New insights in systemic lupus erythematosus: From regulatory T cells to CAR-T-cell strategies

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Systemic lupus erythematous is a heterogeneous autoimmune disease with potentially multiorgan damage. Its complex etiopathogenesis involves genetic, environmental, and hormonal factors, leading to a loss of self-tolerance with autoantibody production and immune complex formation. Given the relevance of autoreactive B lymphocytes, several therapeutic approaches have been made targeting these cells. However, the disease remains incurable, reflecting an unmet need for effective strategies. Novel therapeutic concepts have been investigated to provide more specific and sustainable disease modification compared with continued immunosuppression. Autologous hematopoietic stem cell transplantation has already provided the proof-of-concept that immunodepletion can lead to durable

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treatment-free remissions, albeit with significant treatmentrelated toxicity. In the future, chimeric antigen receptor-T-cell therapies, for example, CD19 chimeric antigen receptor-T, may provide a more effective lymphodepletion and with less toxicity than autologous hematopoietic stem cell transplantation. An emerging field is to enhance immune tolerance by exploiting the suppressive capacities of regulatory T cells, which are dysfunctional in patients with systemic lupus erythematous, and thus resemble promising candidates for adoptive cell therapy. Different approaches have been developed in this area, from polyclonal to genetically engineered regulatory T cells. In this article, we discuss the current evidence and future directions of cellular therapies for the treatment of systemic lupus erythematous, including hematopoietic stem cell transplantation and advanced regulatory T-cell-based cellular therapies. (J Allergy Clin Immunol 2022;150:1289-301.)

Key words: Autoimmune diseases, autoimmunity, CAR-T-cell therapy, cell therapy, HSCT

Systemic lupus erythematosus (SLE) is a heterogeneous autoimmune disease, characterized by a loss of self-tolerance with autoantibody production, cellular-tissue infiltration, and end-organ damage that can potentially lead to serious organ complications and even death.¹ It affects women of childbearing age, with a female to male ratio of about 9:1 commonly reported.² The overall disease prevalence ranges from 20 to 150 per 100,000, and both incidence and prevalence of SLE are continuously increasing with substantial geographical variability.³ The mortality of the disease has continuously improved in recent years due to the improved understanding of the pathogenesis and advances in therapy, resulting in a 15-year survival of currently 85% to 95%.^{4,5} Nevertheless, despite these advances, SLE is still associated with a major burden with differential impact on populations, economic costs, and health-related quality of life. Hence, there is an ongoing and unmet need for novel, disease-specific, effective and safe treatment approaches.

PATHOGENIC INSIGHTS INTO SLE

The immunopathogenesis of SLE is complex and involves genetic, environmental, hormonal, epigenetic, and immunoregulatory factors that act either sequentially or simultaneously on the immune system.¹ A clearance defect of apoptotic cells with accumulation of undigested apoptotic remnants may provoke the first hit in the break of self-tolerance by activating normally quiescent

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Abbreviations used					
allo-HSCT:	Allogeneic HSCT				
BAAR:	B-cell-targeting antibody				
CAR:	Chimeric antigen receptor				
CYC:	Cyclophosphamide				
EBMT:	European Society for Blood and Marrow Transplantation				
FVIII:	Factor VIII				
GvHD:	Graft-versus-host disease				
HSCT:	Hematopoietic stem cell transplantation				
MSC:	Mesenchymal stem cell				
SLE:	Systemic lupus erythematosus				
Tconv:	Conventional T				
TCR:	T-cell receptor				
Treg:	Regulatory T				
T1D:	Type 1 diabetes				

autoreactive lymphocytes that, on repeated or chronic stimulation, may escape self-regulation.⁶ Neutrophils, particularly lowdensity granulocytes, seem to perpetuate the complex interplay between innate and adaptive immune responses, by synthesizing increased levels of proinflammatory cytokines and forming neutrophil extracellular traps, which contain immunostimulatory proteins and autoantigens, including double-stranded DNA.⁷ In addition, IFN-I signaling pathways, mimicking sustained antivirus responses, have been related to lupus disease susceptibility, and seem to contribute to the immunopathology by amplifying autoimmune responses,⁸ for example, by driving autoreactive humoral activity,⁹ ultimately resulting in the generation of autoreactive plasma cells and production of antinuclear antibodies.

Role of B cells

B cells contribute to the immunopathogenesis of SLE via multiple mechanisms. As progenitors of plasma cells, they are central in the generation of pathogenic autoantibodies. In addition, they mediate deleterious functions through antigen presentation to T cells, costimulatory functions via the expression of accessory molecules engaging stimulatory receptors on T cells, and the production of cytokines.¹⁰ The marked B-cell hyperactivity in SLE is reflected by the presence of peripheral blood CD19⁺CD20⁻CD27⁺⁺ plasmablasts that correlate with disease activity and serum anti-double-stranded DNA autoantibody titers.^{11,12} Altered B-cell subset distribution in SLE also includes the predominance of IgD⁻CD27⁻ double-negative B cells that express CD95.¹³ Recent studies indicated that this B-cell subset is enriched for CD11c⁺Tbet⁺ memory B cells¹⁴ that have been associated with autoreactivity,¹⁵ as well as CD19^{low} CXCR5⁻CD21⁻ B cells, characteristic for extrafollicular generation.¹⁶ These memory B-cell subsets share functional properties and transcriptomic signatures with plasmablasts, suggesting their contribution to autoantibody production. In addition, B cells obtained from patients with SLE have altered expression profiles of regulatory checkpoint molecules, such as B- and T-lymphocyte attenuator (BTLA),¹⁷ and display intrinsic abnormalities in signal transduction and immunometabolism.¹⁸ Once activated, memory B cells differentiate into plasmablasts that subsequently migrate to the bone marrow or inflamed tissue to become long-lived memory plasma cells. In SLE, pathogenic autoantibodies are secreted from both subsets. Short-lived plasma cells are usually associated

with lupus flares,¹¹ whereas long-lived plasma cells contribute to the disease chronicity by the continuous secretion of autoantibodies.¹⁹

Role of T cells

In addition to B-cell disturbances, T cells seem to be central in lupus pathology. Particularly, expanded populations of CD4⁺ T follicular-helper and T peripheral helper cells facilitate B-cell activation and autoantibody production,^{20,21} whereas cytotoxic T cells promote local inflammatory responses, for example, in lupus nephritis,²² potentially contributing to tissue injury. A common feature of lupus T cells is an upregulation of interferon response genes, as recently identified in peripheral blood²³ and skin-infiltrating T cells by single-cell transcriptomics.²⁴ On a molecular level, a number of mechanisms, including altered expression and/or activity of protein kinases and phosphatases,^{25,26} and transcription factors^{27,28} are involved in the increased generation of effector T-cell phenotypes, increased expression of proinflammatory (IL-17A and IL-23) cytokines, and reduced expression of immuneregulatory cytokines (IL-2).²⁷ As a consequence, deficient IL-2 production contributes to reduced numbers and altered function of regulatory T (Treg) cells in SLE,^{29,30} facilitates the amplification of inflammation through reduced activation-induced cell death, and plays a role in the development of secondary immune deficiency in SLE, such as reduced function of cytotoxic T cells.31,32

THERAPEUTIC STRATEGIES FOR SLE

Despite the era of modern biological and targeted therapies, a cure for SLE still remains elusive and approved treatments aim to provide a disease modification allowing to control symptoms and organ manifestations.³³ According to recent European League Against Rheumatism recommendations,³⁴ the goal of treatment is to achieve remission,³⁵ or where remission cannot be reached, a state of low disease activity.³⁶ Embedded into a treat-to-target concept, these target criteria continuously need to be monitored by validated lupus activity indices and treatment adapted accordingly.³⁴ Important other recommendations include avoidance of disease flares, reduction in steroid use, improvement of healthrelated quality of life, and prevention of accumulating organ damage. Overarching treatment principles indicate that the treatment of patients should be adapted to multiple disease-specific and patient-specific aspects, especially the individual profile of involved organ manifestations, and should be based on a shared decision.37

Current therapeutic concepts for SLE primarily focus on a chronic suppression of autoreactive immune responses, which may be achieved by conventional immunosuppressive or biologic disease-modifying therapies, targeting cellular or soluble components involved in lupus immunopathology. Because of the complexity of the underlying immune dysregulation in SLE, usually a multitarget therapeutic approach is required to control symptoms and halt progression.³⁷ Nevertheless, although providing more specificity and efficacy, these disease-modifying therapies have to be administered continuously or repeatedly, which may be associated with the cumulative risk of infectious complications or comorbidity, and are cost-effective. Alternatively, high-dose immunosuppression followed by hematopoietic stem cell transplantation (HSCT) has emerged as

an effective on/off therapy that has the capacity to provide longterm, treatment-free remissions, indicating that the resetting of the immune system by depleting autoreactive immunologic memory cells with a consecutive reinduction of immunologic selftolerance has curative potential.³⁷

Disease-modifying therapies

In addition to specific recommendations for antiphospholipid syndrome³⁸ and neuropsychiatric SLE,³⁹ the European League Against Rheumatism/American College of Rheumatology (ACR) taskforce recently updated the recommendations for the treatment of SLE³⁴ and lupus nephritis.⁴⁰ In terms of pharmacologic treatment, use of hydroxychloroquine is recommended for all patients at a dose not exceeding 5 mg/kg of body weight. For chronic maintenance treatment, glucocorticoids should be minimized to less than 7.5 mg/d prednisone equivalent and, if possible, withdrawn. If required, immunosuppressive drugs, such as azathioprine, methotrexate, mycophenolate mofetil, or cyclophosphamide (CYC), may be added. For patients with severe organ involvement and insufficient response to either mycophenolate mofetil or CYC, treatment with rituximab is recommended, despite negative results from randomized controlled trials. The only biologic therapy recommended with a grade A recommendation is the B-cell-activating factortargeting mAb belimumab, which demonstrated efficacy in both renal⁴¹ and nonrenal lupus manifestations.⁴² In addition, the IFN- α receptor-targeting antibody anifrolumab was recently approved as add-on biologic therapy for SLE,⁴³ as well as the novel calcineurin inhibitor voclosporin for lupus nephritis.⁴

Novel therapies currently investigated in clinical phase II or III trials with promising results include Janus kinase inhibitors,⁴⁵ mAbs targeting blood dendritic cell antigen 2,⁴⁶ inhibiting the T-B-cell interaction, for example, with anti-CD40L,⁴⁷ as well as novel B-cell–directed mAbs, such as obinutuzumab (anti-CD20 mAb).⁴⁸ In addition, small pilot studies suggested beneficial clinical responses of plasma cell–depleting approaches using the proteasome inhibitor bortezomib,⁴⁹ atacicept (Transmembrane Activator and Calcium-modulator and cytophilin ligand Interactor-Ig),⁵⁰ and the CD38-targeting mAb daratumumab²³ (Fig 1).

