



Evolução e Desenvolvimento de olhos

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Doutoranda

CONTEÚDO

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Aspectos evolutivos

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Desenvolvimento dos olhos

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Perda de olhos

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Aula Pática



Como olhos se tornaram tão diversificados?

MORFOLÓGICO

FISIOLOGICO

EMBRIONÁRIO

Uma questão de
homologia



Vertebrados



Cefalópodes



Insetos

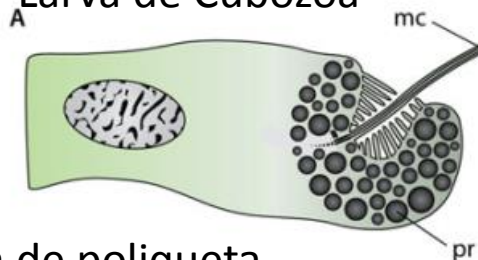


Poliquetas

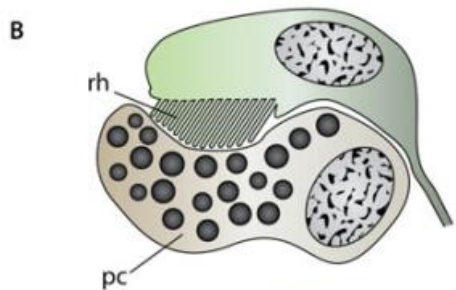
MORFOLÓGICO

Definição do olho

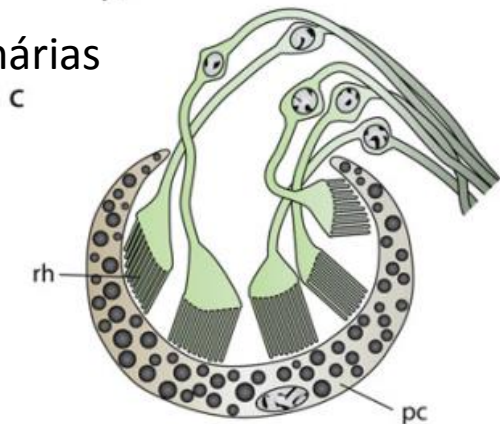
Larva de Cubozoa



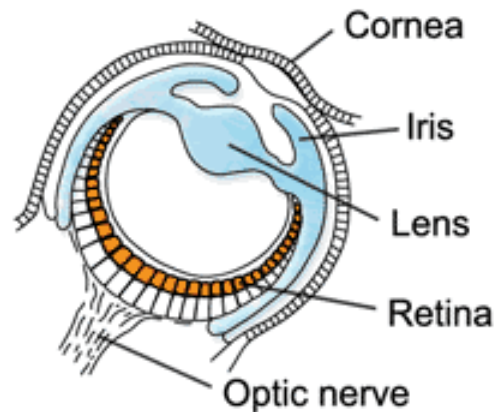
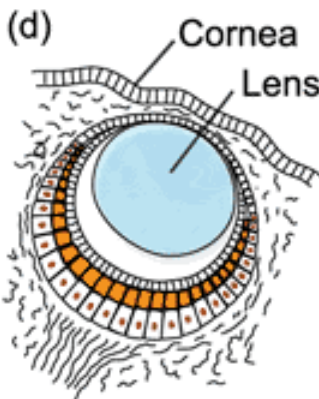
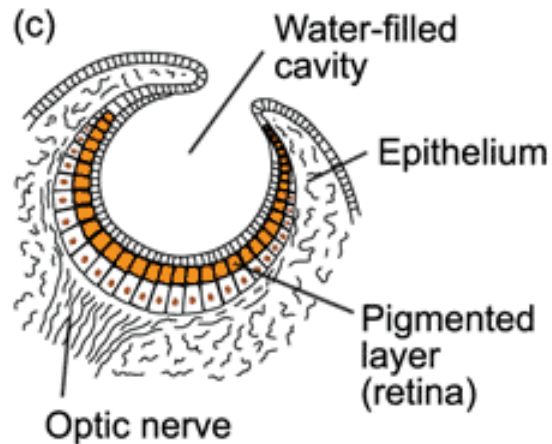
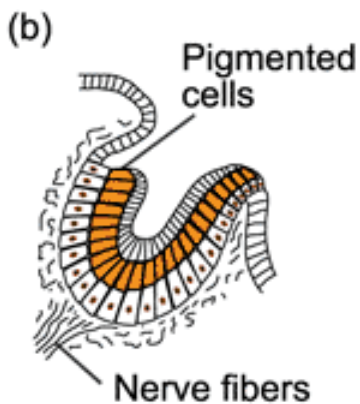
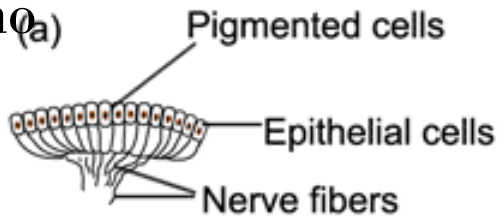
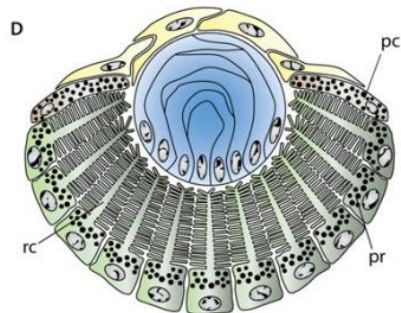
Larva de poliqueta



