The Lancet Infectious Diseases

Application and optimization of RT-PCR in diagnosis of SARS-CoV-2 infection --Manuscript Draft--

Manua arint Number								
Manuscript Number:	THELANCETID-D-20-00567							
Article Type:	Article (Original Research)							
Keywords:	Coronavirus Disease 2019 (COVID-19); Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2); real-time reverse transcription PCR (RT-PCR); Chest CT							
Corresponding Author:	Guan-Min Jiang, M.D Fifth Affiliated Hospital of Sun Yat-sen University Zhuhai, Guangdong CHINA							
First Author:	Guan-Min Jiang, M.D							
Order of Authors:	Guan-Min Jiang, M.D							
	Xiaoshuai Ren							
	Yan Liu							
	Hongtao Chen							
	Wei Liu							
	Zhaowang Guo							
	Yaqin Zhang							
	Chaoqun Chen							
	Jianhui Zhou							
	Qiang Xiao							
	Hong Shan							
Manuscript Region of Origin:	CHINA							
Abstract:	Summary							
	Background							
	Coronavirus Disease 2019 (COVID-19) caused by Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) has become a global threat to public health. Aiming to construct an efficient screening pattern, we comprehensively evaluated the performances of RT-PCR and chest CT in diagnosing COVID-19.							
	Methods							
	The records including demographics, RT-PCR, and CT from 87 confirmed COVID-19 cases and 481 exclusion cases were collected. The diagnostic accuracy of the pharyngeal swab RT-PCR, CT, combination with the second pharyngeal swab RT-PCI or with CT were evaluated individually. Besides, all the stool RT-PCR results were plotted by time to explore the value of stool RT-PCR.							
	Findings							
	Combination of RT-PCR and CT has the higher sensitivity (91.9%,79/86) than RT-PCF alone (78.2%,68/87) or CT alone (66.7%, 54 of 81) or combination of two RT-PCR tests (86.2%,75/87). There was good agreement between RT-PCR and CT (kappa-value, 0.430). In 34 COVID-19 cases with inconsistent results, 94.1% (n=32) are mild infection, 62.5% of which (20/32) showed positive RT-PCR. 46.7% (35/75) COVID-19 patients had at least one positive stool during the course. Two cases had positive stoo earlier than the pharyngeal swabs. Importantly, one patient had consecutive positive stool but negative pharyngeal swabs.							

Powered by Editorial Manager® and ProduXion Manager® from Aries Systems Corporation This preprint research paper has not been peer reviewed. Electronic copy available at: https://ssrn.com/abstract=3546086

Interpretation

Combination of RT-PCR and CT with the highest sensitivity is an optimal pattern to screen COVID-19. RT-PCR is superior to CT in diagnosing mild infections. Stool RT-PCR should be considered as an item for improving discovery rate and hospital discharge. This study shed light for optimizing scheme of screening and monitoring of SARS-CoV-2 infection.

1	Application and optimization of RT-PCR in diagnosis of SARS-CoV-2 infection
2	Xiaoshuai Ren ^{1*} , Yan Liu ^{1*} , Hongtao Chen ^{1*} , Wei Liu ¹ , Zhaowang Guo ¹ , Yaqin Zhang ² , Chaoqun
3	Chen ¹ , Jianhui Zhou ¹ , Qiang Xiao ¹ , Guanmin Jiang ^{1,3#} , Hong Shan ^{4#} .
4	¹ Clinical Laboratory, The Fifth Affiliated Hospital, Sun Yat-Sen University. Zhuhai, 519000, China.
5	² Department of Radiology, The Fifth Affiliated Hospital of Sun Yat-sen University, No. 52 Meihua
6	Dong Road, Zhuhai 519000, Guangdong Province, People's Republic of China
7	³ Central Laboratory, The Fifth Affiliated Hospital of Sun Yat-sen University, Zhuhai, Guangdong,
8	China.
9	⁴ Key Laboratory of Biomedical Imaging of Guangdong Province, Guangdong Provincial Engineering
10	Research Center of Molecular Imaging, The Fifth Affiliated Hospital of Sun Yat-sen University, Zhuhai,
11	519000, Guangdong, China.
12	
13	*These authors contributed equally to this article.
14	
15	[#] These senior authors contributed equally to this article.
16	
17	#Corresponding author:
18	Hong Shan, PhD.
19	Guanmin Jiang, PhD
20	

21 Address for correspondence:

- 22 Hong Shan, Ph.D., Professor
- 23 Mailing address: Key Laboratory of Biomedical Imaging of Guangdong Province, Guangdong
- 24 Provincial Engineering Research Center of Molecular Imaging, The fifth Affiliated Hospital of Sun
- 25 Yat-sen University, No. 52 Meihua Dong Road, Zhuhai 519000, Guangdong Province, People's
- 26 Republic of China. Email: shanhong@mail.sysu.edu.cn
- 27
- 28 Guanmin Jiang, Ph.D., Professor
- 29 Mailing address: Clinical Laboratory, The fifth Affiliated Hospital, Sun Yat-Sen University. Zhuhai,
- 30 519000, China. Email: jianggm3@mail.sysu.edu.cn
- 31
- 32
- 33 Running Title: Diagnosis of COVID-19 Infection
- 34

35 Summary

Background: Coronavirus Disease 2019 (COVID-19) caused by Severe acute respiratory
 syndrome coronavirus 2 (SARS-CoV-2) has become a global threat to public health. Aiming to
 construct an efficient screening pattern, we comprehensively evaluated the performances of RT-PCR
 and chest CT in diagnosing COVID-19.

40 Methods: The records including demographics, RT-PCR, and CT from 87 confirmed COVID-19
41 cases and 481 exclusion cases were collected. The diagnostic accuracy of the pharyngeal swab RT-PCR,
42 CT, combination with the second pharyngeal swab RT-PCR or with CT were evaluated individually.
43 Besides, all the stool RT-PCR results were plotted by time to explore the value of stool RT-PCR.

