CASE REPORT

INTESTINAL AND PULMONARY INFECTION BY Cryptosporidium parvum IN TWO PATIENTS WITH HIV/AIDS

Fábio Tadeu Rodrigues REINA(1), Camila Aparecida RIBEIRO(1), Ronalda Silva de ARAÚJO(2) Maria Helena MATTÉ(2), Roberto Esteves Pires CASTANHO(3), Ioshie Ibara TANAKA(4), Ana Maria Ferreira Sornas VIGGIANI(4) & Luciamáre Perinetti Alves MARTINS(3)

SUMMARY

We describe two patients with HIV/AIDS who presented pulmonary and intestinal infection caused by *Cryptosporidium parvum*, with a fatal outcome. The lack of available description of changes in clinical signs and radiographic characteristics of this disease when it is located in the extra-intestinal region causes low prevalence of early diagnosis and a subsequent lack of treatment.

KEYWORDS: HIV/AIDS; Cryptosporidiosis; Pneumonia; Cryptosporidium parvum.

INTRODUCTION

Cryptosporidium parvum is an obligate intracellular parasite of the Coccidia class that infects the microvilli epithelial cells of the digestive and respiratory systems¹. This parasite is responsible for causing severe diarrhea in approximately 55% of human immunodeficiency virus/ acquired immune deficiency syndrome (HIV/AIDS) patients living in developing countries².

Among the 16 currently described species of *Cryptosporidium*, *C. parvum and C. hominis* are those that predominate in immunocompromised individuals³.

Infection occurs after ingestion of water or food contaminated with oocysts or direct person-to-person or animal-person contact⁴. Respiratory forms of infection can happen upon inhalation of oocysts during an episode of vomiting^{5,6}.

Studies by LOPEZ-VELEZ *et al.*⁷ and CLAVEL *et al.*⁸ found that 30.2% of patients with intestinal cryptosporidiosis also carried extraintestinal infections in both the lungs and bile. The studies mentioned above reported high mortality rates amongst the patients as these cases showed dramatically lower CD4 + cell counts, ultimately reflecting a very severe degree of immunosuppression.

Therefore, we decided it to be of clinical and educational significance

to report our finding of *C. parvum* (identified by molecular testing) in fecal samples and sputum from two HIV/AIDS patients.

This study was submitted and approved by the Research on Human Beings Ethics Committee of the *Marília* Medical School (FAMEMA), under the number 33677514.1.0000.5413. Figure 2 was made using the medical records of patients what are in the custody of the *Serviço de Prontuário de Paciente* (SPP) of *Marília* Medical School. Similarly, the blades for the manufacture of Figure 3 were photomicrographed in Olympus microscope BX41 model coupled to a digital video camera, Olympus, DP 25 model, DP2-BSW software, Olympus, originals of FAMEMA Parasitology and Microbiology Laboratories.

Patient history

Case 1:

On 10/06/2013, a fifty-nine-year-old female patient sought medical care at the emergency ward of the *Marilia* Clinical Hospital. The patient reported a two-month-long history of diarrhea, along with oral moniliasis associated with weight loss of 15 kg +/- over this period. She denied any fever, cough, urinary changes or emesis. The patient reported one prior medical consultation within Brazil's Specialized Care Service (SAE) with the same story and referring indication of highly active antiretroviral therapy (HAART), however, without any confirmed CD4 + result.

⁽¹⁾ Faculdade de Medicina de Marília, Departamento de Moléstias Infecciosas. Marília, SP, Brasil. E-mails: Fábio Tadeu Rodrigues Reina: ftrreina@yahoo.com.br; Camila Aparecida Ribeiro: riber.camila@gmail.com;

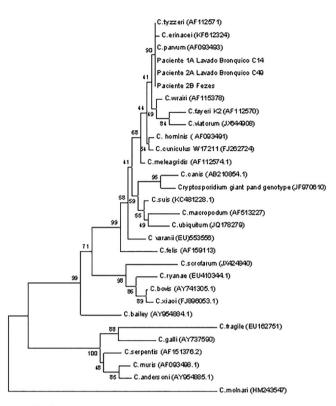
⁽²⁾ Faculdade de Saúde Pública da Universidade de São Paulo, Departamento de Prática de Saúde Pública, São Paulo, SP, Brasil. E-mails: Ronalda Silva de Araújo: ronalda@usp.br; Maria Helena Matté: mhmatte.usp@gmail.com;