planárias



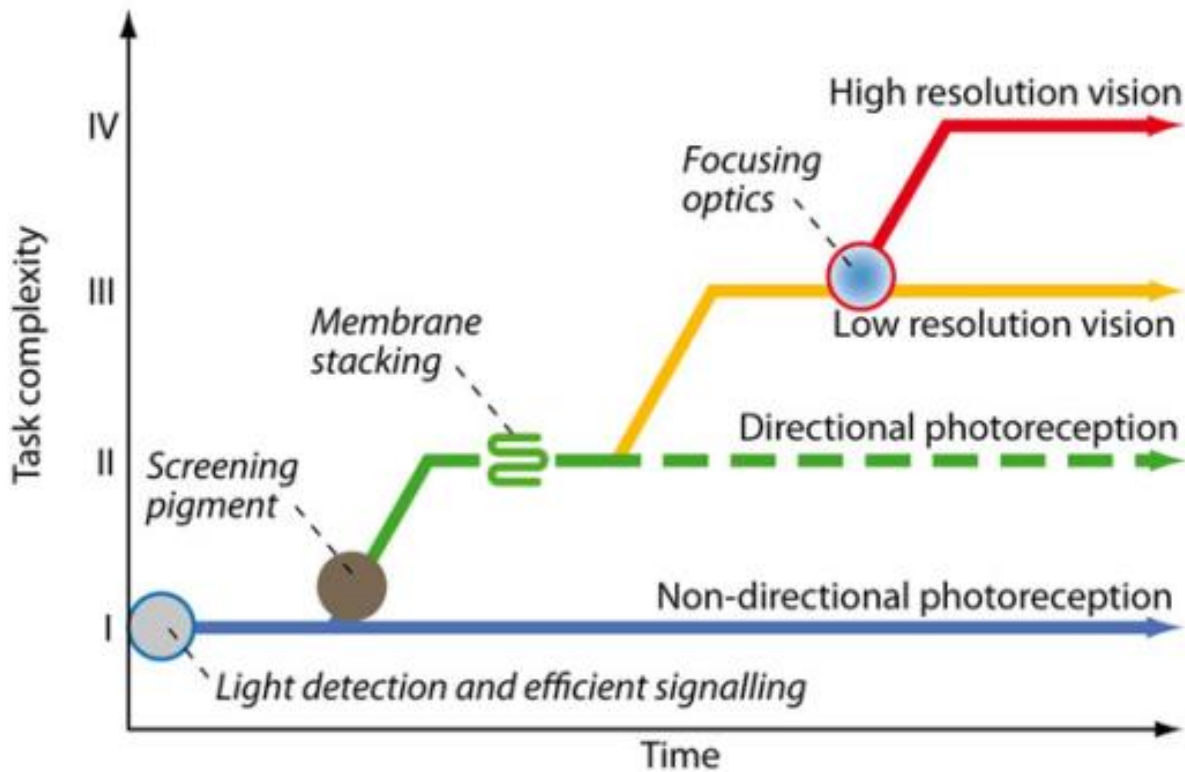
Água viva



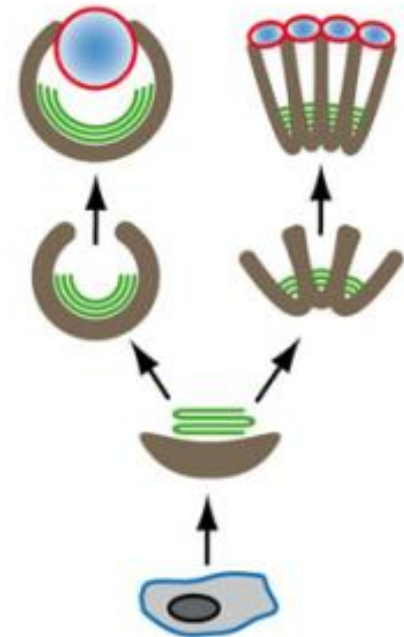
- (a) A pigment spot; (b) a simple pigment cup;
- (c) the simple optic cup found in abalone;