Findings: Combination of RT-PCR and CT has the higher sensitivity (91.9%,79/86) than RT-PCR
alone (78.2%, 68/87) or CT alone (66.7%, 54 of 81) or combination of two RT-PCR tests
(86.2%,75/87). There was good agreement between RT-PCR and CT (kappa-value, 0.430). In 34
COVID-19 cases with inconsistent results, 94.1% (n=32) are mild infection, 62.5% of which (20/32)
showed positive RT-PCR. 46.7% (35/75) COVID-19 patients had at least one positive stool during the
course. Two cases had positive stool earlier than the pharyngeal swabs. Importantly, one patient had
consecutive positive stool but negative pharyngeal swabs.

51 Interpretation: Combination of RT-PCR and CT with the highest sensitivity is an optimal pattern 52 to screen COVID-19. RT-PCR is superior to CT in diagnosing mild infections. Stool RT-PCR should 53 be considered as an item for improving discovery rate and hospital discharge. This study shed light for 54 optimizing scheme of screening and monitoring of SARS-CoV-2 infection.

Funding: This work was supported by the National Natural Science Foundation of China (No.
81502104), National Program on Key Basic Research Project (No. 2018YFC0910600), the Nature
Science Foundation of Guangdong Province, China (Grant No: 2017A030313771 and
2020A151501001) and the Young Teachers Nurturing Program of Sun Yat-Sen University (Grant
No:17ykpy62)

60 61

Keywords: COVID-19, SARS-CoV-2, RT-PCR, chest CT

62

63 Introduction

64 In early December 2019, the first pneumonia cases of unknown origins were identified in Wuhan 65 city, Hubei province, China¹. On Jan 7, a novel coronavirus was discovered using high-throughput sequencing in the throat swab sample of a patient, and is currently named SARS-CoV-2 (previously 66 known as 2019-nCoV) on February 11, 2020 by ICTV^{2,3}. The initial defined cases of COVID-19, were 67 68 epidemiologically linked to the human seafood market in Wuhan, Although later more and more 69 COVID-19 were found without exposure the market but with a history to Wuhan or contact with the patient of COVID-19 pneumonia confirmed^{2,4,5}. Current epidemiologic data indicate the 70 person-to-person transmission of SARS-CoV-2 in hospital and family settings^{2,6,7}. As of February 17, 71 72 2020, more than 71,000 laboratory-confirmed and 1,770 death cases have been documented in China and in other countries worldwide (including the USA, German, japan and South Korea)^{8,9}. The 73 74 mortality rate of SARS-CoV-2 was around 2%. The WHO has recently declared the SARS-CoV-2 a 75 public health emergency of international concern¹⁰. Thus, diagnostic tests specific for this infection are 76 urgently needed for confirming suspected cases, screening patients and conducting virus surveillance.

77 Identification of pathogens mainly includes virus isolation and viral nucleic acid detection. 78 According to the traditional Koch's postulates, virus isolation is the gold standard for virus diagnosis in 79 the laboratory. Thus, based on SARS-CoV-2 possesses a strong capability to infect humans, CDC 80 recommends that clinical virology laboratories should not attempt viral isolation from specimens collected from COVID-19 patients under investigation. Because SARS-CoV-2 is a newly discovered 81 82 virus, the spectrum of the available diagnostic tools is tight. In the early stage, SARS-CoV-2 has been detected in human clinical specimens by next-generation sequencing, cell culture, and electron 83 84 microscopy¹¹. Further development of accurate and rapid methods to identify this emergency 85 respiratory pathogen is still needed.

86 Then the full genome sequence of SARS-CoV-2 (29870-bp, excluding the poly (A) tail) in 87 GenBank (accession number MN908947) was released quickly on January 10, 2020, which is more than 82% identical to those of SARS-CoV and bat SARS-like coronavirus (SL-CoV)¹². On the basis of 88 89 analysis of this complete genomes obtained in this study, several laboratories developed molecular 90 detection tools based on targeting ORF1ab, RNA-dependent RNA polymerase (RdRp) gene N, and E regions of viral spike genes¹³⁻¹⁵. And then the rapid identification of this novel coronavirus is attributed 91 to recent advances in the detection of SARS-CoV-2, including RT-PCR, real-time reverse transcription 92 93 PCR (rRT-PCR), reverse transcription loop-mediated isothermal amplification (RT-LAMP), and 94 microarray-based assays. At present, RT-PCR is a widely used detection technique for SARS-CoV-2 95 and several marketed nucleic acid detection kits for using in clinic¹⁴. Currently, the standard of 96 reference for the COVID-19 pneumonia diagnosis is a positive result in nucleic acid detection assay for 97 the upper and lower respiratory tract specimens and blood, respiratory tract specimens were including 98 nasal and pharyngeal swab specimens, sputum, and bronchoalveolar lavage fluid. And the patients 99 confirmed with the COVID-19 pneumonia had 2 or 3 continuous negative RT-PCR results for 100 nasopharyngeal and throat swab specimens can discharge from hospital. However, the scholar around 101 china indicated that cases of COVID-19 that had 2 or 3 continuous negative RT-PCR results for nasopharyngeal and throat swab specimens before finally laboratory-confirmed[13]. And currently, 102 several reports has reported the positive RT-PCR results for stool of COVID-19 patients^{16,17}. Based on 103 104 the infected patients can potentially shed the SARS-CoV2 through respiratory and fecal-oral routes, 105 The value of RT-PCR results for stool in early diagnosis and monitor of SARS-CoV-2 infection will be 106 study.

107 Fever, cough and dyspnea were the most common symptoms in patients with COVID-19 108 pneumonia. A manifestation similar of those of two other disease caused by coronaviruses, severe acute respiratory syndrome (SARS) and Middle East respiratory syndrome (MERS)¹⁸⁻²⁰. CT is an important 109 method in the diagnosis of lung lesions, and the radiological changes in the lungs of COVID-19 110 111 patients has been characterized²¹. Zhong et al. reported that of 840 COVID-19 patients who underwent 112 CT on admission, around 76.4% manifested abnormal CT imaging features and usually exhibited 113 typical radiological finding of the ground-glass opacity (50%) or bilateral patchy shadowing $(46\%)^{22}$. 114 Based on the "Diagnosis and Treatment Guideline for New Coronavirus Pneumonia (the fifth edition), China". CT scan were used as the clinical diagnostic criteria for COVID-19 pneumonia, but strictly 115 limited in Hubei Province²³. However, the specificity of chest CT is relatively low, alone could not 116 distinguish the SARS-CoV-2 infection from other pathogens well. 117

