce JA, Assa'ad A, Burks AW, Jones SM, Sampson HA, et al. Guidelines for the diagnosis and management of food allergy in the United States: report of the NIAID-sponsored expert panel. *J Allergy Clin Immunol* 2010;126:S1-58.
82. Position Paper: evidence-based versus non evidence-based allergy tests and treatments. 2021. Available at: <https://www.allergy.org.au/hp/papers/position-paper-evidence-based-versus-non-evidence-based-allergy-tests-and-treatments>. Accessed November 7, 2021.
83. Kansu A, Yuce A, Dalgic B, Sekerel BE, Cullu-Cokugras F, Cokugras H. Consensus statement on diagnosis, treatment and follow-up of cow's milk protein allergy among infants and children in Turkey. *Turk J Pediatr* 2016;58:1-11.
84. Lee BW, Aw MM, Chiang WC, Daniel M, George GM, Goh EN, et al. Academy of Medicine, Singapore-Ministry of Health clinical practice guidelines: management of food allergy. *Singapore Med J* 2010;51:599-607.
85. Food allergy. 2021. Available at: <http://www.apapari.org/resources/journal04.asp>. Accessed November 7, 2021.
86. Ebisawa M, Ito K, Fujisawa T. Committee for Japanese Pediatric Guideline for Food Allergy, The Japanese Society of Pediatric Allergy and Clinical Immunology; Japanese Society of Allergy. Japanese guidelines for food allergy 2020. *Allergol Int* 2020;69:370-86.
87. Tham EH, Shek LP, Van Bever HP, Vichyanond P, Ebisawa M, Wong GW, et al. Early introduction of allergenic foods for the prevention of food allergy from an Asian perspective—An Asia Pacific Association of Pediatric Allergy, Respirology & Immunology (APAPARI) consensus statement. *Pediatr Allergy Immunol* 2018;29:18-27.
88. Lee TH, Leung TF, Wong G, Ho M, Duque JR, Li PH, et al. The unmet provision of allergy services in Hong Kong impairs capability for allergy prevention—implications for the Asia Pacific region. *Asian Pac J Allergy Immunol* 2019;37:1-8.
89. Koplin JJ, Perrett KP, Sampson HA. Diagnosing peanut allergy with fewer oral food challenges. *J Allergy Clin Immunol Pract* 2019;7:375-80.
90. Dang TD, Tang M, Choo S, Licciardi PV, Koplin JJ, Martin PE, et al. Increasing the accuracy of peanut allergy diagnosis by using Ara h 2. *J Allergy Clin Immunol* 2012;129:1056-63.
91. Dang TD, Peters RL, Koplin JJ, Dharmage SC, Gurrin LC, Ponsonby AL, et al. Egg allergen specific IgE diversity predicts resolution of egg allergy in the population cohort HealthNuts. *Allergy* 2019;74:318-26.
92. Buyukiryaki B, Cavkaytar O, Sahiner UM, Yilmaz EA, Yavuz ST, Soyer O, et al. Cor a 14, hazelnut-specific IgE, and SPT as a reliable tool in hazelnut allergy diagnosis in Eastern Mediterranean children. *J Allergy Clin Immunol Pract* 2016;4:265-72.e3.
93. Uotila R, Rontynen P, Pelkonen AS, Voutilainen H, Kaarina Kukkonen A, Makela MJ. For hazelnut allergy, component testing of Cor a 9 and Cor a 14 is relevant also in birch-endemic areas. *Allergy* 2020;75:2977-80.
94. Suarez-Farinas M, Suprun M, Kearney P, Getts R, Grishina G, Hayward C, et al. Accurate and reproducible diagnosis of peanut allergy using epitope mapping [published online ahead of print May 15, 2021]. *Allergy*. <https://doi.org/10.1111/all.14905>.
95. Muraro A, Dreborg S, Halken S, Host A, Niggemann B, Aalberse R, et al. Dietary prevention of allergic diseases in infants and small children, part III: critical review of published peer-reviewed observational and interventional studies and final recommendations. *Pediatr Allergy Immunol* 2004;15:291-307.
96. Prescott SL, Tang ML. Australasian Society of Clinical Immunology and Allergy. The Australasian Society of Clinical Immunology and Allergy position statement: summary of allergy prevention in children. *Med J Aust* 2005;182:464-7.