- (d) the complex lensed eye of the marine snail and of the octopus.

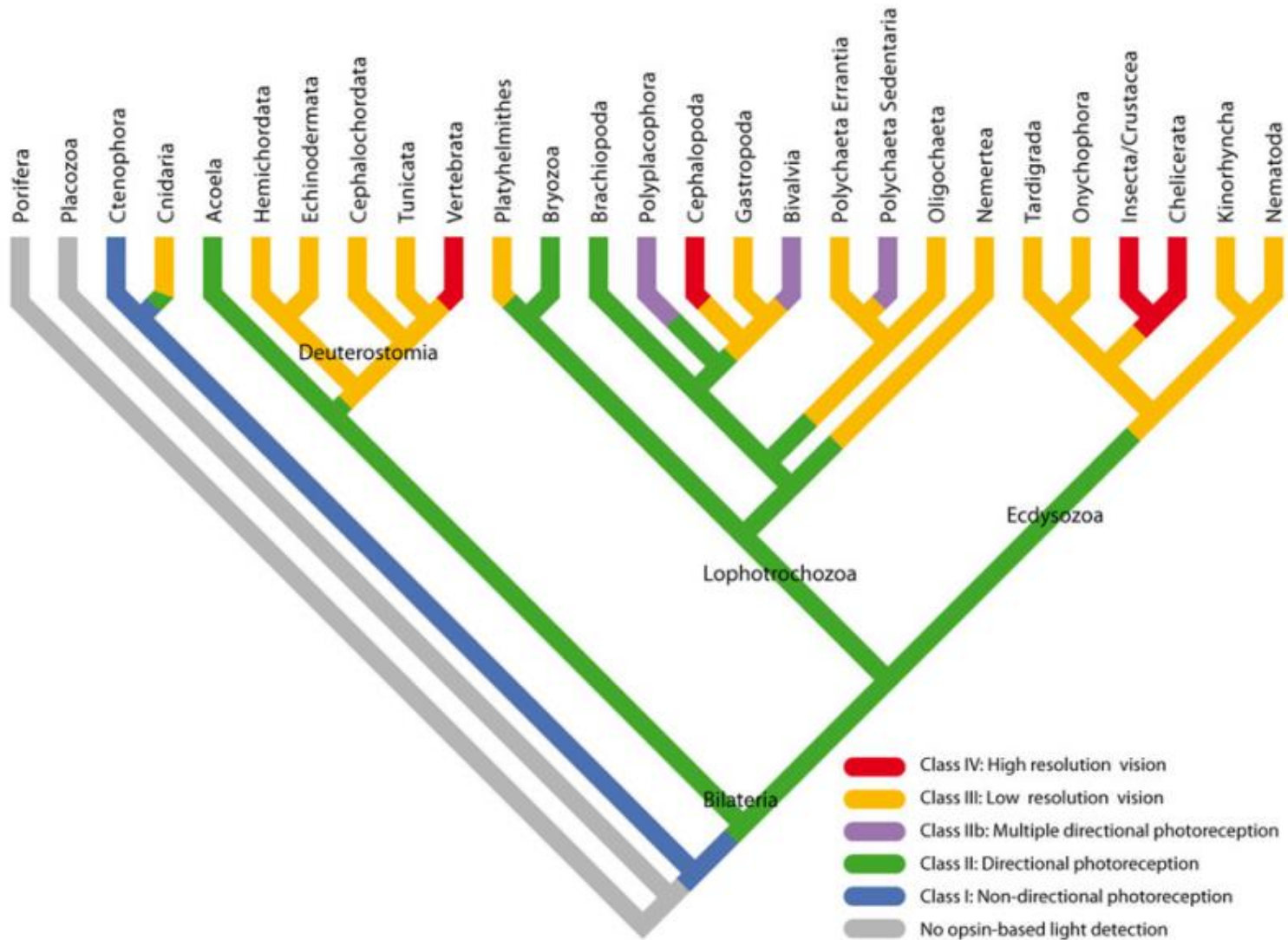
Designed according to Ernst Mayr "What evolution is" (2001 p. 206)