Hematopoietic stem cell transplantation

Initially applied as salvage therapy for life-threatening SLE, autologous HSCT has evolved over the past years into a clinical option for patients with insufficient response to available standard therapies.^{51,52} The basic principle of HSCT is to achieve a broad immune depletion, providing an initial "debulking" of the immunologic memory repertoire, including memory T and B lymphocytes as well as plasma cells that are usually refractory to standard immunosuppression but sensitive to conditioning treatment with anti–thymocyte globulin,⁵³ followed by regeneration of the hematopoietic and immune systems.^{54,55}

To date, more than 300 patients have received autologous HSCT specifically for SLE. Between 1996 and 2020, 112 patients with SLE have been reported within the European Society for Blood and Marrow Transplantation (EBMT) registry (Fig 2). Pooled data from the largest 15 single-center experiences and multicenter trials with 339 patients included indicate a disease-free survival of 50% to 66% at 5 years despite discontinuation of immunosuppressive and other targeted disease-modifying

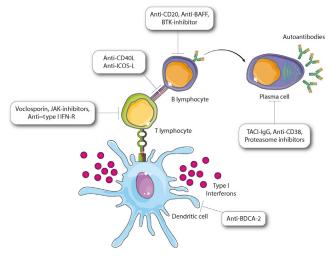
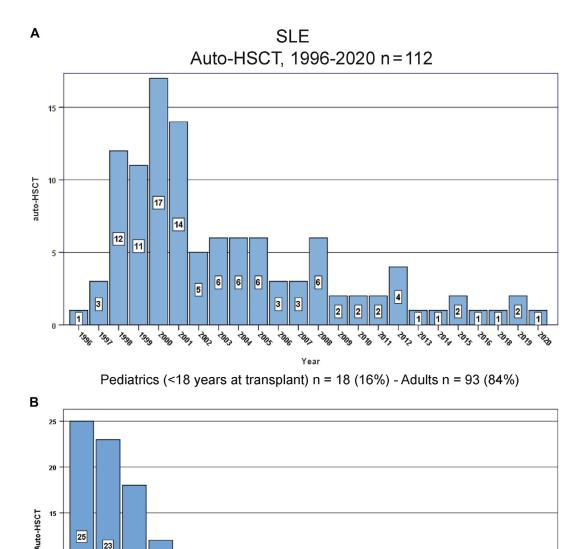


FIG 1. Novel pharmacological targets for SLE. The figure reports some of the targets and their cellular expression of novel drugs currently under evaluation in several clinical trials for SLE. *BAFF*, B-cell-activating factor; *BDCA2*, blood dendritic cell antigen 2; *BTK*, Bruton's tyrosine kinase; *ICOS-L*, inducible T-cell costimulator ligand; *JAK*, Janus kinase; *TACI*, Transmembrane Activator and Calcium-modulator and cytophilin ligand Interactor; *type I IFN-R*, type I interferon receptor.

therapies.⁵⁶ Notably, treatment-related mortality gradually declined from 12% in the first EBMT registry survey in 2004 to less than 5% in most recent reports between 2017 and 2019. Responding patients are usually free of clinical symptoms and may regain seronegativity for antinuclear antibodies, a state referred to as complete clinical and serologic remission "off therapy," which is rarely seen under conventional therapies.³⁵ Compared with continued insufficient or failed chronic immunosuppression, early use of HSCT has also the potential to protect against organ failure and toxicity-related morbidity, such as cardiovascular events, infections, and secondary malignancy, and improve quality of life.⁵⁷ According to previous EBMT recommendations, potential candidates for HSCT would reasonably include those with sustained or relapsed active British Isles Lupus Assessment Group (BILAG) category A remaining steroid dependent after at least 6 months of the best standard therapy, using mycophenolate mofetil or CYC with or without mAbs, with documented evidence of visceral involvement or refractory SLE.^{58,5}

Allogeneic HSCT (allo-HSCT) can be used to restore a dysfunctional immune system, although its wide application has been limited by the risk of graft-versus-host disease (GvHD) and other complications connected to the procedure. Although rare, a retrospective analysis of the EBMT registry published in 2019 reported 5 patients with SLE (2 pediatrics and 3 adults) successfully treated with allo-HSCT⁶⁰ and 3 additional SLE cases in literature achieved a complete remission of autoimmune manifestations after allo-HSCT.⁶¹⁻⁶³ These observations, together with "coincidental" cases of autoimmune diseases in patients undergoing allo-HSCT for hematological malignancies and *in vivo* experiments in mouse models, suggested a potential connection between donor alloreactivity and autoimmune remission, thus suggesting the concept of a putative graft-versus-autoimmunity effect.^{56,64,65}

Collectively, these evidences provide the principle for the use of allo-HSCT as a potential curative approach. Although unlikely to be used widely due to the connected side effects, occasional and



Country FIG 2. Number of HSCTs for SLE. A, The frequency of autologous HSCTs for SLE from 1996 to 2020 included in the EBMT registry. The overall number of pediatric and adult patients is reported. B, The number of HSCTs for SLE by country from 1996 to 2020. Auto-HSCT, Autologous HSCT.

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carefully selected patients may be considered, especially where HSCT risks are lower, potentially with well-matched donors and improvements in allogeneic transplant technique (such as posttransplant CYC and personalized/reduced-toxicity conditioning regimens).56

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Chimeric antigen receptor-T cells

Hungary Spain

Immunotherapy is a promising approach for the depletion of autoreactive cells. Chimeric antigen receptor (CAR)-T cells represent one of the most valuable approaches considering the encouraging results already achieved in other fields.

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CARs are chimeric molecules capable of redirecting the specificity of transduced cells against a target antigen. CARs are composed of 2 major components: the extracellular domain and the intracellular portion. The extracellular domain accounts for the recognition of the target and in most of the cases derives from both the light and the heavy chain of the variable portion of mAb, linked together (single-chain variable fragment).⁶⁶⁻⁶⁸ The intracellular portion mediates the transduction of the signal on antigen binding, and it is composed of 1 or more signaling domain according to the pathway that must be engaged.⁶⁹ These 2 portions are connected together by a linker peptide or spacer.^{70,71}

CARs have been widely studied in the context of cancer. Several commercial and academic autologous CAR-T-cell products targeting B-cell surface antigen CD19 have been approved for B-cell malignancies and multiple myeloma.⁷²

Autoreactive B cells have long been a target for SLE therapy. However, although providing clinical benefit, anti-CD20 mAbs failed to achieve the primary end points in randomized controlled trials. An anti-CD19 CAR-T-cell approach may induce a more robust B-cell depletion compared with the use of B-cell-targeting mAbs (eg, rituximab), especially in tissues in which engineered cells may access more easily. Recently, first data on the use of an anti-CD19 CAR-T-cell strategy in a patient with refractory SLE demonstrated a rapid clinical remission without notable adverse effects, accompanied by sustained depletion of circulating B cells and a rapid disappearance of serum anti-double-stranded DNA antibodies.⁷³ Subsequently, the same group treated an additional 4 patients with SLE with a refractory disease course. Preliminary results on safety and efficacy are encouraging, but data on longterm follow up are warranted.⁷⁴ This first clinical experience builds on preclinical work in mouse models, demonstrating the potential of CAR-T cells to ablate autoantibodies and CD19⁺ B cells, thus improving disease manifestations.⁷⁵

Toxicities, including cytokine-release syndrome and neurologic toxicities, are important side effects associated with CAR-T cells.⁷² These side effects depend on multiple factors, including CAR design, and can be solved using specific strategies.⁷⁶ B-cell aplasia is another well-established consequence of anti-CD19 CAR-T cells, which can last until T cells are functional.⁷⁷ In SLE, the depletion of B cells represents a potential curative approach but might increase the susceptibility to infections.

Collectively, CAR-T cells represent an interesting and promising approach for SLE. Preliminary data are encouraging with convincing efficacy and a favorable safety profile. However, open questions remain, particularly the durability of responses during B-cell repopulation and the identification of an appropriate target population.