SARS-CoV-2 causes extensively outbreak in cold winter. In this season, many other pathogens 118 119 causing pneumonia also become prevalent, even including many viral agent. The infectious diseases 120 share some common characteristics in signs, symptoms and laboratory findings. Therefor it is difficult 121 to differentiate COVID-19 suffers from other pneumonia patients purely depending to the manifestation or routine examination. Therefore, an precision screening scheme is urgent to be 122 123 employed. High sensitive test is pivotal to avoiding secondary transmission by missed diagnosed cases. 124 Meanwhile, the positive predictive value also should be counted, for a number of false positive would 125 bring out not only occupation and cost of healthcare resource, but also increasing infection risk of 126 suspected cases isolated in hospital. In this study, we performed a retrospective study in the 568 cases 127 and compare the efficacy of RT-PCR and CT diagnostic approaches in COVID-19 diagnosis, and to provide evidence for future strategic diagnosis in regions outside Hubei Province. 128

129

130 Methods

131 Data sources

For this retrospective, single center study, we recruited 584 patients from Jan 17 to Feb 11, 2020, 132 133 at The Fifth Affiliated Hospital of Sun Yat-sen University in Zhuhai, China, which is a designated 134 infectious hospital. During this period, RT-PCR and chest CT was performed for consecutive patients including the local residents of Wuhan, outside of Wuhan did have a recent travel to Wuhan or contact 135 136 with people with fever or respiratory symptoms from Wuhan, or had fever or acute respiratory 137 symptoms of unknown cause. Of the 16 patients recruited as of Feb 11, had a suspected diagnosis and 138 were therefore excluded in this study. 87 patients, who were diagnosed as having COVID-19 and 481 139 patients exclusion COVID-19 according to WHO interim guidance, were enrolled in this study. The performances of the first RT-PCR detection in pharyngeal swabs and chest CT were evaluated by 140 141 sensitivity, specificity, youden's index et al. Then the performances of combination of the second 142 RT-PCR, or chest CT were also calculated. Agreement between the two method was analyzed using 143 McNemar Chi-squared test. Finally the all RT-PCR results from pharyngeal and stool were plotted by 144 time to explore the value of stool nucleic detection (Fig 1). The severity of COVID-19 pneumonia was 145 defined based on the international guidelines for community-acquired pneumonia²⁴. Laboratory and CT 146 characteristics data were obtained with standard data collection forms from electronic medical records.

147 The study was approved by The Fifth Affiliated Hospital of Sun Yat-sen University Ethics
148 Committee and written informed consent was obtained from patients involved before enrolment when
149 data were collected retrospectively.

150 RNA Extraction and RT-PCR

The SARS-Cov-2 laboratory test assays were based on the previous WHO recommendation. The upper respiratory tract specimens (pharyngeal and nasal swabs) and stool were obtained from all the cases. Ensure each specimen collected has the name, gender and age of the patient as well as a serial number; any abnormality in the specimen should be noted.

RNA was extracted and tested by real-time RT-PCR with SARS-Cov-2 specific primers and probes according to instruction of Kit. The real-time RT-PCR was carried out in biosafety level 2 facilities at the clinic laboratory. If two targets (RdRp+, E or N +) tested positive by specific RT-PCR, the patients would be considered to be laboratory-confirmed.

159 Negative: no Ct value or Ct \geq 40.

160 Positive: a Ct value < 37.

161 A Ct value between 37-40 is indeterminate. It is required confirmation by repeating. If, when 162 repeated, the Ct value is < 37 the sample is positive, otherwise, it is negative.

Chest CT

163

On admission, the chest CT images were detected among 365 patients. Of the 365 patients, typical and atypical chest CT findings were recorded. According to the Diagnosis and Treatment Program of 2019 New Coronavirus Pneumonia (trial sixth version), The typical findings of chest CT images were bilateral multiple lobular and subsegmental areas of consolidation, bilateral ground glass opacity and subsegmental areas of consolidation. Later chest CT images showed bilateral ground-glass opacity.

169 The findings with image features mentioned in Diagnosis and Treatment Program and interpreted 170 by two radiologists are positive. No imaging abnormalities or exclusion of virus infection are defined 171 as negative. That the abnormal imaging doesn't accordance with the features in trial sixth version, but 172 cannot rule out virological infection were defined as uncertain.

173 Statistical analysis.

Retrieved data were recorded into Microsoft [®] Excel and analyzed. Continuous variables were expressed as median and range deviation. The independent sample t-test and one-sample t-test were utilized to compare significant differences among continuous data. Statistical analysis of agreement was performed using McNemar Chi-squared test. It was regarded as statistical significance when the value was less than 0.05. We used R (version 3.5.0) for all analyses.

3. Results

180

179

3.1 The common characteristics of the patients and specimen in this study

The patients involved in this study summed 568, including 87 COVID-19 and 481 non-COVID19. All of them were the local residents of Wuhan, had a recent travel to Wuhan or contacted with people from Wuhan. And they were all eligible for the epidemiology criteria of suspected cases, and received further medical isolation, examination and diagnosis. Among the patients, 58.2% were males. Throughout the whole spectrum of age, 15-59 years group generates 60.6% of all populations in this study, and 5.7% of patients were aged below 15 years (Table 1). In the disease severity, 83.9% (73 of 87 patients) were mild and 16.1% (14 of 87 patients)were severe.

In the early stage of COVID outbreak, RT-PCR and CT scan were the main measure for screening SARS-CoV-2 infection. Hence we collected the results of the two methods in the 568patints in further. In the period of Jan 17th to Feb 11th, total 1674 specimen from the 568 patients were detected by RT-PCR, which included 1415 pharyngeal and 259 stool (Table1). All patients after being admission received at least one RT-PCR detection for pharyngeal swabs, and 341 patients were subjected to the second RTPCR test for pharyngeal swabs. The stool detected by RT-PCR accounted to 259, 81 of which were from COVID-19 patients. Among the 568 patients, 364 patients were subjected to CT scan, 81 of which were COVID-19 patients. We looked through the case data retrospectively and found that 5
patients without CT scan were admitted to hospital complaining of epidemiology contact. Although
they were eventually diagnosed, they had only mild symptoms of upper respiratory tract infection on
admission and in the course of infection, and no CT scan had been performed.