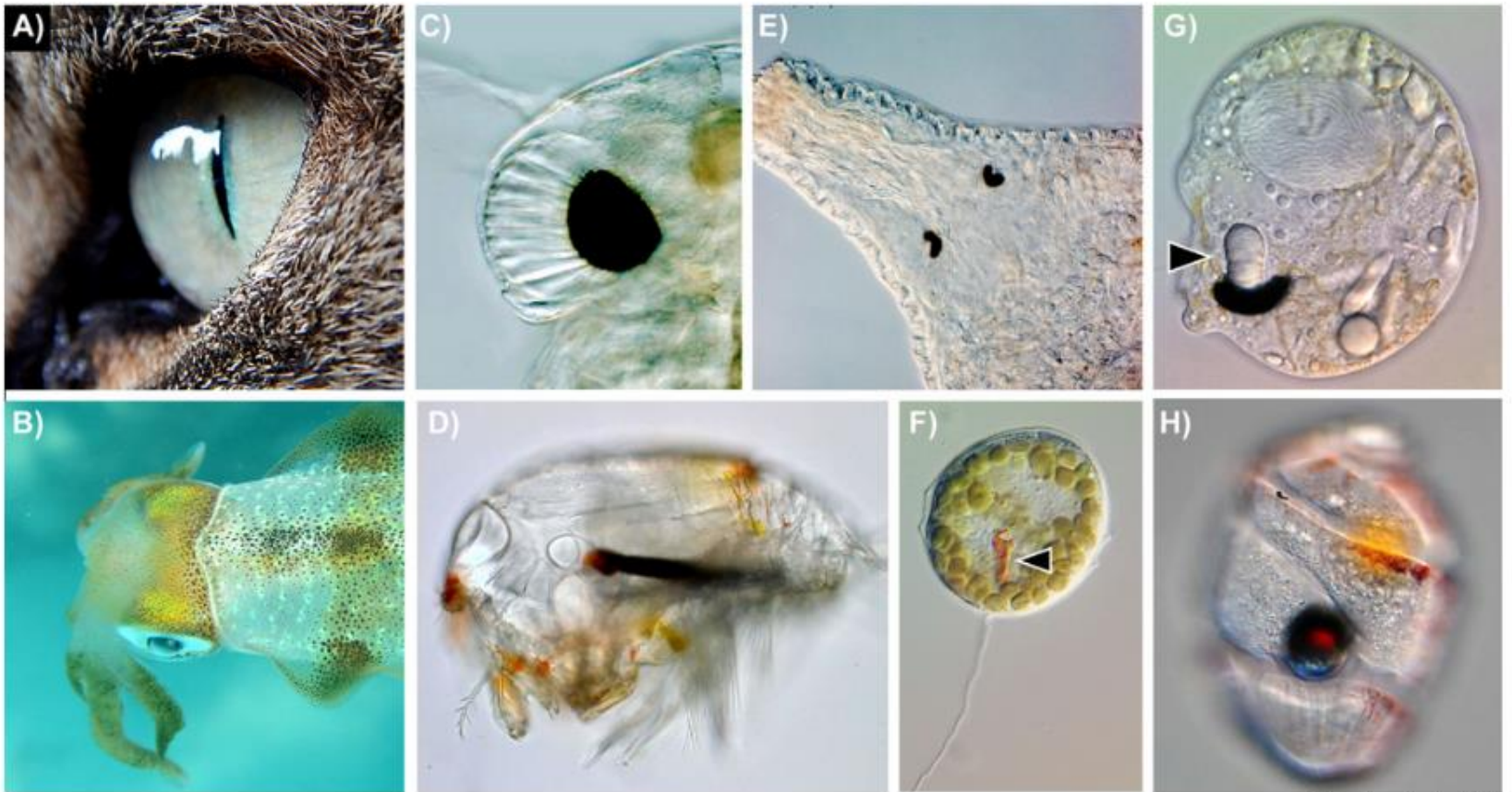
Functional innovations and evolution of behavioural classes