TREG CELLS

Treg cells are a specialized branch of CD4⁺ T lymphocytes endowed with suppressive functions, which maintain the immune tolerance and prevent autoimmunity.⁷⁸ They represent a very heterogeneous population, distributed in secondary lymphoid organs and tissues, phenotypically hardly distinguishable from their conventional counterparts. Treg cells constitutionally express high levels of the IL-2 receptor alpha (CD25), and they are highly enriched in the fraction of CD4⁺CD25^{bright} cells. In addition, they are classically identified as CD127^{low} and Forkhead box P3⁺ cells. However, none of these markers is uniquely expressed by Treg cells, but may also be present on activated conventional T (Tconv) cells.⁷⁹ The Forkhead box P3 transcription factor is essential for Treg-cell development, and its absence causes a severe genetic disease with autoimmune manifestations.^{80,81} Other markers have been described in literature, often associated with highly suppressive Treg-cell subsets.^{82,83}

Treg cells are endowed with immune-suppressive functions and are able to control the activation of the immune system. Tregcell lymphocytes can use different strategies to restrain the activity of immune cells, which can be divided into direct mechanisms, based on cell-to-cell contact, and indirect mechanisms, mediated by third-party molecules or cells.⁸⁴ Their immune suppression is broad and involves multiple components of the immune system. Treg cells suppress T and B lymphocytes via direct and indirect mechanisms. Dendritic cells represent another target, which can be reprogrammed toward an immunesuppressive phenotype (tolerogenic dendritic cells), thus limiting the activation of the adaptive immunity.⁸⁵ Treg-cell activity can also involve monocytes, neutrophils, and natural killer cells.⁸⁶

Treg-cell dysfunctionality in SLE

Several studies investigated the role of Treg cells in SLE, obtaining contradictory results. In 2019, a meta-analysis evaluated 18 published studies about Treg cells in lupus, including a total of 628 patients and 601 healthy controls. Despite a great heterogeneity in the methodology, pooled data indicated a reduction in circulating Treg cells in patients with active SLE. In terms of functionality, Li et al²⁹ reported 3 publications in their meta-analysis: 2 studies showed reduced Treg-cell–suppressive functions in SLE, whereas the third one did not identify any significant difference. Pooled data did not reveal any functional difference between patients and controls. Treg-cell selection, the SLE classification criteria that were used, and different experimental methods might explain this great heterogeneity.²⁹

In 2013, Alexander et al⁸⁷ reported a selective and unique expansion of Forkhead box P3⁺ Helios⁺ Treg cells in patients with SLE compared with both healthy subjects and patients with other autoimmune diseases. These cells were highly proliferative and suppressive and displayed an effector memory phenotype and a restricted T-cell receptor (TCR) repertoire, probably representing a compensatory mechanism to control autoreactive cells in tissues.⁸⁷

Collectively, the role of Treg cells in SLE pathogenesis is still controversial. The development of autoimmune manifestations requires the breakdown of the immune tolerance, which is preserved by Treg cells, but up to now it is unclear whether this could be due to a Treg-cell defect in terms of numbers and/or functionality. Heterogeneity among studies did not help to define this aspect. A harmonization of methods might lead to new insights.

TREG-CELL-BASED THERAPIES

The restoration of the immune tolerance with consequent resolution of the inflammatory response against self-antigens is one of the goals of the treatment of autoimmune diseases. Considering their properties, Treg cells represent the ideal candidate for this kind of therapeutic approach. To this end, several strategies have been developed to enhance the Treg-cell response in SLE.^{88,89}

In vivo induction of Treg cells

Immunosuppressive drugs. The first approach to therapeutically use Treg cells relies on the induction of Treg cells directly *in vivo*, by enhancing their activity and/or persistence. Several drugs used for the treatment of autoimmune diseases can act directly or indirectly on Treg-cell numbers and/or functionality. For example, rapamycin/sirolimus increases the number of Treg cells through the inhibition of the mechanistic target of rapamycin (mTOR) pathway. Sirolimus has been used in several clinical trials alone or in combination with other drugs, proving effective in ameliorating the disease activity. In 2020, 1 meta-analysis of 5 studies involving 149 patients with SLE reported a significant reduction in prednisone dose, and a general improvement in the disease from both a clinical and biochemical point of view.⁹⁰

Steroids have a role in inducing Treg cells acting on the miR-342-3p–mTOR complex 2 axis. In particular, Kim et al⁹¹ demonstrated that Treg cells were essential in mediating the anti-inflammatory properties of dexamethasone in a mouse model of autoimmunity and that their absence completely abrogated its therapeutic activity.

Another approach relies on the indirect boost of Treg-cell expansion. This is the case of CYC used after haploidentical HSCT for the prevention of GvHD. In this context, CYC preferentially depletes proliferating conventional effector T cells due to their low expression of aldehyde dehydrogenase, the enzyme required for its degradation, which, in contrast, is expressed at significantly higher levels by Treg cells and hematopoietic stem cells. This approach proved very effective in reducing the risk of GvHD related to haplo-HSCT by favoring Treg-cell reconstitution and restoring the Treg-cell/Tconv-cell ratio.⁹²

Mesenchymal stem cells. Mesenchymal stem cells represent a subset of cells endowed with immunosuppressive properties, capable of inducing Treg cells and in promoting Tconv-cell differentiation toward a $T_H 2$ phenotype *in vitro* and *in vivo* in several preclinical models. For their properties, several clinical trials used mesenchymal stem cells in patients with SLE. Although mesenchymal stem cells have a good safety profile, their administration showed controversial results in controlling disease manifestations and their efficacy has to be assessed yet.⁹³

IL-2 and muteins. Administration of IL-2, an essential cytokine for Treg-cell growth and survival, may enhance immune regulation. Patients with SLE and lupus-prone mouse models showed an impairment of the IL-2 production, potentially explaining the Treg-cell defect in this condition.⁹⁴ The administration of IL-2 in lupus-prone mice improved the number of circulating Treg cells and the disease severity.⁹⁵

In humans, after the first successful open-label trial in SLE,³⁰ several studies indicated a beneficial effect of low-dose IL-2 alone or in combination with the standard of care in improving disease activity, ameliorating disease manifestations, reducing autoantibodies, and complement consumption.⁹⁶ In addition, low-dose IL-2 increased the number of circulating Treg cells and their proliferative capacity, together with a concomitant reduction in follicular-helper T cells, $T_H 17 \text{ CD4}^+$ lymphocytes, and memory B cells.⁹⁷

Low-dose IL-2 administration displayed a good safety profile. The described adverse events were very few, mild, and transient, principally constituted by local reaction at the injection site. In a minority of patients, fever and influenza-like symptoms were reported but they were transient and mild and did not require specific treatments.⁹⁸

Concerns about IL-2 administration regard its poor pharmacodynamics with frequent and long-term administration, with the risk of expanding Tconv cells, worsening autoimmune manifestations. For this reason, several mutated IL-2 variants (called muteins) have been developed to selectively bind the high-affinity IL-2 receptor that is preferentially expressed by Treg cells, thus enhancing their immunomodulatory properties.⁹⁹ Clinical trials are now ongoing to evaluate their efficacy.¹⁰⁰

A list of clinical trials with adoptive Treg-cell therapy with IL-2 is summarized in Table I.

Different compounds proved effective in boosting Treg-cell expansion *in vivo*, albeit with different efficacy. Classic immunosuppressive drugs are very well known, and they typically can control disease flares but with frequent relapses after their discontinuation. Mesenchymal stem cells showed a good safety profile in several clinical trials but a limited efficacy achieving contradictory results. The difference in the administered dose might explain the discrepancies observed between the preclinical studies and clinical trials in humans.

The administration of low-dose IL-2 and muteins represents a potential novel approach for increasing Treg-cell survival and boosting their suppressive properties, potentially helping to restore the immune tolerance. Several clinical trials showed encouraging results in terms of both efficacy and safety profile. Further studies are required to optimize this approach.

Adoptive transfer of polyclonal Treg cells

A more direct approach is based on the *in vitro* expansion of Treg cells and their subsequent reinfusion. In the last 3 decades, the adoptive cell transfer has progressively emerged as a novel strategy for the treatment of several conditions, in particular for cancer and infectious diseases. In the context of autoimmunity, Treg cells represent ideal candidates for an adoptive transfer due to their abilities to control the activation of the immune system.^{88,89}

Treg cells were used in 3 major contexts: autoimmune diseases, treatment of GvHD, and transplant rejection after solid-organ transplantation. For SLE, the only clinical report regarding adoptive Treg-cell therapy dates back to 2019. Dall'Era et al¹⁰⁴ reported a patient with active cutaneous SLE treated with 10⁸ polyclonal Treg cells, which had been expanded *ex vivo* and labeled with deuterium. After 4 weeks, a marked amelioration of the skin lesions was reported despite reduction of labeled Treg cells in peripheral blood. Skin biopsy revealed an enrichment of Treg cells and IL-17–producing CD4⁺ and CD8⁺ cells and a reduction in IFN- γ -secreting T cells.¹⁰⁴

Most clinical trials with adoptive Treg-cell therapy for autoimmune diseases have been performed for the treatment of type 1 diabetes (T1D). In 2014, Marek-Trzonkowska et al¹⁰⁵ conducted a phase I trial in 12 patients with T1D using autologous polyclonal Treg cells, and demonstrated a clinically significant benefit, with 8 subjects achieving clinical remission, with 2 of them becoming insulin-independent. Similar results have been obtained in a subsequent phase I trial in 2015, including 15 patients with T1D treated with polyclonal autologous Treg

TABLE I. Summary of studies with low-dose IL-2 in S	LE
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No. of patients	Groups	Therapy	Outcomes	Reference
1	IL-2 + SOC	1. 5-3.0 MU/d IL-2 s.c. for 5 d/cycle for 4 cycles (9-16 d between cycles)	Increase in circulating Treg cells during cycles. Improvement in SLEDAI, decrease in immune- suppressive therapy, no new manifestations, decrease in anti-DNA antibodies. No SAE, mild local reactions, transient fever	Humrich et al, ¹⁰¹ 2015
5	IL-2 + SOC	1.5 MU/d for 5 d (1 cycle only)	Increase in circulating Treg cells and Treg-cell–associated markers. Increase in Treg-cell/Tcovn-cell proliferation ratio. Increased proliferation of Tconv cells and NK cells but stable counts	von Spee-Mayer et al, ³⁰ 2016
38	IL-2 + SOC	1 MU every other day for 2 wk/cycle for 3 cycles (14 d between cycles)	Increase in Treg cells and Treg-cell suppression. Reduction in Tfh and $T_H 17 \text{ CD4}^+$ Tconv cells. Improvement in SRI-4, amelioration of symptoms, increase in complement, reduction in anti-DNA antibodies and proteinuria. No SAE, mild local reactions, influenza-like symptoms	He et al, ⁹⁶ 2016
12	IL-2 + SOC	0.75-3.0 MU/d IL-2 s.c. for 5 d/cycle for 4 cycles (9-16 d between cycles)	Increase in circulating Treg cells and Treg-cell-associated markers. Increase in Treg-cell/Tcovn-cell proliferation ratio. Increased proliferation of CD8 ⁺ Tconv cells and NK cells. Reduction in Tfh Tconv cells. Reduction in circulating B cells. Improvement in SLEDAI, amelioration of symptoms, increase in complement, reduction in anti-DNA antibodies. No SAE, mild local reactions, influenza-like symptoms. Transient increase in acute-phase proteins	Humrich et al, ⁹⁷ 2019
30	18 patients IL-2 + SOC and 12 patients SOC	1 MU every other day for 2 wk/cycle for 3 cycles (14 d between cycles)	Increase in Treg cells during cycles. Higher remission rate in the IL-2 group at 10 wk, improved renal outcomes. No SAE, mild local reactions, influenza-like symptoms, nausea, diarrhea	Shao et al, ¹⁰² 2019
50	IL-2 + rapamycin	100 WU 3-5 d/mo + rapamycin 0.5 mg every other day for 24 wk	Increase in Treg cells during cycles. Reduced T _H 17 Tconv-cell/ Treg-cell ratio. Improvement in SLEDAI, decrease in immune- suppressive therapy. No SAE	Zhao et al, ¹⁰³ 2019
60	30 patients IL-2 + SOC and 30 patients placebo + SOC	1 MU every other day for 2 wk/cycle for 3 cycles (14 d between cycles)	Increase in Treg cells and NK cells in the IL-2 group. Improvement in SRI-4, amelioration of symptoms, higher remission rate of lupus nephritis, increase in complement, reduction in anti-DNA antibodies in the IL-2 group. No SAE, mild local reactions, influenza-like symptoms	He et al, ⁹⁸ 2020

The table reports the most relevant studies regarding low-dose IL-2 in patients with SLE, describing the number of enrolled patients, groups of treatment, the schedule of administration, a brief description of the outcome, and the bibliographic reference.