97. Agostoni C, Decsi T, Fewtrell M, Goulet O, Kolacek S, Koletzko B, et al. Complementary feeding: a commentary by the ESPGHAN Committee on Nutrition. *J Pediatr Gastroenterol Nutr* 2008;46:99-110.
98. Greer FR, Sicherer SH, Burks AW, Committee on Nutrition, Section on Allergy and Immunology. The effects of early nutritional interventions on the

- development of atopic disease in infants and children: the role of maternal dietary restriction, breastfeeding, hydrolyzed formulas, and timing of introduction of allergenic complementary foods. *Pediatrics* 2019;143:e20190281.
99. Host A, Halken S, Muraro A, Dreborg S, Niggemann B, Aalberse R, et al. Dietary prevention of allergic diseases in infants and small children. *Pediatr Allergy Immunol* 2008;19:1-4.
 100. Muche-Borowski C, Kopp M, Reese I, Sitter H, Werfel T, Schafer T. Allergy prevention. *Dtsch Arztebl Int* 2009;106:625-31.
 101. Boyce JA, Assa'ad A, Burks AW, Jones SM, Sampson HA, Wood RA, et al. Guidelines for the diagnosis and management of food allergy in the United States: summary of the NIAID-Sponsored Expert Panel Report. *J Allergy Clin Immunol* 2010;126:1105-18.
 102. Infant Feeding Advice. 2010. Available at: https://www.allergy.org.au/images/stories/hp/info/ASCI_A_Infant_Feeding_Advice_2010.pdf. Accessed November 7, 2021.
 103. AMS-MOH Clinical Practice Guidelines: management of food allergy. 2010. Available at: https://www.moh.gov.sg/docs/librariesprovider4/guidelines/cpg_management-of-food-allergy.pdf. Accessed November 7, 2021.
 104. Pelkonen AS, Kuitunen M, Dunder T, Reijonen T, Valovirta E, Makela MJ, et al. Allergy in children: practical recommendations of the Finnish Allergy Programme 2008-2018 for prevention, diagnosis, and treatment. *Pediatr Allergy Immunol* 2012;23:103-16.
 105. Chan ES, Cummings C; Canadian Paediatric Society, Community Paediatrics Committee and Allergy Section. Dietary exposures and allergy prevention in high-risk infants: a joint statement with the Canadian Society of Allergy and Clinical Immunology. *Paediatr Child Health* 2013;18:545-54.
 106. Schafer T, Bauer CP, Beyer K, Bufe A, Friedrichs F, Gieler U, et al. S3-Guideline on allergy prevention: 2014 update: Guideline of the German Society for Allergology and Clinical Immunology (DGAKI) and the German Society for Pediatric and Adolescent Medicine (DGKJ). *Allergo J Int* 2014;23:186-99.
 107. Muraro A, Halken S, Arshad SH, Beyer K, Dubois AE, Du Toit G, et al. EAACI food allergy and anaphylaxis guidelines. Primary prevention of food allergy. *Allergy* 2014;69:590-601.
 108. Fleischer DM, Sicherer S, Greenhawt M, Campbell D, Chan E, Muraro A, et al. Consensus communication on early peanut introduction and the prevention of peanut allergy in high-risk infants. *Ann Allergy Asthma Immunol* 2015;115:87-90.
 109. Fiocchi A, Pawankar R, Cuello-Garcia C, Ahn K, Al-Hammadi S, Agarwal A, et al. World Allergy Organization-McMaster University Guidelines for Allergic Disease Prevention (GLAD-P): Probiotics. *World Allergy Organ J* 2015;8:4.
 110. di Mauro G, Bernardini R, Barberi S, Capuano A, Correria A, De' Angelis GL, et al. Prevention of food and airway allergy: consensus of the Italian Society of Preventive and Social Paediatrics, the Italian Society of Paediatric Allergy and Immunology, and Italian Society of Pediatrics. *World Allergy Organ J* 2016;9:28.
 111. Chan AW, Chan JK, Tam AY, Leung TF, Lee TH. Guidelines for allergy prevention in Hong Kong. *Hong Kong Med J* 2016;22:279-85.