Corresponding eye evolution



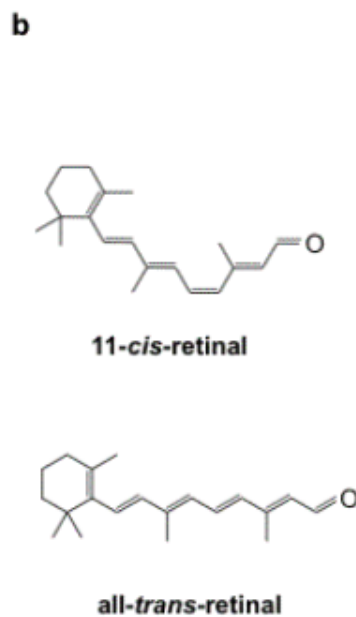
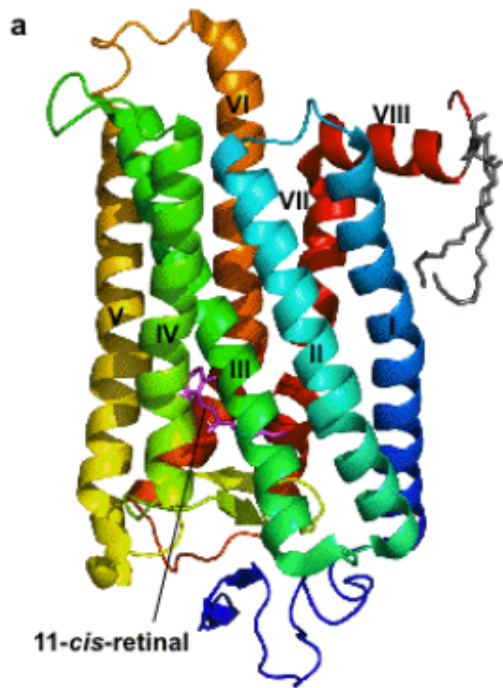