3.2 The performance evaluation of the RT-PCR, CT scan and combination patterns indiagnosis of COVID-19

201 In the beginning of the SARS-CoV-2 outbreak, RT-PCR for pharyngeal and CT scan is the main 202 screening strategy. Aiming to evaluate the property of the two methods in diagnosing SARS-CoV-2 203 infection, the performance indexes of RTPCR detection and CT scan were computed respectively. The 204 results were shown in Table2. The sensitivity and specificity of RT-PCR for pharyngeal were 78.2% 205 and 98.8%, positive predictive value and negative predictive value were 91.9% and 96.2, and Youden's 206 index was 0.770, which indicated overall performance. In the methodology evaluation of CT scan, 207 criteria was defined firstly as an image diagnostic method. According the standard above, the data 208 about the CT scan performance were evaluated. The sensitivity and specificity were 66.7% and 68.2%, 209 lower than RT-PCR of pharyngeal. positive predictive and negative predictive value were 56.8% and 210 92.3% and Youden's index was 0.343 (Table2).

Besides that, we also analyze the agreement of the two methods (The cases with uncertain CT results were not involved in statistical analysis due to statistical limitations). As is shown in Table3, there was statistically significance between the two methods (p<0.001). Moreover, there was statistically differences in the diagnosis of non-COVID-19 patients(p<0.001), but not in COVID-19 patients (p=0.734). The agreement was good to fair agreement (kappa value, 0.430), and the adjusted agreement was 72.8% in all patients.

Given that the time on receiving detection can cause influences on sensitivity, the time differences between RT-PCR for pharyngeal and CT scan were compared. The average time of nucleic acid screening was earlier than that of CT scan statistically (-0.7390days, -3625~1.674) (Table 4). The COVID-19 group received an of CT scan later than the non-COVID-19 group in average (-1.7160,-5.185~1.753 v.s. -0.4594, -2.391~1.472, p=0.002) (Table 4).

222 In purpose to explore the characteristics of the RT-PCR and CT scan in diagnosis of SARS-CoV-2 223 infections, 127 cases with inconsistent diagnosis by two methods were selected for further analysis, 224 which is consisted of 34 COVID-19 patients and 93 non-COVID-19 cases (Fig.2). Notably mild cases 225 count to 94.1% in COVID-19 suffers (32/34) (Fig2.A). 20 of the 32 cases showed RT-PCR positive 226 & CT negative or RT-PCR positive & CT uncertain, suggested the priority of RT-PCR in identifying 227 mild infections. There are still 12 cases with RT-PCR negative & CT positive or RT-PCR negative & 228 CT uncertain, we are inclined to consider unqualified sampling or low viral load in early stage were 229 responsible for the false discovery of RT-PCR. Only two inconsistent cases were severe infections 230 (Fig2.A)., indicating both of the methods reached the good accordance. Besides that, we also focused 231 the results pattern in 93 non-COVID 19, that mainly presented RT-PCR negative & CT positive (n=40) 232 and RT-PCR negative & CT uncertain (n=49) (Fig2.B). That is 95.3% (89/93) patients possessed 233 negative RT-PCR but abnormal CT results. We concluded that CT scan is an morphology detection, 234 not pathogen identification, hence, it was difficult to differentiate SARS-CoV-2 from other viruses or 235 pathogens accurately.

It is notable that both of the methods acquired the sensitivity less than 80% in screening
 SARS-CoV-22 infections, which is not ideal enough for the diagnosis of infectious diseases with severe
 consequences. To develop more appropriate detection scheme, the performances of combination

RT-PCR and CT were evaluated in future. The 568 cases being subjected to first RT-PCR and second
RT-PCR for pharyngeal and 341 being subjected to CT scan were analyzed. The performance indexes
were shown in Table1. The results showed that the sensitivity of RT-PCR in parallel with CT scan was
the highest(91.9%), which was higher than that of parallel with second nucleic acid (86.2%) (Table1).
But the specificity of two nucleic acid detections was significantly higher than that of combination of
nucleic acid and CT, suggesting that nucleic acid in parallel with CT was more appropriate to screen

245 SARS-CoV-22 infections, and two nucleic acid tests for exclusion diagnosis maybe more suitable.

246 3.3 The value of nucleotide detection in stool was evaluated in COVID-19 patients.

247 It is reported that alive virus can survive in stool of COVID-19 patients. According our data, 8.6% 248 patients (Table1) cannot be identified by combination of RT-PCR for pharyngeal swab and CT scan. 249 Since that, can the RT-PCR test for stool be an efficiency assisting examination? In this study, there 250 were 75 COVID-19 cases subjecting to stool nucleotide detection. Therefore, all of the RT-PCR results 251 from 75 case with stool nucleotide detection were plotted along time axis (Figure 3). It was 252 demonstrated that 35 cases had at least one positive results and the discovery rate is 46.7%. The stool 253 presented earlier positive than the throat swab in two cases (patient No. 21, 38) (Figure 3) . Moreover, 254 pharyngeal swab of No.21 patient had been always negative until the end of the study. There were 16 255 patients with remaining positive results of stool after two consecutive negative results of pharyngeal 256 swabs (patient No.2,8,13,15,18,21,24,26,27,29,30,35,39,40,42,48) during their hospitalization 257 (Figure 3). Importantly, of the 14 discharged patients, two cases had stools being negative later than 258 pharyngeal swabs (Patient No.2,10) (Figure 3). The data above suggested that the detection of fecal 259 nucleic acid could be employed to improve the discovery rate and might be developed as an indicator 260 of monitoring and de-isolation.

261 262

Discussion

SARS-CoV-2, a novel betacoronavirus are a major cause of symptomatic respiratory tract infection in all age groups worldwide^{11,16,25}. Timely and accurate diagnosis of the virus enables appropriate treatment of infections. RT-PCR is widely deployed in diagnostic virology. In the case of a public health emergency, proficient diagnostic laboratories can rely on this robust technology to establish new diagnostic tests within their routine services before pre-formulated assays become available.