 112. HKIA position paper on prevention of peanut allergy in high risk infants. 2016. Available at: [http://www.allergy.org.hk/HKIA%20-%20Guidelines%20for%20Prevention%20of%20Peanut%20Allergy%20\(Final\).pdf](http://www.allergy.org.hk/HKIA%20-%20Guidelines%20for%20Prevention%20of%20Peanut%20Allergy%20(Final).pdf). Accessed November 7, 2021.
 113. Cuello-Garcia CA, Fiocchi A, Pawankar R, Yepes-Nunez JJ, Morgano GP, Zhang Y, et al. World Allergy Organization-McMaster University Guidelines for Allergic Disease Prevention (GLAD-P): Prebiotics. *World Allergy Organ J* 2016;9:10.
 114. Togias A, Cooper SF, Acebal ML, Assa'ad A, Baker JR Jr, Beck LA, et al. Addendum guidelines for the prevention of peanut allergy in the United States: report of the National Institute of Allergy and Infectious Diseases-sponsored expert panel. *Ann Allergy Asthma Immunol* 2017;118:166-73.e7.
 115. Stiefel G, Anagnostou K, Boyle RJ, Brathwaite N, Ewan P, Fox AT, et al. BSACI guideline for the diagnosis and management of peanut and tree nut allergy. *Clin Exp Allergy* 2017;47:719-39.
 116. Fewtrell M, Bronsky J, Campoy C, Domellof M, Embleton N, Fidler Mis N, et al. Complementary feeding: a position paper by the European Society for Paediatric Gastroenterology, Hepatology, and Nutrition (ESPGHAN) Committee on Nutrition. *J Pediatr Gastroenterol Nutr* 2017;64:119-32.
 117. Ebisawa M, Ito K, Fujisawa T; Committee for Japanese Pediatric Guideline for Food Allergy, The Japanese Society of Pediatric Allergy and Clinical Immunology, The Japanese Society of Allergology. Japanese guidelines for food allergy 2017. *Allergol Int* 2017;66:248-64.
 118. Recto MST, Genuino MLG, Castor MAR, Casis-Hao RJ, Tamondong-Lachica DR, Sales MIV, et al. Dietary primary prevention of allergic diseases in children: the Philippine guidelines. *Asia Pac Allergy* 2017;7:102-14.
 119. Turner PJ, Feeney M, Meyer R, Perkin MR, Fox AT. Implementing primary prevention of food allergy in infants: new BSACI guidance published. *Clin Exp Allergy* 2018;48:912-5.
 120. Assessing the health benefits and risks of the introduction of peanut and hen's egg into the infant diet before six months of age in the UK. 2018. Available at: <https://cot.food.gov.uk/sites/default/files/jointsacncotallergystatementfinal2.pdf>. Accessed November 7, 2021.
 121. Abrams EM, Hildebrand K, Blair B, Chan ES. Timing of introduction of allergenic solids for infants at high risk. *Paediatr Child Health* 2019;24:56-7.
 122. Infant feeding and allergy prevention. 2020. Available at: https://www.allergy.org.au/images/pcc/ASCI_A_Guidelines_Infant_Feeding_and_Allergy_Prevention_2020.pdf. Accessed November 7, 2021.
 123. Primary prevention of allergy in at-risk infants Consensus Statement. 2019. Available at: [https://www.ams.edu.sg/view-pdf.aspx?file=media%5C5649_fi_819.pdf&ofile=Consensus+Statement+on+Primary+Prevention+of+Allergy+in+At-Risk+Infants+\(FINAL\).pdf](https://www.ams.edu.sg/view-pdf.aspx?file=media%5C5649_fi_819.pdf&ofile=Consensus+Statement+on+Primary+Prevention+of+Allergy+in+At-Risk+Infants+(FINAL).pdf). Accessed November 7, 2021.
 124. Wang H, Ma L, Tan Q, Chen J, Li P, Tang J-P, et al. Chinese Expert Consensus on the Diagnosis and Management of Food Allergy in Children With Atopic Dermatitis#. *Int J Dermatol Venereol* 2020;3:135-41.
 125. Matthai J, Sathiashekar M, Poddar U, Sibal A, Srivastava A, Waikar Y, et al. Guidelines on diagnosis and management of cow's milk protein allergy. *Indian Pediatr* 2020;57:723-9.