⁽³⁾ Faculdade de Medicina de Marília, Disciplina de Parasitologia. Marília, SP, Brasil. E-mails: Roberto Esteves Pires Castanho: castamho@famema.br; Luciamáre Perinetti Alves Martins: luciamarepam@gmail.com

⁽⁴⁾ Faculdade de Medicina de Marília, Disciplina de Microbiologia, Marília, SP, Brasil. E-mails: Ioshie Ibara Tanaka: yoibara@hotmail.com; Ana Maria Ferreira Sornas Viggiani: ana.sornas@bol.com.br

Correspondence to: Luciamáre Perinetti Alves Martins. E-mail: luciamarepam@gmail.com

Reina FTR, Ribeiro CA, Araújo RS, Matté MH, Castanho REP, Tanaka II, Viggiani AMFS, Martins LPA. Intestinal and pulmonary infection by *Cryptosporidium parvum* in two patients with HIV/AIDS. Rev Inst Med Trop Sao Paulo. 2016;58:21.



0.005

Fig. 1 - A phylogenetic tree was inferred using the neighbor-joining method²⁸ with 1000 bootstrap replicates, and evolutionary distances calculated using the Kimura 2-parameter²⁹ method. Evolutionary analyzes were performed by MEGA5 software³⁰.



Fig. 2 - A: Nonspecific radiographic changes presented by Patient 1. B: Patient 2 without significant radiographic changes.

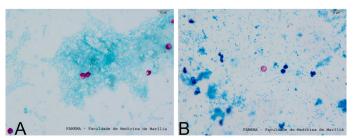


Fig. 3 - A: *Cryptosporidium* spp. oocysts in faeces stained by the Kinyoun method. **B:** *Cryptosporidium* spp. oocysts in sputum stained by the Ziehl-Neelsen method.

On the day of treatment, the patient had a physical examination that determined a poor general condition: pale; dehydrated; Heart rate: 68 bpm; Blood pressure: 80 x 50 mmhg; afebrile.

Lungs: Vesicular murmur: present with bibasilar crepitant rales.

Heart: S₁ and S₂ heart sounds normal and regular; without murmurs.

Abdomen: excavated, bowel sounds present and hypoactive, limp and painless.

Lower limb: weak pulse, with capillary refill time: 3 seconds.

At this time, supportive measures were instituted, including a broad spectrum treatment for opportunistic diseases. Laboratory tests were requested, including for detection of *Cryptosporidium* spp., which ultimately proved positive in both stool samples (method Kinyoun) and sputum (the Ziehl-Neelsen).

Case 2:

On 10/18/2013, a forty-four-year-old female patient was admitted to the Infectious Diseases Ward of *Marilia* Clinical Hospital. The patient reported experiencing diarrhea for approximately the previous three months. During this period, the patient experienced a weight loss of 10 kg, associated with an unmeasured fever and dry cough. On the day of admission, the patient presented a normal general condition: conscious, oriented, dehydrated and pale. Heart: S₁ and S₂ heart sounds normal; without murmur Heart rate: 86 bpm, Lungs: Vesicular murmur: present, no adventitious sounds (Saturation: 98%).

Supportive measures and laboratory tests were performed.

Treatment of esophageal candidiasis was initiated upon hospitalization. An Acid-Alcohol Resistant Bacilli (AARB) exam was performed with negative results, and the analysis of stool and sputum samples for *Cryptosporidium* returned positive.