NK, Natural killer; SAE, serious adverse events; s.c., subcutaneously; SLEDAI, Systemic Lupus Erythematosus Disease Activity Index; SOC, standard of care; T/h, T follicular-helper.

cells (cell dose: 0.05×10^8 to 26×10^8 cells/patient). Deuterium-prelabeled cells were detectable from the first day of infusion, peaking at 7 to 14 days and reducing by 75% of the initial dose after 3 months, while after 1 year, Treg cells were detectable only in 4 patients.¹⁰⁶ More comprehensive analyses of polyclonal Treg-cell clinical trials have been published elsewhere.^{107,108}

Although the experience in SLE is limited, data on adoptive polyclonal Treg-cell transfer in autoimmunity collectively proved feasibility and safety, albeit with only moderate efficacy. This may be explained by the low level of Treg-cell persistence *in vivo* at least in peripheral blood and the limited number of disease-relevant antigen-specific cells in the final cell product. In fact, as suggested by mouse studies, antigen-specific cells are superior than polyclonal Treg cells in controlling autoimmune responses.^{89,109} The direct expansion of adequate numbers of antigen-specific Treg cells is cumbersome and greatly limits the development of efficacious adoptive Treg-cell therapies. The advent of genome editing techniques allowing the generation of high numbers of antigen-specific Treg cells greatly boosted this field.

Genetically engineered Treg cells

Modern techniques for efficient genome editing allows the generation of engineered T cells. So far, the major field of

application of these approaches is represented by cancer, where genome editing has been largely used to redirect T-cell specificity and increase their potency and/or their safety profile.

However, genome editing can also be used in the context of autoimmunity to increase the number of disease-relevant antigenspecific regulatory cells. In particular, 2 different strategies have been developed: TCR-redirected regulatory cells and CAR-Treg cells.

TCR-redirected Treg cells

The first TCR gene transfer approach was already reported in the nineties by Clay et al,¹¹⁰ who efficiently transduced human T cells with a melanoma-specific TCR using a retroviral vector, and the first study with engineered TCRs in cancer was published in 2006.¹¹¹ Subsequently, several other TCRs have been identified in cancer.⁶⁹

Preclinical studies demonstrated the feasibility and the efficacy of TCR gene transfer for the treatment of autoimmune diseases, especially for T1D. In addition, Brusko et al¹¹² redirected Treg-cell specificity using an antityrosinase TCR, specifically a melanoma antigen. The authors demonstrated the feasibility of the process and the capacity of engineered Treg cells to respond, expand, and exert suppressive capacities in the presence of the cognate antigen.¹¹²

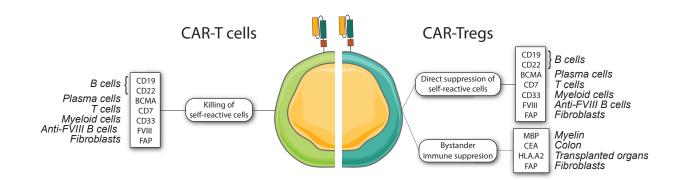


FIG 3. CAR-T-cell strategies comparison. A comparison between conventional CAR-T cells and CAR-Treg cells is reported with a list of the most-studied targets. For each molecule, the associated cellular target is also reported. In addition, for each target, the therapeutic strategy is provided. CAR-T cells can be used to selectively deplete target components relevant for the autoreactive process. CAR-Treg cells can control the autoimmune process by exerting an immune-regulatory activity. Two different strategies can be adopted with CAR-Treg cells: a direct suppression of target cells through a cell-to-cell contact or a broader locoregional immune suppression, especially localized in target organs. *BCMA*, B-cell maturation antigen; *CEA*, carcino-embryonic antigen; *FAP*, fibroblast activation protein; *MBP*, myelin basic protein.

Disease	Target	Starting date	Identifier	Study design	End points
Renal transplantation	HLA-A2	March 2021	NCT04817774	Phase I/IIa multicenter open-label trial	Safety and tolerability Prevention of rejection
Liver transplantation	HLA-A2	January 2022	NCT05234190	Phase I/IIa multicenter open-label trial	Safety and tolerability Prevention of rejection Immunosuppressive withdrawa
R/R CD19 ⁺ B-ALL	CD19	November 2022	NCT05114837	Phase I/IIa single-center open-label trial	Safety and tolerability Antitumor efficacy

The table reports a list of actively recruiting clinical trials with CAR-Treg cells updated to June 2022. The underlying condition, the CAR target, the starting date, the trial identifier, the study design, and the declared primary and secondary end points are reported. Active CAR-Treg-cell clinical trials are found on www.clinicatrials.gov. *R/R CD19*⁺ *B-ALL*, Refractory/relapsing CD19⁺ B-acute lymphoblastic leukemia.

In 2017, 2 publications reported the development of isletspecific TCR-redirected Treg cells. Yeh et al¹¹³ isolated an antiglutamic acid decarboxylase TCR and generated anti-glutamic acid decarboxylase Treg cells, which controlled *in vitro* the proliferation of anti-glutamic acid decarboxylase Tconv cells. Hull et al¹¹⁴ cloned 2 TCRs, specific for tyrosine phosphatase (IA2) and insulin, respectively. TCR-redirected Treg cells showed antigen-specific immune-suppressive capacities *in vitro*.

Kim et al reported the efficacy of TCR-redirected Treg cells in 2 other contexts. Anti–factor VIII (FVIII) Treg cells prevented the immunization against recombinant FVIII in hemophilia.¹¹⁵ In 2018, the same group showed the feasibility of the same approach in the context of central nervous system inflammation. They generated anti-MBP Treg cells, which displayed antigen-specific immune-suppressive capacities *in vitro* and *in vivo* in a mouse model of experimental autoimmune encephalitis.¹¹⁶

CAR-Treg cells

CARs represent a potential solution to redirect Treg-cell specificity, and several preclinical studies proved the efficacy of CAR-Treg cells in autoimmunity. In 2009, Hombach et al¹¹⁷ generated anti–carcinoembryonic antigen CAR-Treg cells and

demonstrated that these cells inhibited carcinoembryonic antigen⁺ tumor rejection by antigen-specific Tconv cells.¹¹⁷ Lee et al¹¹⁸ obtained similar results in a model of CD19⁺ acute Bcell leukemia, showing how anti-CD19 CAR-Treg cells in the tumor site abrogated the activity of conventional anti-CD19 CAR-T cells. In 2020, Imura et al¹¹⁹ demonstrated the efficacy of anti-CD19 CAR-Treg cells in controlling B-cell proliferation and antibody production *in vitro* and in reducing the risk of developing GvHD in a xenograft mouse model.

In 2016, MacDonald et al¹²⁰ generated anti–HLA-A02 CAR-Treg cells. They showed that engineered lymphocytes displayed antigen-specific suppressive properties *in vitro* and prevented xenograft GvHD induced by HLA-A2⁺ PBMCs *in vivo*. In 2017, 2 groups confirmed these results in xenograft mouse models, demonstrating that anti–HLA-A02 CAR-Treg cells blocked the rejection of transplanted human skin.^{121,122}

Scott's group used CAR-Treg cells for the prevention of immune responses against recombinant FVIII in hemophilia. In 2017, Yoon et al¹²³ generated anti-FVIII CAR-Treg cells that blocked FVIII-specific Tconv-cell responses and suppressed the generation of anti-FVIII antibodies both *in vitro* and *in vivo*. Subsequently, in 2018, the same group generated a BAAR, a B-cell–targeting antibody, a novel strategy to selectively block anti-FVIII

B cells, demonstrating how BAAR-Treg cells retained their suppressive capacities and blocked anti-FVIII antibody production both *in vitro* and *in vivo*¹²⁴ (Fig 3). Considering these promising results, first clinical trials involving CAR-Treg cells have been developed, from which one is already enrolling patients, whereas others are expected to start in the next months. An updated list of trials as of June 2022 is depicted in Table II.

Collectively, several preclinical studies demonstrated the feasibility and functionality of engineered Treg cells. TCRs and CARs represent 2 different strategies, both effective in redirecting Treg-cell–suppressive capacities in an antigen-specific manner, although none of them has been specifically used in SLE. TCRs and CARs have some differences, and the choice of one or the other approach depends on the target antigen. TCRs can recognize both extracellular and intracellular antigens but are MHC-restricted. CARs are limited to extracellular molecule but are MHC-independent.⁸⁸ Engineered Treg cells can exert their suppressive functions directly acting on self-reactive cells. In addition, they can exert a locoregional immunosuppression by targeting an antigen expressed in specific tissues to control locally the inflammatory response,⁸⁴ as in the case of anti-MBP engineered Treg cells.¹¹⁶

Given the complex pathogenesis of SLE with autoreactive responses against multiple self-antigens, the identification of a potential target for an adoptive Treg-cell therapy remains a therapeutic challenge. CARs specific for molecules expressed by pathogenic cells, such as CD19 for B cells⁷³ or B-cell maturation antigen for plasma cells,¹²⁵ represent a possible solution. However, these targets are poorly specific for autoreactive cells, being expressed also by their protective counterparts.