 126. Malaysia Allergy Prevention (MAP): guidelines for Healthcare Professionals. Available at: <https://www.allergysai.org/pdf/guideline-map.pdf>. Accessed November 7, 2021.
 127. Netting MJ, Campbell DE, Koplin JJ, Beck KM, McWilliam V, Dharmage SC, et al. An Australian consensus on infant feeding guidelines to prevent food allergy: outcomes from the Australian Infant Feeding Summit. *J Allergy Clin Immunol Pract* 2017;5:1617-24.
 128. Statement on the role of hydrolysed cows' milk formulae in influencing the development of atopic outcomes and autoimmune disease. 2016. Available at: <https://cot.food.gov.uk/sites/default/files/finalstatement-hydrolysedformula.pdf>. Accessed November 7, 2021.
 129. DesRoches A, Infante-Rivard C, Paradis L, Paradis J, Haddad E. Peanut allergy: is maternal transmission of antigens during pregnancy and breastfeeding a risk factor? *J Invest Allergol Clin Immunol* 2010;20:289-94.
 130. Sicherer SH, Wood RA, Stablein D, Lindblad R, Burks AW, Liu AH, et al. Maternal consumption of peanut during pregnancy is associated with peanut sensitization in atopic infants. *J Allergy Clin Immunol* 2010;126:1191-7.
 131. Frank L, Marian A, Visser M, Weinberg E, Potter PC. Exposure to peanuts in utero and in infancy and the development of sensitization to peanut allergens in young children. *Pediatr Allergy Immunol* 1999;10:27-32.
 132. Netting MJ, Middleton PF, Makrides M. Does maternal diet during pregnancy and lactation affect outcomes in offspring? A systematic review of food-based approaches. *Nutrition* 2014;30:1225-41.
 133. Bischoff SC, Mayer JH, Manns MP. Allergy and the gut. *Int Arch Allergy Immunol* 2000;121:270-83.
 134. Fisher HR, Du Toit G, Bahnsen HT, Lack G. The challenges of preventing food allergy: lessons learned from LEAP and EAT. *Ann Allergy Asthma Immunol* 2018;121:313-9.
 135. Du Toit G, Roberts G, Sayre PH, Bahnsen HT, Radulovic S, Santos AF, et al. Randomized trial of peanut consumption in infants at risk for peanut allergy. *N Engl J Med* 2015;372:803-13.
 136. Perkin MR, Logan K, Tseng A, Raji B, Ayis S, Peacock J, et al. Randomized trial of introduction of allergenic foods in breast-fed infants. *N Engl J Med* 2016;374:1733-43.
 137. Soriano VX, Peters RL, Ponsonby AL, Dharmage SC, Perrett KP, Field MJ, et al. Earlier ingestion of peanut after changes to infant feeding guidelines: the Early-Nuts study. *J Allergy Clin Immunol* 2019;144:1327-35.e5.
 138. EFSA Panel on Nutrition, Novel Foods and Food Allergens (NDA), Castenmiller J, de Henauw S, Hirsch-Ernst KI, Kearney J, Knutsen HK, et al. Appropriate age range for introduction of complementary feeding into an infant's diet. *EFSA J* 2019;17:e05780.
 139. Muraro A, Werfel T, Hoffmann-Sommergruber K, Roberts G, Beyer K, Bindslev-Jensen C, et al. EAACI food allergy and anaphylaxis guidelines: diagnosis and management of food allergy. *Allergy* 2014;69:1008-25.
 140. D'Auria E, Peroni DG, Sartorio MUA, Verduci E, Zuccotti GV, Venter C. The role of diet diversity and diet indices on allergy outcomes. *Front Pediatr* 2020;8:545.
 141. Venter C, Maslin K, Holloway JW, Silveira LJ, Fleischer DM, Dean T, et al. Different measures of diet diversity during infancy and the association with childhood food allergy in a UK birth cohort study. *J Allergy Clin Immunol Pract* 2020;8:2017-26.
 142. Venter C, Greenhawt M, Meyer RW, Agostoni C, Reese I, du Toit G, et al. EAACI position paper on diet diversity in pregnancy, infancy and childhood: novel

- concepts and implications for studies in allergy and asthma. *Allergy* 2020;75:497-523.