269 In our study, the sensitivity of RT-PCR was greater than that other reports (78% vs 30%-50%) in the first assay. The reasons for the high sensitivity of our series may include^{26,27}: The first was samples, 270 in some hospital, the patients' nasopharyngeal or oropharyngeal swabs were collected for testing the 271 272 SARS-CoV-2 separately. In our study, the detection specimen was the patient's pharyngeal and nasal 273 swabs combine, compared to the pharyngeal or nasal swab only, indicating higher sensitivity at initial 274 screening. We show that the strategy for the detection of viral RNA in pharyngeal and nasal swabs 275 used for SARS-CoV-2 diagnosis is not perfect. We also found that the virus are present in several stool 276 swabs of patients when pharyngeal and nasal swabs detection negative. Based on the infected patients 277 can potentially shed this pathogen through respiratory and fecal-oral routes, we applied test for oral and 278 stool swabs which could greatly improved detection positive rate.

Among the 87 laboratory-confirmed cases, there were still 19 cases with RT-PCR results testing negative in the first assay. In the false negatives can be caused by poor sample quality, such as respiratory tract samples collected from the oropharynx; collection that is too early or late in the progression of the disease, in the early stages, the number of viruses in the body is not enough to be detected. samples that have not been properly stored, transported, or processed, SARS-CoV-2 coronavirus is RNA virus, which is prone to death and degradation. In the process of collecting samples and transporting them to the laboratory for testing, it takes a long time and the nucleic acid is easy to degrade, so it is not easy to detect positive. At last, technical factors, including virus mutation and PCR inhibition.

In our study, the sensitivity of RT-PCR was higher than that of the chest CT (78.5% vs 66.7%, respectively). The reasons for the relatively lower efficiency of chest CT is may include, 1) The major of COVID-19 patients was mild, in the early detection, there is no features or typical features of chest CT in our study. 2) We divided three groups in the 365 patients, one group is COVID-19 positive, the other group is COVID-19 negative. Our results support RT-PCR combine chest CT for first screening for COVID-19 for patients with clinical and epidemiologic features.

294 To date, two team from China have reported to succeed isolating alive SARS-CoV-2 in 295 COVID-19 patients feces (data unpublished). According to these study, some of the provisions have 296 been supplemented about strictly handling the patient's secretions. Many coronaviruses can be 297 transmitted through oral-fecal route by infecting intestines. SARS-CoV-2 belongs to lineage B, 298 betacoronavirus (β -CoV) genus. The other member in this lineage is SARS-CoV, responsible for 299 SARS outbreak in 2003 in China, can be detected in the stool of patients²⁸. MERS, lineage C β -CoVs, has been proved existing in the feces^{29,30}. For it is not too surprise to detect SARS-CoV-2 in stool. It is 300 301 reported that SARS-CoV-2 RNA has been detected in the stool of a patient in the USA^{16,17}. However, 302 our study give an important hint that in a big portion of COVID-19 patients' stool are there the RNA of 303 SARS-CoV-2 and up to 16 patients presented positive stool after two negative pharyngeal swabs, which indicates stool nucleotide has potential role in monitoring infection as an supplement item. 304 305 Moreover, what deserve attention more is patients No.22, who had none positive pharyngeal swabs in 306 all the stage of infection, but positive stool. Until the end of the observation, the patient were still under 307 treatment. Although it can't be absolutely excluded that pharyngeal presented false negative or shifting 308 positive later, at least stool detection need be involved in examination for the strongly suspected 309 persons with negative pharyngeal swab. Besides that, that stool remain positives in a later stage of 310 infection suggests that fecal nucleic acid negative conversion should be included in the discharge 311 criteria. In our center, there was no recurrence infection in 17 discharged patients, for the stool 312 nucleotide had been detected before being out of hospitalization. Of course, we just provide the 313 evidence the RNA in the stool. whether SARS-CoV-2 can be transmitted by oral to fecal need to be 314 studied in future.

Combination of pharyngeal RT-PCR and chest CT with higher sensitivity is an reasonable option to screen SARS-CoV-2 infection patients. Two pharyngeal RT-PCR detections with higher specificity can be used in exclude diagnosis. RT-PCR has the more advantage in screening mild infection comparing to chest CT. RT-PCR of stool should be adopted to improve discovery rate and counted as an item for discharging from hospital. Our study shed light for the optional scheme of the clinical diagnosis and monitoring of SARS-CoV-2 infection.

- 321
- 322 Declaration of interests

323 All authors declare no competing interests
324
325 Informed consent
326 None
327

328 Reference

Hui DS, E IA, Madani TA, et al. The continuing 2019-nCoV epidemic threat of novel coronaviruses to global health - The latest 2019 novel coronavirus outbreak in Wuhan, China.
 International journal of infectious diseases : IJID : official publication of the International Society for Infectious Diseases 2020; 91: 264-6.

Huang C, Wang Y, Li X, et al. Clinical features of patients infected with 2019 novel coronavirus
in Wuhan, China. *Lancet (London, England)* 2020; **395**(10223): 497-506.

335 3. Severe acute respiratory syndrome-related coronavirus: The species and its viruses – a statement
336 of the Coronavirus Study Group. *BioRxiv*.

4. Chang, Lin M, Wei L, et al. Epidemiologic and Clinical Characteristics of Novel Coronavirus
Infections Involving 13 Patients Outside Wuhan, China. *JAMA* 2020.

Lai CC, Shih TP, Ko WC, Tang HJ, Hsueh PR. Severe acute respiratory syndrome coronavirus 2
(SARS-CoV-2) and coronavirus disease-2019 (COVID-19): The epidemic and the challenges. *International journal of antimicrobial agents* 2020: 105924.

6. Chan JF, Yuan S, Kok KH, et al. A familial cluster of pneumonia associated with the 2019 novel
coronavirus indicating person-to-person transmission: a study of a family cluster. *Lancet (London, England)* 2020; **395**(10223): 514-23.

7. Chen N, Zhou M, Dong X, et al. Epidemiological and clinical characteristics of 99 cases of 2019
novel coronavirus pneumonia in Wuhan, China: a descriptive study. *Lancet (London, England)* 2020;
395(10223): 507-13.

8. WHO. Novel coronavirus—Japan (ex-China). Jan 17, 2020.
https://www.who.int/csr/don/17-january-2020-novel-coronavirusjapan-ex-china/en/ (accessed Jan 19, 2020). 2020.