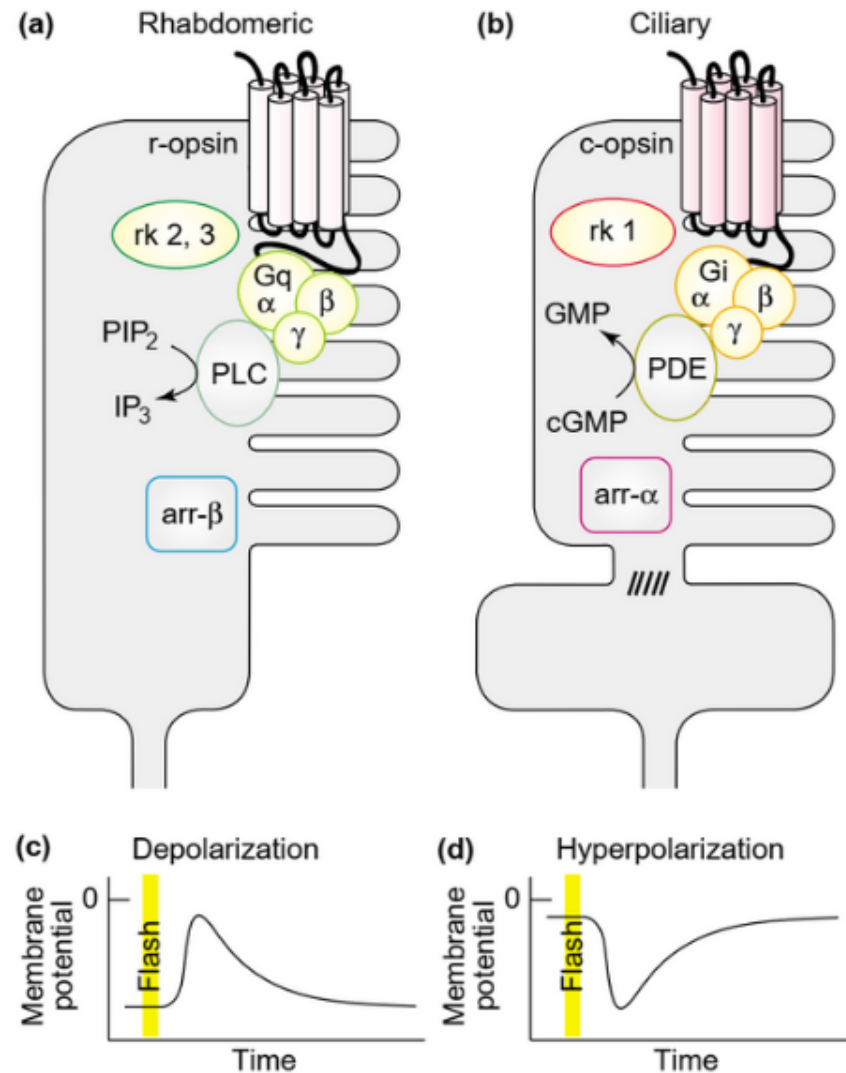
Gavelis, et al., 2017

Todos os olhos compartilham a mesma célula fotorreceptora?

FISIOLÓGICO



Opsinas - proteínas receptoras

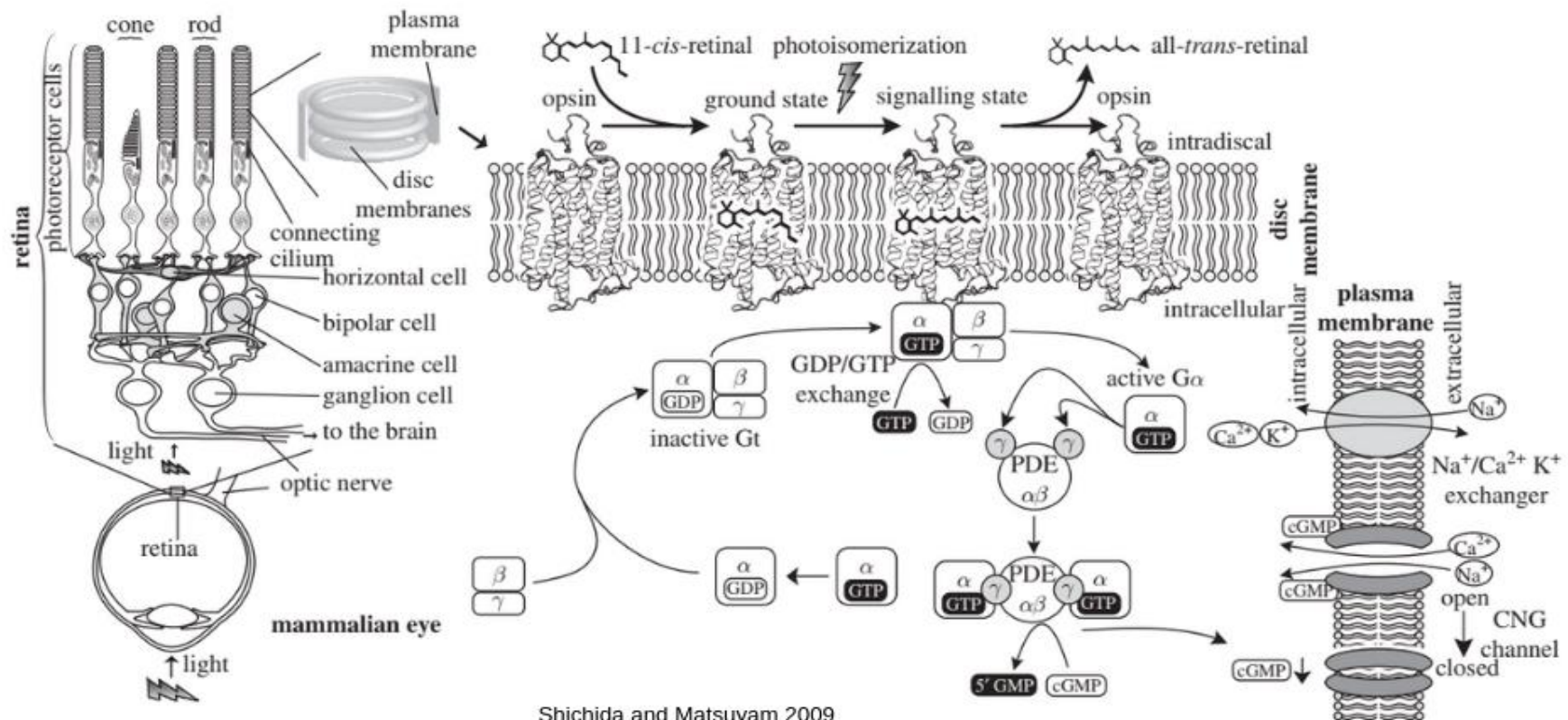


Mesmo caminho de fototransdução

- 1 - Percepção da luz por rodopsinas
- 2 - Rodopsina ativa G proteína
- 3 - G proteína ativa uma cascata de sinalização enzimática
- 4 - resposta elétrica percebida pelas células fotorreceptoras