143. Sweeney A, Sampath V, Nadeau KC. Early intervention of atopic dermatitis as a preventive strategy for progression of food allergy. *Allergy Asthma Clin Immunol* 2021;17:30.
 144. Brough HA, Lanser BJ, Sindher SB, Teng JMC, Leung DYM, Venter C, et al. Early intervention and prevention of allergic diseases [published online ahead of print July 13, 2021]. *Allergy*. <https://doi.org/10.1111/all.15006>.
 145. Sampson HA, Aceves S, Bock SA, James J, Jones S, Lang D, et al. Food allergy: a practice parameter update—2014. *J Allergy Clin Immunol* 2014;134:1016-25.e43.
 146. Begin P, Chan ES, Kim H, Wagner M, Cellier MS, Favron-Godbout C, et al. CSACI guidelines for the ethical, evidence-based and patient-oriented clinical practice of oral immunotherapy in IgE-mediated food allergy. *Allergy Asthma Clin Immunol* 2020;16:20.
 147. Nurmatov U, Dhimi S, Arasi S, Pajno GB, Fernandez-Rivas M, Muraro A, et al. Allergen immunotherapy for IgE-mediated food allergy: a systematic review and meta-analysis. *Allergy* 2017;72:1133-47.
 148. Pajno GB, Fernandez-Rivas M, Arasi S, Roberts G, Akdis CA, Alvaro-Lozano M, et al. EAACI Guidelines on allergen immunotherapy: IgE-mediated food allergy. *Allergy* 2018;73:799-815.
 149. European Commission approves first treatment for peanut allergy. Available at: <https://www.europeanpharmaceuticalreview.com/news/138359/european-commission-approves-first-treatment-for-peanut-allergy/>. Accessed November 7, 2021.
 150. Chinthrajah RS, Purington N, Andorf S, Long A, O'Laughlin KL, Lyu SC, et al. Sustained outcomes in oral immunotherapy for peanut allergy (POISED study): a large, randomised, double-blind, placebo-controlled, phase 2 study. *Lancet* 2019;394:1437-49.
 151. Yu S, Smith A, Hass S, Wu E, Chai X, Zhou J, et al. The risk reduction of accidental exposure-related systemic allergic reactions extrapolated based on food challenge data after 1 year of peanut oral immunotherapy. *Adv Ther* 2021;38:4321-32.
 152. Position paper—oral immunotherapy for food allergy. 2021. Available at: <https://www.allergy.org.au/hp/papers/ascia-oral-immunotherapy-for-food-allergy>. Accessed November 7, 2021.
 153. Randomised, controlled trial evaluating the effectiveness of probiotic and egg oral immunotherapy at inducing desensitisation or tolerance in participants with egg allergy compared with placebo (Probiotic Egg Allergen Oral Immunotherapy for Treatment of Egg Allergy: PEAT study). 2019. Available at: <http://anzctr.org.au/Trial/Registration/TrialReview.aspx?ACTRN=12619000480189>. Accessed November 7, 2021.
 154. Follow-on study of a multicentre randomised, controlled trial (PPOIT-003) evaluating the long-term safety and efficacy of probiotic and peanut oral immunotherapy (PPOIT) compared with oral immunotherapy (OIT) alone and with placebo in peanut allergic individuals. 2019. Available at: <http://www.anzctr.org.au/Trial/Registration/TrialReview.aspx?id=377706>. Accessed November 7, 2021.
 155. An open-label study of probiotic and hen's egg or cow's milk oral immunotherapy (Probiotic and Egg or Milk Oral Immunotherapy: PrEMO study). 2018. Available at: <http://anzctr.org.au/Trial/Registration/TrialReview.aspx?ACTRN=12619000306112>. Accessed November 7, 2021.
 156. Oral peanut immunotherapy with a modified dietary starch adjuvant for treatment of peanut allergy in children aged 10-16 years. 2017. Available at: <http://anzctr.org.au/Trial/Registration/TrialReview.aspx?ACTRN=12617000914369>. Accessed November 7, 2021.
 157. Diagnóstico y tratamiento de la alergia alimentaria en niños. 2011. Available at: <https://cenetec-difusion.com/gpc-sns/?p=1740>. Accessed November 7, 2021.