9. WHO. Novel coronavirus—Republic of Korea (ex-China). Jan 21, 2020.
https://www.who.int/csr/don/21-january-2020-novelcoronavirus-republic-of-korea-ex-china/en/
(accessed Jan 23, 2020). 2020.

WHO. Clinical management of severe acute respiratory infection when Novel coronavirus (nCoV)
 infection is suspected: interim guidance. Jan 11, 2020. accessed Jan 20, 2020.

Thu N, Zhang D, Wang W, et al. A Novel Coronavirus from Patients with Pneumonia in China, *2019. N Engl J Med* 2020; 382(8): 727-33.

I2. Zhou P, Yang XL, Wang XG, et al. A pneumonia outbreak associated with a new coronavirus ofprobable bat origin. *Nature* 2020.

360 13. Chen L, Liu W, Zhang Q, et al. RNA based mNGS approach identifies a novel human coronavirus
361 from two individual pneumonia cases in 2019 Wuhan outbreak. *Emerg Microbes Infect* 2020; 9(1):
362 313-9.

363 14. Chu DKW, Pan Y, Cheng SMS, et al. Molecular Diagnosis of a Novel Coronavirus (2019-nCoV)
364 Causing an Outbreak of Pneumonia. *Clin Chem* 2020.

365 15. Corman VM, Landt O, Kaiser M, et al. Detection of 2019 novel coronavirus (2019-nCoV) by
 366 real-time RT-PCR. *Euro Surveill* 2020; 25(3).

367 16. Holshue ML, DeBolt C, Lindquist S, et al. First Case of 2019 Novel Coronavirus in the United
368 States. *N Engl J Med* 2020.

369 17. Zhang W, Du RH, Li B, et al. Molecular and serological investigation of 2019-nCoV infected

patients: implication of multiple shedding routes. *Emerg Microbes Infect* 2020; **9**(1): 386-9.

371 18. Assiri A, Al-Tawfiq JA, Al-Rabeeah AA, et al. Epidemiological, demographic, and clinical

- 372 characteristics of 47 cases of Middle East respiratory syndrome coronavirus disease from Saudi Arabia:
- a descriptive study. *Lancet Infect Dis* 2013; **13**(9): 752-61.
- 19. Lee N, Hui D, Wu A, et al. A major outbreak of severe acute respiratory syndrome in Hong Kong. *N Engl J Med* 2003; **348**(20): 1986-94.

20. Ksiazek TG, Erdman D, Goldsmith CS, et al. A novel coronavirus associated with severe acute
respiratory syndrome. *N Engl J Med* 2003; 348(20): 1953-66.

- 378 21. Heshui Shi XH, Nanchuan Jiang, Yukun Cao, Osamah Alwalid, Jin Gu, Yanqing Fan, Chuansheng
- Zheng. Radiological findings from 81 patients with COVID-19 pneumonia in Wuhan, China: a
 descriptive study. *Lancet Infect Dis* 2020.
- Wei-jie Guan Z-yN, Yu Hu, Wen-hua Liang, Chun-quan, Jian-xing He, Lei Liu, Hong Shan, .
 Clinical characteristics of 2019 novel coronavirus infection in China. *British Medical Journal* 2020.
- 23. China NHCotPsRo. New coronavirus pneumonia prevention and control program (the fifthedition).
- 24. Metlay JP, Waterer GW, Long AC, et al. Diagnosis and Treatment of Adults with
 Community-acquired Pneumonia. An Official Clinical Practice Guideline of the American Thoracic
 Society and Infectious Diseases Society of America. *Am J Respir Crit Care Med* 2019; 200(7):
 e45-e67.
- 25. Rothe C, Schunk M, Sothmann P, et al. Transmission of 2019-nCoV Infection from an
 Asymptomatic Contact in Germany. *N Engl J Med* 2020.
- 26. Bernheim A, Mei X, Huang M, et al. Chest CT Findings in Coronavirus Disease-19 (COVID-19):
 Relationship to Duration of Infection. *Radiology* 2020: 200463.
- 393 27. Fang Y, Zhang H, Xie J, et al. Sensitivity of Chest CT for COVID-19: Comparison to RT-PCR.
 394 *Radiology* 2020: 200432.
- 28. Chan KH, Poon LL, Cheng VC, et al. Detection of SARS coronavirus in patients with suspected
- **396** SARS. *Emerging infectious diseases* 2004; **10**(2): 294-9.
- 29. Corman VM, Albarrak AM, Omrani AS, et al. Viral Shedding and Antibody Response in 37
 Patients With Middle East Respiratory Syndrome Coronavirus Infection. *Clinical infectious diseases :*
- an official publication of the Infectious Diseases Society of America 2016; **62**(4): 477-83.
- 30. Zhou J, Li C, Zhao G, et al. Human intestinal tract serves as an alternative infection route for
 Middle East respiratory syndrome coronavirus. *Sci Adv* 2017; 3(11): eaao4966.
- 402
- 403

404	Fig1. Flowcha	rt for patient	inclusion
-----	---------------	----------------	-----------

407 Fig2. The analysis of 127 cases with disagreement of the first RT-PCR detection and CT scan. A. In 408 COVID-19 patients. B. In non- COVID-19 patients. In the legend, figures indicates the number of 409 cases. 410 411 412 Fig3. The qRT-PCR results were plotted along time axis in COVID-19 patients (n=75). Shapes and 413 colors were used to represent sample types and results, respectively. Circle indicates pararenal swab 414 and triangle indicates stool. Indigo indicates negative result and pale pink positive result. The dots 415 filled with black represents severe patients. To the deadline of the study, 14 patients (patients

416 No.2,3,4,5,10,11,12,17,23,32,33,54,62,63) had discharged from hospital.
417

Characteristic		COVID-19	non- COVID-19	%
Gender				
	male	40	294	58.8(334/568)
	female	47	187	41.2(234/568)
Age				
	0-14	5	28	5.8(33/568)
	15-49	44	300	60.6(344/568)
	50-64	25	88	19.9(113/568)
	≥65	13	65	13.7(78/568)
Disease				
severity				
	Mild	73	-	83.9(73/87)
	Severe	14		16.1(14/87)
All specimen for q-RT-PC	R			
(n=1674)				
Pharynge	al	623	792	84.5(1415/1674)
	The first	87	481	40.1(568/1415)
	The second	87	254	17.0(241/1415)
stool		181	78	16.0(259/1674)
CT scan (n=364)		81	283	