BAARs could increase the specificity: these molecules can selectively target self-reactive B lymphocytes and in SLE they might target pathogenic cells.¹²⁴ Finally, for selective applications, locoregional immune suppression might represent a solution, like anti-MBP for neurologic manifestations.¹¹⁶

In conclusion, the identification of a specific target for SLE is still cumbersome. TCRs and CARs are 2 complementary approaches with specific characteristics. Alternative strategies may be used with already available molecules, especially in specific situations. Some suggestions are reported in Fig 3.

CONCLUSIONS AND FUTURE DIRECTIONS

SLE is a complex disease characterized by a breakdown of the immunologic self-tolerance.¹ Autoreactive B and T cells, in conjunction with key players of the innate immune system, play a central role in the disease pathogenesis.^{10,21} Several therapeutic approaches have been attempted so far, many of them with promising results, yielding a therapeutic concept of disease modification that controls symptoms and halts progression but provides no curative potential.³⁷ New insights into SLE pathogenesis have led to the development of more specific drugs, in particular mAbs, which specifically target disease-relevant molecules.⁴¹ Nevertheless, targeted biologic therapies usually require continuous administration to control disease manifestations, and may be associated with the cumulative risk of infections and comorbidity.

A barrier for long-term remissions represents the autoreactive immunologic memory, which is usually formed long before symptoms of the disease occur, and which is mostly refractory to available biologic therapies, particularly autoantibody-secreting memory plasma cells.¹⁹ To control self-reactive cells, in particular memory ones, 2 different approaches have been developed: the first one relies on eliminating autoreactive immune cells (immunoablation), and the second one aims at restoring the immune tolerance (immune regulation).

To obtain the immunoablation of self-reactive cells, autologous HSCT (and rarely allogeneic HSCT) has been used in patients with SLE and already provided the proof-of-concept that longterm remissions can be achieved after resetting the immune system into a self-tolerant state. However, autologous HSCT can be associated with considerable transplant-related mortality and other long-term complications, such as secondary autoimmune diseases.⁵⁷ Furthermore, it remains unclear what particular components of the memory compartment need to be targeted and how deep the lymphocyte lineage depletion will be required to achieve sustainable responses in SLE. In this regard, it will be of interest to follow the results of ongoing CD19 CAR-T-cell therapies in SLE.73,75 Compared with mAbs, this strategy has the advantage of a broader depletion of autoreactive B cells, especially those maintained in inflamed tissues. First results already indicate that this approach provided a therapeutically relevant depletion of B cells along with significant reductions in autoantibodies. However, whether such deep B-cell lineage depletion is sufficient to induce durable responses, or further memory compartments, such as CD19-negative plasma cells or T cells, to be target in addition remains unclear.

Another therapeutic option to control chronic autoimmune responses in SLE is to foster immune regulation, aiming at restoring the immune tolerance. Considering the central role of Treg cells in maintaining self-tolerance, they represent the ideal candidate for such a kind of approach. Several therapeutic approaches are under investigation, either directly "fueling" Treg cells *in vivo*, for example, with IL-2 or muteins, ^{30,96,100} or by *ex vivo* manipulation followed by a subsequent Treg-cell reinfusion (Fig 4). Clinical trials of adoptive Treg-cell therapies demonstrated feasibility and a proof of efficacy. However, the use of polyclonal Treg cells achieved only modest and transient clinical results, probably due to a low number of disease-relevant Treg cells in the cell product or due to a reduced persistence *in vivo*.¹⁰⁹ New strategies rely on the use of engineered regulatory cells, which represent a possible solution to overcome these limitations.⁸⁹

One of the greatest limitations to the use of engineered cells is represented by the cost of the procedure, thus probably limiting its applicability to selected patients.¹²⁶ Specific criteria should be identified to select those patients eligible for an adoptive cell therapy. As for CD19 CAR-T-cell therapies, potential candidates are those with persistent and progressive disease, despite conventional or biologic therapies. In addition, specific biomarkers, for example, those related to IL-2 deficiency, may be helpful for patient selection. Some candidate biomarkers have been reviewed elsewhere.¹²⁷

Beside the costs, manufacturing is another limiting factor for the widespread use of adoptive cell therapy. Nevertheless, the CAR-T experience in cancer revolutionized the field, increasing the availability of both academic and commercial products. Hopefully in the future, availability and costs of manufacturing facilities will gradually improve, especially with the development of decentralized units for the production of the cells.^{128,129}

Safety is another key aspect of adoptive T-cell therapy, especially in the context of autoimmunity, given the chronicity of the condition compared with a life-threatening disease such as

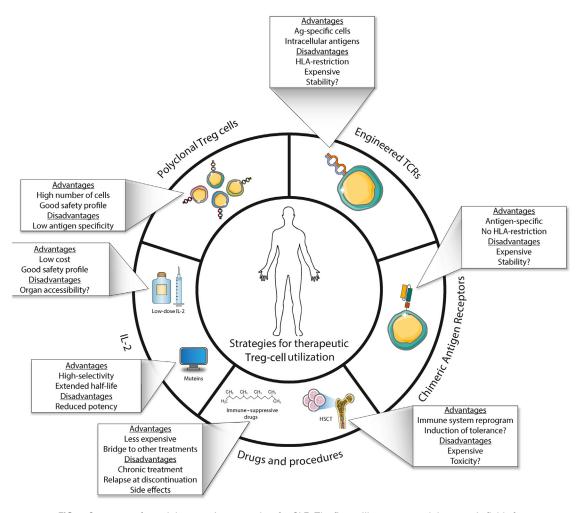


FIG 4. Summary of novel therapeutic approaches for SLE. The figure illustrates novel therapeutic fields for SLE with their advantages and disadvantages. In particular, the use of adoptive Treg-cell therapy is described as either polyclonal or engineered Treg cells. CARs and engineered TCRs can be used to generate engineered Treg cells. To increase Treg-cell activity, low-dose IL-2 and muteins are reported. Finally, the use of immunomodulatory approaches such as immune-suppressive drugs or HSCT is illustrated.

cancer. Conventional CAR-T cells have several well-established side effects in cancer,¹³⁰ whose incidence in SLE has to be addressed in future clinical trials. Regarding Treg cells, several articles demonstrated their instability in chronic inflammatory environments with their reprogramming toward conventional effector cells.¹³¹ Addressing this issue is crucial, especially with CAR-Treg cells, where the reprogramming of the engineered lymphocytes to proinflammatory cells might worsen the underlying condition.

Collectively, SLE treatment is rapidly evolving and new approaches are currently under investigation. Better understanding of the pathologic mechanisms and recent advances in cell manufacturing have generated the development of new, specific therapies to fundamentally modify the cellular interactions and clinical outcomes. The precise role of novel cellular therapies in the future treatment algorithm of SLE remains to be determined. According to current guidelines, SLE therapies should be embedded into a concept of *disease modification* that "requires minimizing disease activity with the fewest treatmentassociated toxicity and slowing or preventing organ damage progression."³³ In this regard, available therapies suppressing immune reactions with or without the use of biologic drugs are sufficient to achieve fundamental treatment goals in most patients with SLE. Therefore, in the near future, application of novel cellular therapies will still be restricted to patients with high risk for mortality or disease progression. According to recent data, such patients are reasonably those "not at target," that is, not achieving lupus low disease activity, who have a significantly increased risk of mortality, accumulating organ damage, and poor quality of life.¹³²

Accumulating data from clinical trials or single experiences with HSCT or CAR T-cell therapies already demonstrated that a vast immune depletion as "on-off" therapy may provide durable responses. These promising data may set a scene for a future ambitious treatment goal of achieving therapy-free long remissions, which could become realistic in the future. Likewise, data from novel Treg-cell-based therapies providing a concept of *immune modulation* are promising and may have a future role in providing clinical remission with low toxicity. In the future, more data are required to evaluate the risk-benefit ratio of individual novel cellular therapies, and most importantly to identify patients who will benefit most from such therapies.