 158. Guía Clínica Alergia a Proteína de Leche de Vaca. Santiago: Minsal; 2012. Available at: <https://www.minsal.cl/portal/url/item/dd7c4cf4c183c58de040010165016b2a.pdf>. Accessed November 7, 2021.
 159. Comité Nacional de A. Food allergy in children: recommendations for diagnosis and treatment [in Spanish]. *Arch Argent Pediatr* 2018;116:s1-9.
 160. SolÉ D, Silva LR, Cocco RR, Ferreira CT, Sarni RO, Oliveira LC, et al. Consenso Brasileiro sobre Alergia Alimentar: 2018—Parte 2—Diagnóstico, tratamento e prevençã,o. Documento conjunto elaborado pela Sociedade Brasileira de Pediatria e Associaçã,o Brasileira de Alergia e Imunologia. *BJAI* 2018;2:39-82.
 161. Bagés MC, Chinchilla Mejía CF, Ortíz Piedrahita C, Plata García CE, Puello Mendoza EM, Quintero Hernández OJ, et al. Recomendaciones sobre diagnóstico y tratamiento de la alergia a la proteína de la leche de vaca en población pediátrica colombiana. Posición de expertos. *Revista colombiana de Gastroenterología* 2020;35:54-64.
 162. Cow milk protein allergy. Available at: <http://www.thaipediatics.org/Media/media-20161208102750.pdf>. Accessed November 7, 2021.
 163. Guidelines for the management of cow's milk protein allergy in children. 2012. Available at: <https://www.allergymtai.org/pdf/guideline-cows-milk.pdf>. Accessed November 7, 2021.
 164. Guidelines for the diagnosis and management of cow's milk protein allergy (CMPA) in Hong Kong. 2014. Available at: [https://www.allergy.org.hk/HKIA%20-%20Guidelines%20for%20the%20Diagnosis%20and%20Management%20Cow's%20Milk%20Protein%20Allergy%20\(CMPA\)%20in%20Hong%20Kong%20\(Final\).pdf](https://www.allergy.org.hk/HKIA%20-%20Guidelines%20for%20the%20Diagnosis%20and%20Management%20Cow's%20Milk%20Protein%20Allergy%20(CMPA)%20in%20Hong%20Kong%20(Final).pdf). Accessed November 7, 2021.
 165. Munasir Z, Muktiarti D. The management of food allergy in Indonesia. *Asia Pac Allergy* 2013;3:23-8.
 166. Yanagida N, Okada Y, Sato S, Ebisawa M. New approach for food allergy management using low-dose oral food challenges and low-dose oral immunotherapies. *Allergol Int* 2016;65:135-40.
 167. Vickery BP, Berglund JP, Burk CM, Fine JP, Kim EH, Kim JI, et al. Early oral immunotherapy in peanut-allergic preschool children is safe and highly effective. *J Allergy Clin Immunol* 2017;139:173-81.e8.
 168. Andorf S, Purington N, Block WM, Long AJ, Tupa D, Brittain E, et al. Anti-IgE treatment with oral immunotherapy in multifoed allergic participants: a double-blind, randomised, controlled trial. *Lancet Gastroenterol Hepatol* 2018;3:85-94.
 169. Cao S, Borro M, Alonzi S, Sindher S, Nadeau K, Chinthrajah RS. Improvement in health-related quality of life in food-allergic patients: a meta-analysis. *J Allergy Clin Immunol Pract* 2021;9:3705-14.
 170. Warren CM, Roach A, Das R, Casale TB, Vickery BP, Wasserman RL, et al. Oral immunotherapy-related awareness, attitudes, and experiences among a nationally-representative sample of food allergy patients/caregivers [published online ahead of print July 29, 2021]. *J Allergy Clin Immunol Pract*. <https://doi.org/10.1016/j.jaip.2021.07.020>.
 171. Sampath V, Sindher SB, Zhang W, Nadeau KC. New treatment directions in food allergy. *Ann Allergy Asthma Immunol* 2018;120:254-62.
 172. Bohle B, Werfel T. Treatment approaches to food allergy [published online ahead of print June 9, 2021]. *Handb Exp Pharmacol*. https://doi.org/10.1007/164_2021_496.