Table1. The common characteristics of the patients and specimen involved in this study

2

3

4

Table2. Performance of the RT-PCR test and CT scan in diagnosing SARS-CoV2 infection

Detection	Total	Sensitivity%	Specificity%	Positive	Negative	Positive	Negative	Youden's
				predictive predictive 1		likelihood	likelihood	Index
				value%	value%	ratio	ratio	
RT-PCR	568	78.2(68/87)	98.8(475/481)	91.9(68/74)	96.2(475/494)	65.17	0.221	0.770
CT^{a}	364	66.7(54/81)	68.2(193/283)	56.8(54/95)	92.3(193/209)	2.10	0.488	0.343
RT&RT	241	86.2(75/87)	93.5(144/154)	88.2(75/85)	92.3(144/156)	13.26	0.148	0.797
RT&CT ^a	370	91.4(74/81)	66.8(189/283)	62.2(74/119)	97.9(189/193)	2.75	0.129	0.581

^a 60 cases manifested uncertain CT scan results, including 11 COVID-19 patients and 49
non-COVID-19 ones.

CT		RT-I	PCR		1	- 1 :			
CT scan ^a		negative	positive	p value ^b	kappa	adjusted agreement% ^c			
Total	negative	193	16	< 0.001	0.430	72.8			
	positive	51	44						
		244	60						
non-COVID-19	negative	189	4	< 0.001	0.006	50.7			
	positive	40	1						
		229	5						
COVID-19	negative	4	12	0.734	0.047	52.4			
	positive	11	43						
		15	55						

Table3. Comparison of RT-PCR and CT scan in diagnosis of SARS-CoV2 infection

9 ^a The cases with uncertain CT scan results were not included in the paired-Chi-squared test.

¹⁰ ^bStatistical analysis was performed using McNemar Chi-squared test with significance at the p <0.05

11 level.

12 °A kappa statistic of ≥ 0.7 represents excellent agreement, 0.40 to 0.7 represents good to fair agreement,

and <0.40 represents poor agreement

Table 4. Statistics of the day differences between the first RT-PCR detection and CT scan

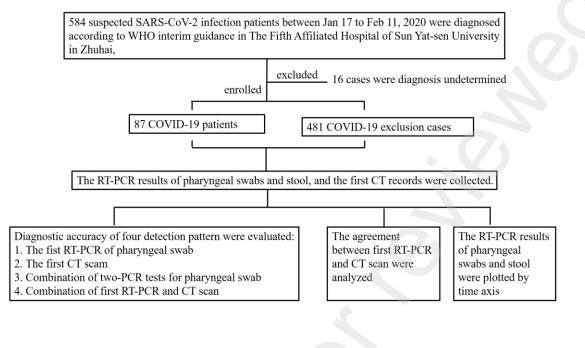
		Mean (95%CI)	p value*	p value#
All patients		-0.7390 (1.674~ -3.652	< 0.001	
All patients	Non-COVID-19	-0.4594 (1.472~ -2.391)	< 0.001	0.000
	COVID-19	-1.7160 (1.753~ -5.185)	< 0.001	0.002

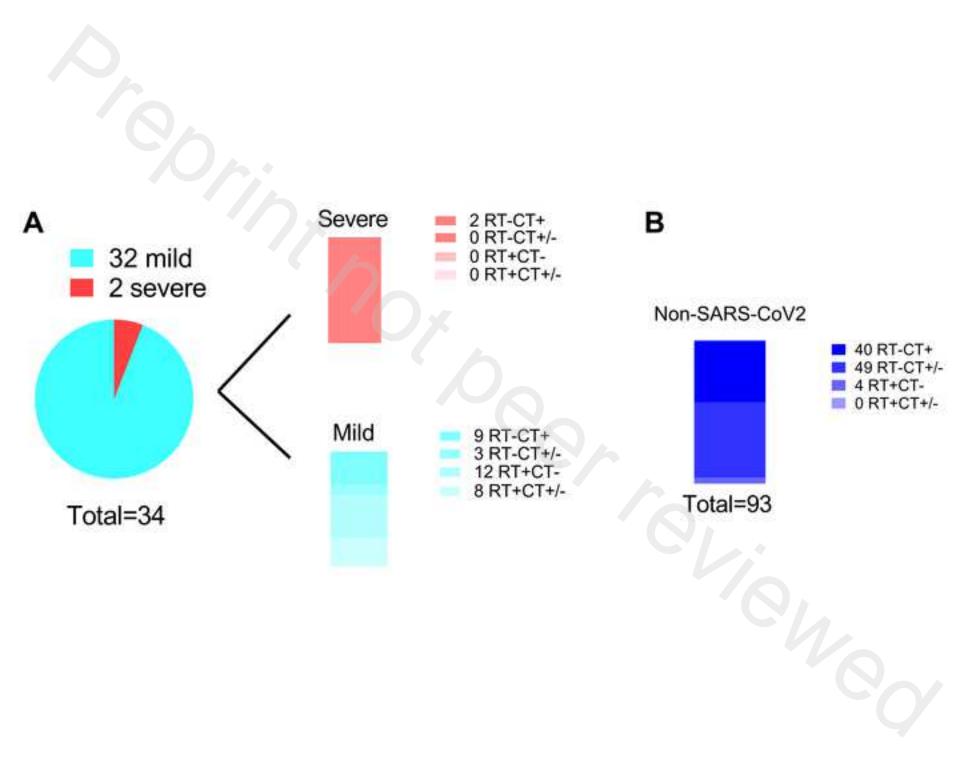
		COVID-19	-1.7160 (1.753~ -5.185)	<0.001
16	The day differences	between the first	RT-PCR detection and CT scan	were performed statistical
17	analysis.			
18		ere used to test stat	istical significance between the me	ean of group and zero at the
19	p <0.05 level.		-	
20		e t test were used	to test statistical significance of the	he day differences between
21	non-COVID-19group			
22		C		
23				