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REFERENCES

- 1. Tsokos GC. Systemic lupus erythematosus. N Engl J Med 2011;365:2110-21.
- Alexander T, Hedrich CM. Systemic lupus erythematosus are children miniature adults? Clin Immunol 2022;234:108907.
- 3. Carter EE, Barr SG, Clarke AE. The global burden of SLE: prevalence, health disparities and socioeconomic impact. Nat Rev Rheumatol 2016;12: 605-20.
- 4. Tektonidou MG, Lewandowski LB, Hu J, Dasgupta A, Ward MM. Survival in adults and children with systemic lupus erythematosus: a systematic review and Bayesian meta-analysis of studies from 1950 to 2016. Ann Rheum Dis 2017;76:2009-16.
- Yen EY, Shaheen M, Woo JMP, Mercer N, Li N, McCurdy DK, et al. 46-year trends in systemic lupus erythematosus mortality in the United States, 1968 to 2013. Ann Intern Med 2017;167:777.
- Wahren-Herlenius M, Dörner T. Immunopathogenic mechanisms of systemic autoimmune disease. Lancet 2013;382:819-31.
- Gupta S, Kaplan MJ. The role of neutrophils and NETosis in autoimmune and renal diseases. Nat Rev Nephrol 2016;12:402-13.
- Crow MK, Ronnblom L. Type I interferons in host defence and inflammatory diseases. Lupus Sci Med 2019;6:e000336.
- Barnas JL, Albrecht J, Meednu N, Alzamareh DF, Baker C, McDavid A, et al. B cell activation and plasma cell differentiation are promoted by IFN-λ in systemic lupus erythematosus. J Immunol 2021;207:2660-72.
- Shen P, Fillatreau S. Antibody-independent functions of B cells: a focus on cytokines. Nat Rev Immunol 2015;15:441-51.
- Odendahl M, Jacobi A, Hansen A, Feist E, Hiepe F, Burmester GR, et al. Disturbed peripheral B lymphocyte homeostasis in systemic lupus erythematosus. J Immunol 2000;165:5970-9.
- Jacobi AM, Mei H, Hoyer BF, Mumtaz IM, Thiele K, Radbruch A, et al. HLA-DRhigh/CD27high plasmablasts indicate active disease in patients with systemic lupus erythematosus. Ann Rheum Dis 2010;69:305-8.
- 13. Jacobi AM, Reiter K, Mackay M, Aranow C, Hiepe F, Radbruch A, et al. Activated memory B cell subsets correlate with disease activity in systemic lupus erythematosus: delineation by expression of CD27, IgD, and CD95. Arthritis Rheum 2008;58:1762-73.
- Rincon-Arevalo H, Wiedemann A, Stefanski A-L, Lettau M, Szelinski F, Fuchs S, et al. Deep phenotyping of CD11c+ B cells in systemic autoimmunity and controls. Front Immunol 2021;12:659.
- Wang S, Wang J, Kumar V, Karnell JL, Naiman B, Gross PS, et al. IL-21 drives expansion and plasma cell differentiation of autoreactive CD11chiT-bet+ B cells in SLE. Nat Commun 2018;9:1758.
- Szelinski F, Stefanski AL, Wiedemann A, Schrezenmeier E, Rincon-Arevalo H, Reiter K, et al. Antigen-experienced CXCR5-CD19 low B cells are plasmablast precursors expanded in SLE [published online ahead of print April 20, 2022]. Arthritis Rheumatol. https://doi.org/10.1101/2021.05.25.21257784
- Wiedemann A, Lettau M, Weißenberg SY, Stefanski A-L, Schrezenmeier E-V, Rincon-Arevalo H, et al. BTLA expression and function are impaired on SLE B cells. Front Immunol 2021;12:1402.
- Zhang X, Yang Y, Jing L, Zhai W, Zhang H, Ma Q, et al. Pyruvate kinase M2 contributes to TLR-mediated inflammation and autoimmunity by promoting Pyk2 activation. Front Immunol 2021;12:1655.
- Hiepe F, Dörner T, Hauser AE, Hoyer BF, Mei H, Radbruch A. Long-lived autoreactive plasma cells drive persistent autoimmune inflammation. Nat Rev Rheumatol 2011;7:170-8.
- Liarski VM, Kaverina N, Chang A, Brandt D, Yanez D, Talasnik L, et al. Cell distance mapping identifies functional T follicular helper cells in inflamed human renal tissue. Sci Transl Med 2014;6:230ra46.
- Makiyama A, Chiba A, Noto D, Murayama G, Yamaji K, Tamura N, et al. Expanded circulating peripheral helper T cells in systemic lupus erythematosus: association with disease activity and B cell differentiation. Rheumatology 2019; 58:1861-9.

- Arazi A, Rao DA, Berthier CC, Davidson A, Liu Y, Hoover PJ, et al. The immune cell landscape in kidneys of patients with lupus nephritis. Nat Immunol 2019;20: 902-14.
- Ostendorf L, Burns M, Durek P, Heinz GA, Heinrich F, Garantziotis P, et al. Targeting CD38 with daratumumab in refractory systemic lupus erythematosus. N Engl J Med 2020;383:1149-55.
- Dunlap GS, Billi AC, Ma F, Maz MP, Tsoi LC, Wasikowski R, et al. Single cell transcriptomics reveals distinct effector profiles of infiltrating T cells in lupus skin and kidney. JCI Insight 2022;7:e156341.
- Suárez-Rojas G, Crispín JC. Dysregulated protein kinase/phosphatase networks in SLE T cells. Clin Immunol 2022;236:108952.
- Tsokos GC. Autoimmunity and organ damage in systemic lupus erythematosus. Nat Immunol 2020;21:605-14.
- Hedrich CM. Mechanistic aspects of epigenetic dysregulation in SLE. Clin Immunol 2018;196:3-11.
- 28. Hofmann SR, Mäbert K, Kapplusch F, Russ S, Northey S, Beresford MW, et al. cAMP response element modulator α induces dual specificity protein phosphatase 4 to promote effector T cells in juvenile-onset lupus. J Immunol 2019;203: 2807-16.
- **29.** Li W, Deng C, Yang H, Wang G. The regulatory T cell in active systemic lupus erythematosus patients: a systemic review and meta-analysis. Front Immunol 2019;10:159.
- 30. von Spee-Mayer C, Siegert E, Abdirama D, Rose A, Klaus A, Alexander T, et al. Low-dose interleukin-2 selectively corrects regulatory T cell defects in patients with systemic lupus erythematosus. Ann Rheum Dis 2016;75:1407-15.
- Hedrich CM, Crispin JC, Tsokos GC. Epigenetic regulation of cytokine expression in systemic lupus erythematosus with special focus on T cells. Autoimmunity 2014;47:234-41.
- 32. Katsuyama E, Suarez-Fueyo A, Bradley SJ, Mizui M, Marin AV, Mulki L, et al. The CD38/NAD/SIRTUIN1/EZH2 axis mitigates cytotoxic CD8 T cell function and identifies patients with SLE prone to infections. Cell Rep 2020;30:112-23.e4.
- 33. van Vollenhoven R, Askanase AD, Bomback AS, Bruce IN, Carroll A, Dall'Era M, et al. Conceptual framework for defining disease modification in systemic lupus erythematosus: a call for formal criteria. Lupus Sci Med 2022;9:e000634.
- 34. Fanouriakis A, Kostopoulou M, Alunno A, Aringer M, Bajema I, Boletis JN, et al. 2019 Update of the EULAR recommendations for the management of systemic lupus erythematosus. Ann Rheum Dis 2019;78:736-45.
- 35. van Vollenhoven R, Voskuyl A, Bertsias G, Aranow C, Aringer M, Arnaud L, et al. A framework for remission in SLE: consensus findings from a large international task force on definitions of remission in SLE (DORIS). Ann Rheum Dis 2017;76:554-61.
- 36. Franklyn K, Lau CS, Navarra SV, Louthrenoo W, Lateef A, Hamijoyo L, et al. Definition and initial validation of a Lupus Low Disease Activity State (LLDAS). Ann Rheum Dis 2016;75:1615-21.
- Dörner T, Furie R. Novel paradigms in systemic lupus erythematosus. Lancet 2019;393:2344-58.
- Limper M, Scirè CA, Talarico R, Amoura Z, Avcin T, Basile M, et al. Antiphospholipid syndrome: state of the art on clinical practice guidelines. RMD Open 2018;4:e000785.
- 39. Pamfil C, Fanouriakis A, Damian L, Rinzis M, Sidiropoulos P, Tsivgoulis G, et al. EULAR recommendations for neuropsychiatric systemic lupus erythematosus vs usual care: results from two European centres. Rheumatology 2015;54:1270-8.
- 40. Fanouriakis A, Kostopoulou M, Cheema K, Anders H-J, Aringer M, Bajema I, et al. 2019 Update of the Joint European League Against Rheumatism and European Renal Association–European Dialysis and Transplant Association (EULAR/ERA–EDTA) recommendations for the management of lupus nephritis. Ann Rheum Dis 2020;79:713-23.
- Furie R, Rovin BH, Houssiau F, Malvar A, Teng YKO, Contreras G, et al. Twoyear, randomized, controlled trial of belimumab in lupus nephritis. N Engl J Med 2020;383:1117-28.
- 42. Furie R, Petri M, Zamani O, Cervera R, Wallace DJ, Tegzová D, et al. A phase III, randomized, placebo-controlled study of belimumab, a monoclonal antibody that inhibits B lymphocyte stimulator, in patients with systemic lupus erythematosus. Arthritis Rheum 2011;63:3918-30.
- Morand EF, Furie R, Tanaka Y, Bruce IN, Askanase AD, Richez C, et al. Trial of anifrolumab in active systemic lupus erythematosus. N Engl J Med 2020;382: 211-21.
- Rovin BH, Teng YKO, Ginzler EM, Arriens C, Caster DJ, Romero-Diaz J, et al. Efficacy and safety of voclosporin versus placebo for lupus nephritis (AURORA 1): a double-blind, randomised, multicentre, placebo-controlled, phase 3 trial. Lancet 2021;397:2070-80.
- 45. Wallace DJ, Furie RA, Tanaka Y, Kalunian KC, Mosca M, Petri MA, et al. Baricitinib for systemic lupus erythematosus: a double-blind, randomised, placebocontrolled, phase 2 trial. Lancet 2018;392:222-31.