Molecular testing: Stool and sputum samples from both patients were submitted to a specific detection of Cryptosporidium using Nested PCR method. The fragment of the 18S rRNA subunit was amplified by nested PCR by using the forward primers: SCL1F 5'-CTGGTTGATCCTGCCAGTAG-3' and reverse CPB-DIAGR 5'-AAGGTGCTGAAGGAGTAAGG-3' which amplify 1035 pb and SSU F 5'-GGAAGGGTTTTTATTGTAAGATAAAG-3' and SSUR 5'-AAGGAGTAA GGATCCACCACAA- 3' which amplify about 826 pb as previously described by COUPE et al.9 and XIAO et al.10 to identify of Cryptosporidium spp. The fragments of the secondary PCR were purified with GFXTM PCR DNA Gel Band Purification Kit (GE-UK) and directly sequenced. The nucleotide sequences obtained were analyzed and compared with those registered in GenBank, and phylogenetic analysis were used on the aligned sequences to assess relationships among sequences that allowed the identification of Cryptosporidium parvum in both patients.

DISCUSSION

The low number of diagnosed cases of extraintestinal cryptosporidiosis,

Reina FTR, Ribeiro CA, Araújo RS, Matté MH, Castanho REP, Tanaka II, Viggiani AMFS, Martins LPA. Intestinal and pulmonary infection by *Cryptosporidium parvum* in two patients with HIV/AIDS. **Rev Inst Med Trop Sao Paulo. 2016;58:**21.

especially of pulmonary location, is largely due to the absence of specific clinical signs, as well as the presence of radiological abnormalities, which can be confused with other opportunistic infections that commonly affect patients with immune impairment.

In our cases, parasitological examination by Ziehl-Neelsen and Kinyoun techniques demonstrated the presence of *Cryptosporidium* spp. in the sputum and stool (Fig. 3) of two patients. Confirming the presence of the specific species, *C. parvum*, was performed by nested PCR of the 18S rRNA region in sputum and feces of patient 2, and sputum of patient 1. Sequencing of the samples was then performed and aligned with the sequences available in the GenBank database, showing similarity with the *C. parvum* species, as can be observed in Figure 1.

There is no sufficient molecular data available to support information on the prevalence of *C. parvum* in clinical cases in Brazil. ASSIS *et al.*¹¹ detected *Cryptosporidium parvum* in 10 HIV-positive patients in *Minas Gerais*, suggesting a possible zoonotic transmission of this species of *Cryptosporidium*. However, many studies showed that *C. hominis*, along with *C. parvum*, are the most frequent species observe in humans worldwide^{12,13}.

The differentiation of *Cryptosporidium* to species level depends on the identification by specifics molecular methods, which limits the availability of information on the spread of this species of *Cryptosporidium* in human clinical cases, mainly in Brazil¹². Therefore, the epidemiological importance of reporting these cases is bolstered by the fact that the transmission of this parasite occurs from personperson¹⁴, since certain promiscuous habits can happen primarily amongst practitioners of oro-anal intercourse, putting these individuals at specific risk of infection^{12,15,16}.

Although the results demonstrated the presence of *C. parvum* in stool and sputum, indicating the possibility that pulmonary infection has occurred through inhalation of oocysts during an episode of vomiting, as mentioned by some authors⁶, one cannot rule out the spread by hematogenous route, since it is described as the presence of the parasite within macrophages¹⁷.

GENTILE *et al.*¹⁸ reported finding *Cryptosporidium* oocysts within blood vessels and studies of MARTINEZ *et al.*¹⁹ in an *in vitro* study conducted in mice, demonstrated that *Cryptosporidium* can multiply within macrophages, resisting the action of lysosomal enzymes, making it difficult to control the infection in immunocompromised patients in case that this possibility occurred in humans.

Although radiographic changes occurred only in Patient 1, (Fig. 2), specific treatment for cryptosporidiosis was not instituted, because until then there was no effective treatment for cryptosporidiosis in HIV/AIDS patients. Several authors showed the limited effectiveness of drugs like Nitazoxanide, Macrolides, or Paromomycin for treating cryptosporidiosis in immunocompromised patients, even if HAART was associated^{20,21,22,23}. Recently, promising results have been obtained by CASTELLANOS-GONZALEZ *et al.* (2013)²⁴ when they treated immunosuppressed mice infected with *C. parvum* using Calcium-dependent protein kinases inhibitor (CDPK1 inhibitor).