±

1	Fig1. Flowchart for patient inclusion
2	
3	
4	Fig2. The analysis of 127 cases with disagreement of the first RT-PCR detection and CT scan. A. In
5	COVID-19 patients. B. In non- COVID-19 patients. In the legend, figures indicates the number of
6	cases.
7	
8	
9	Fig3. The qRT-PCR results were plotted along time axis in COVID-19 patients (n=75). Shapes and
10	colors were used to represent sample types and results, respectively. Circle indicates pararenal swab
11	and triangle indicates stool. Indigo indicates negative result and pale pink positive result. The dots
12	filled with black represents severe patients. To the deadline of the study, 14 patients (patients
13	No.2,3,4,5,10,11,12,17,23,32,33,54,62,63) had discharged from hospital.
14	
15	

This preprint research paper has not been peer reviewed. Electronic copy available at: https://ssrn.com/abstract=3546086





Figu	ire 1	2	4	5	6	7	8	9	10	11	12	13	14	15	16	17	19	19	20	21	22	23	24	25	26			
	9-•▲	2	4	5	0	0	•	9 0∆	•		12	13	14	15	0	0	10	19	20	21	22	23	24	20	20			
1	8 - • 🔺						•	0	0					0	0	0												
	7 - ▲ 6 -●▲					0		o۵		•▲		0	o∆		0 0	0 0			0					o۵	0			
	5 - • 4 -							oΔ					oΔ	oΔ	o∆ o	∆ ⊙	0		0 0			0		0 0	oΔ			
1	3 -												oΔ	04	0	0			0				oΔ	0	•			
	2-●▲ 1-●▲		•		0	oΔ	• ▲		•				oΔ	0	0	0	∆ o	∆ o	0∆ 0	oΔ		0	•	0 0	0∆ 0			
	0 - 9 - 🔺		oΔ			0	•	oΔ	oΔ		•			0∆ 0∆		0	o o∆	0	0 0∆			0		0	0 0			
	8 -	-								•				0	oΔ	oΔ	0	0	04			Ŭ.	-	0	oΔ			
	7 - 6 -		• 	•		0	•	0	0	•	•	0		0 0			0 0						0	οΔ 0	οΔ 0			
	5 - 4 -•	•	0	∆ 0	oΔ	0	•	0	0	•	•	0	 0	0 0	0	oΔ	0 0	oΔ	 ⊙∆			0Δ	•	0	0 0			
	3 -												0			0	0							0	0			
	2-• 1-	•		•	•									0				0	0	•	•	•	• 	0				
	0 -•	•	0	•	0	0	•	0	0	•	•	0	0	0	0	0	0	0	0	0	0	0	0	0	0			
	27	28	29	30	31	32	33	35	36	37	38	39	41	44	45	46	47	48	49	50	51	52	53	54	55			
1	9 -																											
1	8 - 7 -																									Dis	sease severit	y
<u>ក្ខ</u> 1	6 - 0∆	o۵																								•	mild	
H I I	5 - 4 -		• •	∆ 0		•				o۵																•	severe	
la 1 2 1 2	3 - 2 -0∆	o o∆	0∆ 0	o∆	o∆ o∆		0			0 0	0	oΔ			o∆ 0			0	0	0		0		•				
Bu 1	1 - 0		0	0		• 🔺				0					0	•	-	Ŭ	0	o	oΔ			•		sar	mple.type	
har)	0 -0 9 -0 ∆	0 0	0 0	0 0∆	0 0	•	0 0	0	oΔ	0 0	oΔ	0 0	•	00	oΔ	0 0				0	oΔ	0	oΔ	•	0 0	0	swab	
st pl	8 -0 7 -0∆	o 0∆	0 0	o∆ o∆ 0	o ₀∆	•	o o∆		0	0 0	0 0	∆ o	•	0 0	0 0	0	o∆		0 0	0 0	0	0	•		0 0∆		fecal	
er fir	6 - 0	0	0	0		•	οΔ	o		0	0			0	0		0			0	0		o۵		04			
afte	5 - 0 4 - 0	0 0	 0	0 0	0		0 0		 0	οΔ 0		04	•	οΔ 0	oΔ	0			 ○				0			res	sult	
	3 - <mark>0</mark> 2 -		0			• 🔺	0	o∆	0 0	0 0	0 0	0 0		0 0	0 0	oΔ		•		oΔ	oΔ	0	o∆ 0			•	negative	
	1 - 0	0	0 0	0	0	•	0 0	0	0	0	0	0		0					oΔ			0	0 0 0	•		•	positive	
	0 -0	0	0	0	0	•	0	0	0	0	0	0	• 🔺	0	0	0	0	oΔ	0	0	0	0	0	•	0			
	56	58	60	61	62	63	64	67	68	70	71	72	73	74	75	76	77	78	79	80	81	82	83	84	87			
	9 -																											
	8 - 7 -																							0				
	6 - 5 -																							0				
1	4 -																							oΔ				
	3 - 2 -0	0 0																										
1	1 - 0 0 - 0	0	0 0		oΔ	oΔ	•	0 0	0			0						0										
	9 -	0 0∆	0		٥۵	oΔ	0			•			0					0	•		0							
	8 - <mark>0</mark> 7 -	0			oΔ	oΔ		0 0	0	•▲	•	0	•		0	0		0 0	•		0	• • •	0	Δ	0 0			
	6 - 0	•	•				0		0		•	o	0		oΔ	0	0			0		•			0			
	5 - 4 -0	oΔ	0		0	0	Δ	0	0				0 0		0	0 0	0	0 0	• ▲	0	0	•	0		0 0∆			
	3 - △ 2 -0	0Δ	o ∆	0 0	o∆	o∆		0	0	• ▲	•	о о	∆ ⊙	∆ o		0	oΔ	o o∆	•				0	0	•			
	1 - 0		0	0			0	Δ	Δ				0			oΔ		oΔ		0	Δ			0				
l	0-0	o is pr	o	o nt res		o∆ ch pr	o	o has	• not			0 Diat	0 (io))	0 ∆		o	<u>م</u> م	•		o∆ blo.c	•	toc:/		0	0 m/ab	otroot	2546096	

This preprint research paper has not been peeparient No. Electronic copy available at: https://ssrn.com/abstract=3546086