- 46. Van Vollenhoven R, Furie R, Kalunian K, Navarra S, Romero-Diaz J, Werth V, et al. POS0698 BIIB059 demonstrated a consistent therapeutic effect on SRI-4 response across subgroups of participants with systemic lupus erythematosus in the LILAC phase 2 study. Ann Rheum Dis 2021;80:597-8.
- 47. Furie RA, Bruce IN, Dörner T, Leon MG, Leszczyński P, Urowitz M, et al. Phase 2, randomized, placebo-controlled trial of dapirolizumab pegol in patients with moderate-to-severe active systemic lupus erythematosus. Rheumatology 2021; 60:5397-407.
- 48. Furie RA, Aroca G, Cascino MD, Garg JP, Rovin BH, Alvarez A, et al. B-cell depletion with obinutuzumab for the treatment of proliferative lupus nephritis: a randomised, double-blind, placebo-controlled trial. Ann Rheum Dis 2022;81:100-7.
- 49. Alexander T, Sarfert R, Klotsche J, Kühl AA, Rubbert-Roth A, Lorenz H-M, et al. The proteasome inhibitior bortezomib depletes plasma cells and ameliorates clinical manifestations of refractory systemic lupus erythematosus. Ann Rheum Dis 2015;74:1474-8.
- Merrill JT, Wallace DJ, Wax S, Kao A, Fraser PA, Chang P, et al. Efficacy and safety of atacicept in patients with systemic lupus erythematosus. Arthritis Rheumatol 2018;70:266-76.
- 51. Duarte RF, Labopin M, Bader P, Basak GW, Bonini C, Chabannon C, et al. Indications for haematopoietic stem cell transplantation for haematological diseases, solid tumours and immune disorders: current practice in Europe, 2019. Bone Marrow Transplant 2019;54:1525-52.
- 52. Alexander T, Greco R, Snowden JA. Hematopoietic stem cell transplantation for autoimmune disease. Annu Rev Med 2021;72:215-28.
- Zand MS, Vo T, Pellegrin T, Felgar R, Liesveld JL, Ifthikharuddin JJ, et al. Apoptosis and complement-mediated lysis of myeloma cells by polyclonal rabbit antithymocyte globulin. Blood 2006;107:2895-903.
- 54. Alexander T, Arnold R, Hiepe F, Radbruch A. Resetting the immune system with immunoablation and autologous haematopoietic stem cell transplantation in autoimmune diseases. Clin Exp Rheumatol 2016;34:53-7.
- Swart JF, Delemarre EM, van Wijk F, Boelens J-J, Kuball J, van Laar JM, et al. Haematopoietic stem cell transplantation for autoimmune diseases. Nat Rev Rheumatol 2017;13:244-56.
- Burt RK, Farge D, Ruiz MA, Saccardi R, Snowden JA. Hematopoietic stem cell transplantation and cellular therapies for autoimmune diseases. Boca Raton: CRC Press; 2021.
- 57. Burt RK, Han X, Gozdziak P, Yaung K, Morgan A, Clendenan AM, et al. Five year follow-up after autologous peripheral blood hematopoietic stem cell transplantation for refractory, chronic, corticosteroid-dependent systemic lupus erythematosus: effect of conditioning regimen on outcome. Bone Marrow Transplant 2018;53:692-700.
- 58. Illei GG, Cervera R, Burt RK, Doria A, Hiepe F, Jayne D, et al. Current state and future directions of autologous hematopoietic stem cell transplantation in systemic lupus erythematosus. Ann Rheum Dis 2011;70:2071-4.
- 59. Snowden JA, Saccardi R, Allez M, Ardizzone S, Arnold R, Cervera R, et al. Haematopoietic SCT in severe autoimmune diseases: updated guidelines of the European Group for Blood and Marrow Transplantation. Bone Marrow Transplant 2012;47:770-90.
- 60. Greco R, Labopin M, Badoglio M, Veys P, Furtado Silva JM, Abinun M, et al. Allogeneic HSCT for autoimmune diseases: a retrospective study from the EBMT ADWP, IEWP, and PDWP Working Parties. Front Immunol 2019;10:1570.
- 61. Khorshid O, Hosing C, Bibawi S, Ueno N, Reveille J, Mayes MD, et al. Nonmyeloablative stem cell transplant in a patient with advanced systemic sclerosis and systemic lupus erythematosus. J Rheumatol 2004;31:2513-6.
- 62. Lu Q, Lu L, Niu X, Guo Y, Parino GR, Liu D. Non-myeloablative allogeneic stem cell transplant in a patient with refractory systemic lupus erythematosus. Bone Marrow Transplant 2006;37:979-81.
- 63. Marmont AM, Bacigalupo A, Gualandi F, Bregante S, van Lint MT, Geroldi S. Systemic lupus erythematosus complicated with thymoma and pure red cell aplasia (PCRA). CR of both complications following thymectomy and allogeneic haematopoietic SCT (HSCT), but persistence of antinuclear antibodies (ANA). Bone Marrow Transplant 2014;49:982-3.
- 64. Marmont AM, Gualandi F, Van Lint MT, Bacigalupo A. Refractory Evans' syndrome treated with allogeneic SCT followed by DLI. Demonstration of a graftversus-autoimmunity effect. Bone Marrow Transplant 2003;31:399-402.
- 65. Van Wijmeersch B, Sprangers B, Rutgeerts O, Lenaerts C, Landuyt W, Waer M, et al. Allogeneic bone marrow transplantation in models of experimental autoimmune encephalomyelitis: evidence for a graft-versus-autoimmunity effect. Biol Blood Marrow Transplant 2007;13:627-37.
- 66. Eshhar Z, Waks T, Gross G, Schindler DG. Specific activation and targeting of cytotoxic lymphocytes through chimeric single chains consisting of antibodybinding domains and the gamma or zeta subunits of the immunoglobulin and T-cell receptors. Proc Natl Acad Sci U S A 1993;90:720-4.

- 67. Schultz LM, Davis KL, Baggott C, Chaudry C, Marcy AC, Mavroukakis S, et al. Phase 1 study of CD19/CD22 bispecific chimeric antigen receptor (CAR) therapy in children and young adults with B cell acute lymphoblastic leukemia (ALL). Blood 2018;132:198.
- Ellebrecht CT, Bhoj VG, Nace A, Choi EJ, Mao X, Cho MJ, et al. Reengineering chimeric antigen receptor T cells for targeted therapy of autoimmune disease. Science 2016;353:179-84.
- Sadelain M, Rivière I, Riddell S. Therapeutic T cell engineering. Nature 2017; 545:423-31.
- Hudecek M, Sommermeyer D, Kosasih PL, Silva-Benedict A, Liu L, Rader C, et al. The nonsignaling extracellular spacer domain of chimeric antigen receptors is decisive for in vivo antitumor activity. Cancer Immunol Res 2015;3:125-35.
- Casucci M, Falcone L, Camisa B, Norelli M, Porcellini S, Stornaiuolo A, et al. Extracellular NGFR spacers allow efficient tracking and enrichment of fully functional CAR-T cells co-expressing a suicide gene. Front Immunol 2018;9:507.
- 72. Hayden PJ, Roddie C, Bader P, Basak GW, Bonig H, Bonini C, et al. Management of adults and children receiving CAR T-cell therapy: 2021 best practice recommendations of the European Society for Blood and Marrow Transplantation (EBMT) and the Joint Accreditation Committee of ISCT and EBMT (JACIE) and the European Haematol. Ann Oncol 2022;33:259-75.
- Mougiakakos D, Krönke G, Völkl S, Kretschmann S, Aigner M, Kharboutli S, et al. CD19-targeted CAR T cells in refractory systemic lupus erythematosus. N Engl J Med 2021;385:567-9.
- 74. Schett G, Boeltz S, Müller F, Kleyer A, Völkl S, Aigner M, et al. OP0279 CAR-T cell treatment of refractory systemic lupus erythematosus—safety and preliminary efficacy data from the first four patients. Ann Rheum Dis 2022;81:185.
- 75. Kansal R, Richardson N, Neeli I, Khawaja S, Chamberlain D, Ghani M, et al. Sustained B cell depletion by CD19-targeted CAR T cells is a highly effective treatment for murine lupus. Sci Transl Med 2019;11:eaav1648.
- 76. Brudno JN, Lam N, Vanasse D, Shen Y, Rose JJ, Rossi J, et al. Safety and feasibility of anti-CD19 CAR T cells with fully human binding domains in patients with B-cell lymphoma. Nat Med 2020;26:270-80.
- Logue JM, Zucchetti E, Bachmeier CA, Krivenko GS, Larson V, Ninh D, et al. Immune reconstitution and associated infections following axicabtagene ciloleucel in relapsed or refractory large B-cell lymphoma. Haematologica 2020;106: 978-86.
- Sakaguchi S, Sakaguchi N, Asano M, Itoh M, Toda M. Immunologic selftolerance maintained by activated T cells expressing IL-2 receptor alpha-chains (CD25). Breakdown of a single mechanism of self-tolerance causes various autoimmune diseases. J Immunol 1995;155:1151-64.
- 79. Fraser H, Safinia N, Grageda N, Thirkell S, Lowe K, Fry LJ, et al. A rapamycinbased GMP-compatible process for the isolation and expansion of regulatory T cells for clinical trials. Mol Ther Methods Clin Dev 2018;8:198-209.
- Lu L, Barbi J, Pan F. The regulation of immune tolerance by FOXP3. Nat Rev Immunol 2017;17:703-17.
- Barzaghi F, Passerini L, Bacchetta R. Immune dysregulation, polyendocrinopathy, enteropathy, x-linked syndrome: a paradigm of immunodeficiency with autoimmunity. Front Immunol 2012;3:211.
- 82. Miyara M, Yoshioka Y, Kitoh A, Shima T, Wing K, Niwa A, et al. Functional delineation and differentiation dynamics of human CD4+ T cells expressing the FoxP3 transcription factor. Immunity 2009;30:899-911.
- Fan H, Ren D, Hou Y. TLR7, a third signal for the robust generation of spontaneous germinal center B cells in systemic lupus erythematosus. Cell Mol Immunol 2018;15:286-8.
- Romano M, Fanelli G, Albany CJ, Giganti G, Lombardi G. Past, present, and future of regulatory T cell therapy in transplantation and autoimmunity. Front Immunol 2019;10:43.
- 85. Onishi Y, Fehervari Z, Yamaguchi T, Sakaguchi S. Foxp3+ natural regulatory T cells preferentially form aggregates on dendritic cells in vitro and actively inhibit their maturation. Proc Natl Acad Sci 2008;105:10113-8.
- Lewkowicz N, Mycko MP, Przygodzka P, Ćwiklińska H, Cichalewska M, Matysiak M, et al. Induction of human IL-10-producing neutrophils by LPS-stimulated Treg cells and IL-10. Mucosal Immunol 2016;9:364-78.
- Alexander T, Sattler A, Templin L, Kohler S, Groß C, Meisel A, et al. Foxp3+ Helios+ regulatory T cells are expanded in active systemic lupus erythematosus. Ann Rheum Dis 2013;72:1549-58.
- Ferreira LMR, Muller YD, Bluestone JA, Tang Q. Next-generation regulatory T cell therapy. Nat Rev Drug Discov 2019;18:749-69.
- Raffin C, Vo LT, Bluestone JA. Treg cell-based therapies: challenges and perspectives. Nat Rev Immunol 2020;20:158-72.
- 90. Peng L, Wu C, Hong R, Sun Y, Qian J, Zhao J, et al. Clinical efficacy and safety of sirolimus in systemic lupus erythematosus: a real-world study and meta-analysis. Ther Adv Musculoskelet Dis 2020;12:1759720X2095333.