Therefore at this time, we choose to treat the most prevalent infectious

diseases in these patients, and antiretroviral therapy HAART was initiated to recovery the cellular immune system²⁵, since CD4+ cells are decisive for the acquired immune response²⁶. However, in a short space of time, there was a worsening of the general condition of Patient 1, and she progressed to death. In relation to Patient 2, as she had been clinically stable, the patient was discharged with guidelines for outpatient follow-up and established HAART. However, the patient did not return to her scheduled consultations, resulting in an interruption in medical care and, consequently, the maintenance treatment, leading to a fatal outcome.

Seeing as a pulmonary infection, it is considered a rare complication in its intestinal form²⁷, combined with the high mortality rate of these cases, the lack of formal description of clinical and radiographical changes seen upon extra-intestinal localization of this parasite, cause an extremely low rate of early diagnosis and untimely treatment.

REFERENCES

- 1. Current WL, Garcia LS. Cryptosporidiosis. Clin Microbiol Rev. 1991;4:325-58.
- Soto M, Velásquez G, Cuervo C, Galvis MT, Botero D. Criptosporidiosis respiratoria en un paciente con SIDA. Acta Med Colomb. 1997;22:148-50.
- Meamar AR, Guyot K, Certad G, Dei-Cas E, Mohraz M, Mohebali M, et al. Molecular characterization of *Cryptosporidium* isolates from humans and animals in Iran. Appl Environ Microbiol. 2007;73:1033-5.
- Kosek M, Alcantara C, Lima AA, Guerrant RL. Cryptosporidiosis: an update. Lancet Infect Dis. 2001;1:262-9.
- Albuquerque YM, Silva MC, Lima AL, Magalhães V. Pulmonary cryptosporidiosis in AIDS patients, an underdiagnosed disease. J Bras Pneumol. 2012;38:530-2.
- Corti M, Villafañe MF, Muzzio E, Bava J, Abuín JC, Palmieri OJ. Pulmonary cryptosporidiosis in AIDS patients. Rev Argent Microbiol. 2008;40:106-8.
- López-Vélez R, Tarazona R, Garcia Camacho A, Gomez-Mampaso E, Guerrero A, Moreira V, et al. Intestinal and extraintestinal cryptosporidiosis in AIDS patients. Eur J Clin Microbiol Infect Dis. 1995;14:677-81.
- Clavel A, Arnal AC, Sánchez EC, Cuesta J, Letona S, Amiguet JA, et al. Respiratory cryptosporidiosis: case series and review of the literature. Infection. 1996;24:341-6.
- Coupe S, Sarfati C, Hamane S, Derouin F. Detection of *Cryptosporidium* and identification to the species level by nested PCR and restriction fragment length polymorphism. J Clin Microbiol. 2005;43:1017-23.
- Xiao L, Alderisio K, Limor J, Royer M, Lal AA. Identification of species and sources of *Cryptosporidium* oocysts in storm waters with a small-subunit rRNA-based diagnostic and genotyping tool. Appl Environ Microbiol. 2000;66:5492-8.
- Assis DC, Resende DV, Cabrine-Santos M, Correia D, Oliveira-Silva MB. Prevalence and genetic characterization of *Cryptosporidium* spp. and *Cystoisospora belli* in HIV-infected patients. Rev Inst Med Trop Sao Paulo. 2013;55:149-54.
- Chalmers RM, Davies AP. Minireview: clinical cryptosporidiosis. Exp Parasitol. 2010;124:138-46.
- Lucca P, De Gaspari EN, Bozzoli LM, Funada MR, Silva SO, Iuliano W, et al. Molecular characterization of *Cryptosporidium* spp. from HIV infected patients from an urban area of Brazil. Rev Inst Med Trop Sao Paulo. 2009;51:341-3.
- Araújo AJ, Kanamura HY, Almeida ME, Gomes AH, Pinto TH, Da Silva AJ. Genotypic identification of *Cryptosporidium* spp. isolated from HIV-infected patients and immunocompetent children of São Paulo, Brazil. Rev Inst Med Trop Sao Paulo. 2008;50:139-43.

Reina FTR, Ribeiro CA, Araújo RS, Matté MH, Castanho REP, Tanaka II, Viggiani AMFS, Martins LPA. Intestinal and pulmonary infection by *Cryptosporidium parvum* in two patients with HIV/AIDS. Rev Inst Med Trop Sao Paulo. 2016;58:21.

- Bachur TP, Vale JM, Coêlho IC, Queiroz TR, Chaves CS. Enteric parasitic infections in HIV/AIDS patients before and after the highly active antiretroviral therapy. Braz J Infect Dis. 2008;12:115-22.
- Keystone JS, Keystone DL, Proctor EM. Intestinal parasitic infections in homosexual men: prevalence, symptoms and factors in transmission. Can Med Assoc J. 1980;123:512-4.
- Ma P, Villanueva TG, Kaufman D, Gillooley JF. Respiratory cryptosporidiosis in the acquired immune deficiency syndrome. Use of modified cold Kinyoun and Hemacolor stains for rapid diagnoses. JAMA. 1984;252:1298-301.
- Gentile G, Baldassarri L, Caprioli A, Donelli G, Venditti M, Avvisati G, et al. Colonic vascular invasion as a possible route of extraintestinal cryptosporidiosis. Am J Med. 1987;82:574-5.
- Martinez F, Mascaro C, Rosales MJ, Diaz J, Cifuentes J, Osuna A. *In vitro* multiplication of *Cryptosporidium parvum* in mouse peritoneal macrophages. Vet Parasitol. 1992;42:27-31.
- Abubakar I, Aliyu SH, Arumugam C, Hunter PR, Usman NK. Prevention and treatment of cryptosporidiosis in immunocompromised patients. Cochrane Database Syst Rev. 2007;1:CD004932
- Cabada MM, White AC Jr. Treatment of cryptosporidiosis: do we know what we think we know? Curr Opin Infect Dis. 2010;23:494-9.
- Checkley W, White AC Jr, Jaganath D, Arrowood MJ, Chalmers RM, Chen XM, et al. A review of the global burden, novel diagnostics, therapeutics, and vaccine targets for *Cryptosporidium*. Lancet Infect Dis. 2015;15:85-94.

- Kurniawan A, Dwintasari SW, Connelly L, Nichols RA, Yunihastuti E, Karyadi T, et al. *Cryptosporidium* species from human immunodeficiency-infected patients with chronic diarrhea in Jakarta, Indonesia. Ann Epidemiol. 2013;23:720-3.
- Castellanos-Gonzales A, White AC Jr, Ojo KK, Vidadala RS, Zhang Z, Reid MC, et al. A novel calcium-dependent protein kinase inhibitor as a lead compound for treating cryptosporidiosis. J Infect Dis. 2013;208:1342-8.
- Miao YM, Awad-El-Kariem FM, Franzen C, Ellis DS, Müller A, Counihan HM, et al. Eradication of cryptosporidia and microsporidia following successful antiretroviral therapy. J Acquir Immune Defic Syndr. 2000;25:124-9.
- Patenburg B, Dann SM, Wang HC, Robinson P, Castellanos-Gonzalez A, Lewis DE, et al. Intestinal immune response to human *Cryptosporidium* sp. infection. Infect Immun. 2008;76:23-9.
- Dupont C, Bougnoux ME, Turner L, Rouveix E, Dorra M. Microbiological findings about pulmonary cryptosporidiosis in two AIDS patients. J Clin Microbiol. 1996;34:227-9.
- Saitou N, Nei M. The neighbor-joining method: a new method for reconstructing phylogenetic trees. Mol Biol Evol. 1987;4:406-25.
- Tamura K, Nei M, Kumar S. Prospects for inferring very large phylogenies by using the neighbor-joining method. Proc Natl Acad Sci USA. 2004;101:11030-5.
- Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S. MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. Mol Biol Evol. 2011;28:2731-9.

Received: 27 November 2014 Accepted: 11 June 2015