- 91. Kim D, Nguyen QT, Lee J, Lee SH, Janocha A, Kim S, et al. Anti-inflammatory roles of glucocorticoids are mediated by Foxp3+ regulatory T cells via a miR-342-dependent mechanism. Immunity 2020;53:581-96.e5.
- Ikegawa S, Matsuoka KI. Harnessing Treg homeostasis to optimize posttransplant immunity: current concepts and future perspectives. Front Immunol 2021;12:1-15.
- 93. Li A, Guo F, Pan Q, Chen S, Chen J, Liu H, et al. Mesenchymal stem cell therapy: hope for patients with systemic lupus erythematosus. Front Immunol 2021;12: 4062.
- 94. Humrich JY, Morbach H, Undeutsch R, Enghard P, Rosenberger S, Weigert O, et al. Homeostatic imbalance of regulatory and effector T cells due to IL-2 deprivation amplifies murine lupus. Proc Natl Acad Sci 2010;107:204-9.
- Mizui M, Koga T, Lieberman LA, Beltran J, Yoshida N, Johnson MC, et al. IL-2 protects lupus-prone mice from multiple end-organ damage by limiting CD4-CD8- IL-17-producing T cells. J Immunol 2014;193:2168-77.
- 96. He J, Zhang X, Wei Y, Sun X, Chen Y, Deng J, et al. Low-dose interleukin-2 treatment selectively modulates CD4(+) T cell subsets in patients with systemic lupus erythematosus. Nat Med 2016;22:991-3.
- 97. Humrich JY, von Spee-Mayer C, Siegert E, Bertolo M, Rose A, Abdirama D, et al. Low-dose interleukin-2 therapy in refractory systemic lupus erythematosus: an investigator-initiated, single-centre phase 1 and 2a clinical trial. Lancet Rheumatol 2019;1:e44-54.
- **98.** He J, Zhang R, Shao M, Zhao X, Miao M, Chen J, et al. Efficacy and safety of low-dose IL-2 in the treatment of systemic lupus erythematosus: a randomised, double-blind, placebo-controlled trial. Ann Rheum Dis 2020;79: 141-9.
- Hernandez R, Põder J, LaPorte KM, Malek TR. Engineering IL-2 for immunotherapy of autoimmunity and cancer [published online ahead of print February 25, 2022]. Nat Rev Immunol. https://doi.org/10.1038/s41577-022-00680-w.
- Mullard A. Restoring IL-2 to its cancer immunotherapy glory. Nat Rev Drug Discov 2021;20:163-5.
- 101. Humrich JY, von Spee-Mayer C, Siegert E, Alexander T, Hiepe F, Radbruch A, et al. Rapid induction of clinical remission by low-dose interleukin-2 in a patient with refractory SLE. Ann Rheum Dis 2015;74:791-2.
- 102. Shao M, He J, Zhang R, Zhang X, Yang Y, Li C, et al. Interleukin-2 deficiency associated with renal impairment in systemic lupus erythematosus. J Interferon Cytokine Res 2019;39:117-24.
- 103. Zhao C, Chu Y, Liang Z, Zhang B, Wang X, Jing X, et al. Low dose of IL-2 combined with rapamycin restores and maintains the long-term balance of Th17/Treg cells in refractory SLE patients. BMC Immunol 2019;20:32.
- 104. Dall'Era M, Pauli ML, Remedios K, Taravati K, Sandova PM, Putnam AL, et al. Adoptive Treg cell therapy in a patient with systemic lupus erythematosus. Arthritis Rheumatol 2019;71:431-40.
- 105. Marek-Trzonkowska N, Myśliwiec M, Dobyszuk A, Grabowska M, Derkowska I, Juścińska J, et al. Therapy of type 1 diabetes with CD4(+)CD25(high) CD127-regulatory T cells prolongs survival of pancreatic islets—results of one year follow-up. Clin Immunol 2014;153:23-30.
- 106. Bluestone JA, Buckner JH, Fitch M, Gitelman SE, Gupta S, Hellerstein MK, et al. Type 1 diabetes immunotherapy using polyclonal regulatory T cells. Sci Transl Med 2015;7:315ra189.
- Pilat N, Sprent J. Treg therapies revisited: tolerance beyond deletion. Front Immunol 2021;11:3663.
- 108. Goswami TK, Singh M, Dhawan M, Mitra S, Bin ET, Rabaan AA, et al. Regulatory T cells (Tregs) and their therapeutic potential against autoimmune disorders – advances and challenges. Hum Vaccin Immunother 2022;18:2035117.
- 109. Tang Q, Henriksen KJ, Bi M, Finger EB, Szot G, Ye J, et al. In vitro-expanded antigen-specific regulatory T cells suppress autoimmune diabetes. J Exp Med 2004;199:1455-65.
- 110. Clay TM, Custer MC, Sachs J, Hwu P, Rosenberg SA, Nishimura MI. Efficient transfer of a tumor antigen-reactive TCR to human peripheral blood lymphocytes confers anti-tumor reactivity. J Immunol 1999;163:507-13.
- 111. Morgan RA, Dudley ME, Wunderlich JR, Hughes MS, Yang JC, Sherry RM, et al. Cancer regression in patients after transfer of genetically engineered lymphocytes. Science 2006;314:126-9.

- 112. Brusko TM, Koya RC, Zhu S, Lee MR, Putnam AL, McClymont SA, et al. Human antigen-specific regulatory T cells generated by T cell receptor gene transfer. PLoS One 2010;5:e11726.
- 113. Yeh W-I, Seay HR, Newby B, Posgai AL, Moniz FB, Michels A, et al. Avidity and bystander suppressive capacity of human regulatory T cells expressing de novo autoreactive T-cell receptors in type 1 diabetes. Front Immunol 2017;8: 1-13.
- 114. Hull CM, Nickolay LE, Estorninho M, Richardson MW, Riley JL, Peakman M, et al. Generation of human islet-specific regulatory T cells by TCR gene transfer. J Autoimmun 2017;79:63-73.
- 115. Kim YC, Zhang A-H, Su Y, Rieder SA, Rossi RJ, Ettinger RA, et al. Engineered antigen-specific human regulatory T cells: immunosuppression of FVIII-specific T- and B-cell responses. Blood 2015;125:1107-15.
- 116. Kim YC, Zhang A-H, Yoon J, Culp WE, Lees JR, Wucherpfennig KW, et al. Engineered MBP-specific human Tregs ameliorate MOG-induced EAE through IL-2-triggered inhibition of effector T cells. J Autoimmun 2018;92:77-86.
- 117. Hombach AA, Kofler D, Rappl G, Abken H. Redirecting human CD4+CD25+ regulatory T cells from the peripheral blood with pre-defined target specificity. Gene Ther 2009;16:1088-96.
- 118. Lee JC, Hayman E, Pegram HJ, Santos E, Heller G, Sadelain M, et al. In vivo inhibition of human CD19-targeted effector T cells by natural T regulatory cells in a xenotransplant murine model of B cell malignancy. Cancer Res 2011;71: 2871-81.
- 119. Imura Y, Ando M, Kondo T, Ito M, Yoshimura A. CD19-targeted CAR regulatory T cells suppress B cell pathology without GvHD. JCI insight 2020;5:e136185.
- 120. MacDonald KG, Hoeppli RE, Huang Q, Gillies J, Luciani DS, Orban PC, et al. Alloantigen-specific regulatory T cells generated with a chimeric antigen receptor. J Clin Invest 2016;126:1413-24.
- 121. Boardman DA, Philippeos C, Fruhwirth GO, Ibrahim MAA, Hannen RF, Cooper D, et al. Expression of a chimeric antigen receptor specific for donor HLA class I enhances the potency of human regulatory T cells in preventing human skin transplant rejection. Am J Transplant 2017;17:931-43.
- 122. Noyan F, Zimmermann K, Hardtke-Wolenski M, Knoefel A, Schulde E, Geffers R, et al. Prevention of allograft rejection by use of regulatory T cells with an MHC-specific chimeric antigen receptor. Am J Transplant 2017;17:917-30.
- 123. Yoon J, Schmidt A, Zhang A-H, Königs C, Kim YC, Scott DW. FVIII-specific human chimeric antigen receptor T-regulatory cells suppress T- and B-cell responses to FVIII. Blood 2017;129:238-45.
- 124. Zhang A-H, Yoon J, Kim YC, Scott DW. Targeting antigen-specific B cells using antigen-expressing transduced regulatory T cells. J Immunol 2018;201: 1434-41.
- 125. Feng D, Sun J. Overview of anti-BCMA CAR-T immunotherapy for multiple myeloma and relapsed/refractory multiple myeloma. Scand J Immunol 2020;92: e12910.
- 126. Lyman GH, Nguyen A, Snyder S, Gitlin M, Chung KC. Economic evaluation of chimeric antigen receptor T-cell therapy by site of care among patients with relapsed or refractory large B-cell lymphoma. JAMA Netw Open 2020;3: e202072.
- 127. Capecchi R, Puxeddu I, Pratesi F, Migliorini P. New biomarkers in SLE: from bench to bedside. Rheumatology 2020;59:v12-8.
- 128. Ran T, Eichmüller SB, Schmidt P, Schlander M. Cost of decentralized CAR T-cell production in an academic nonprofit setting. Int J Cancer 2020;147:3438-45.
- 129. Lam C, Meinert E, Yang A, Cui Z. Comparison between centralized and decentralized supply chains of autologous chimeric antigen receptor T-cell therapies: a UK case study based on discrete event simulation. Cytotherapy 2021;23:433-51.
- 130. Neelapu SS, Tummala S, Kebriaei P, Wierda W, Gutierrez C, Locke FL, et al. Chimeric antigen receptor T-cell therapy—assessment and management of toxicities. Nat Rev Clin Oncol 2018;15:47-62.
- 131. Komatsu N, Okamoto K, Sawa S, Nakashima T, Oh-Hora M, Kodama T, et al. Pathogenic conversion of Foxp3 + T cells into TH17 cells in autoimmune arthritis. Nat Med 2014;20:62-8.
- 132. Kandane-Rathnayake R, Louthrenoo W, Hoi A, Luo S-F, Wu Y-JJ, Chen Y-H, et al. 'Not at target': prevalence and consequences of inadequate disease control in systemic lupus erythematosus—a multinational observational cohort study. Arthritis Res Ther 2022;24:70.