Existem 2 tipos de células fotorreceptoras

- Ciliares - superfície fotossensível derivada de um cílio.
- Rabdômérico - derivada de microvilosidades da superfície da célula



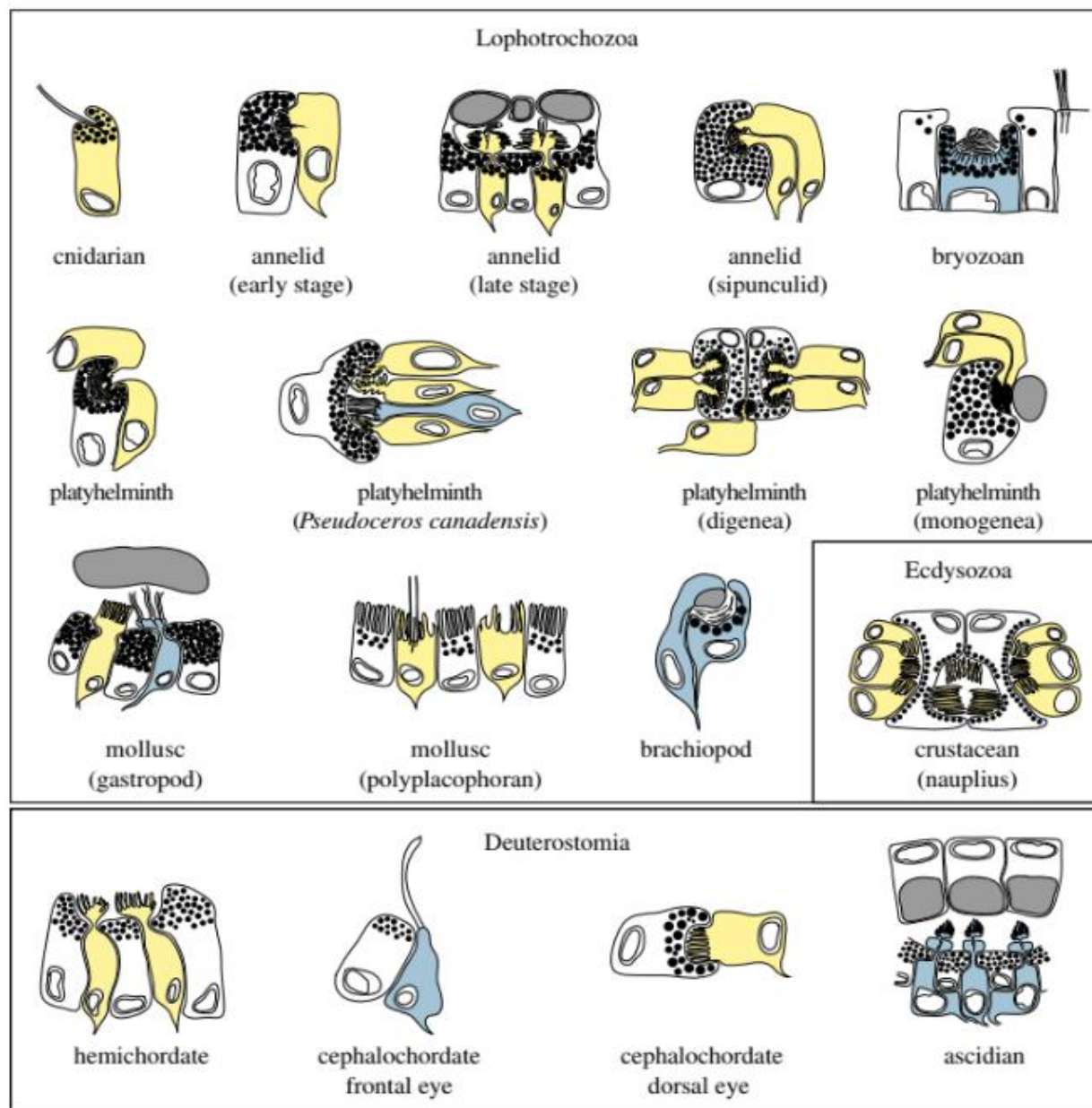
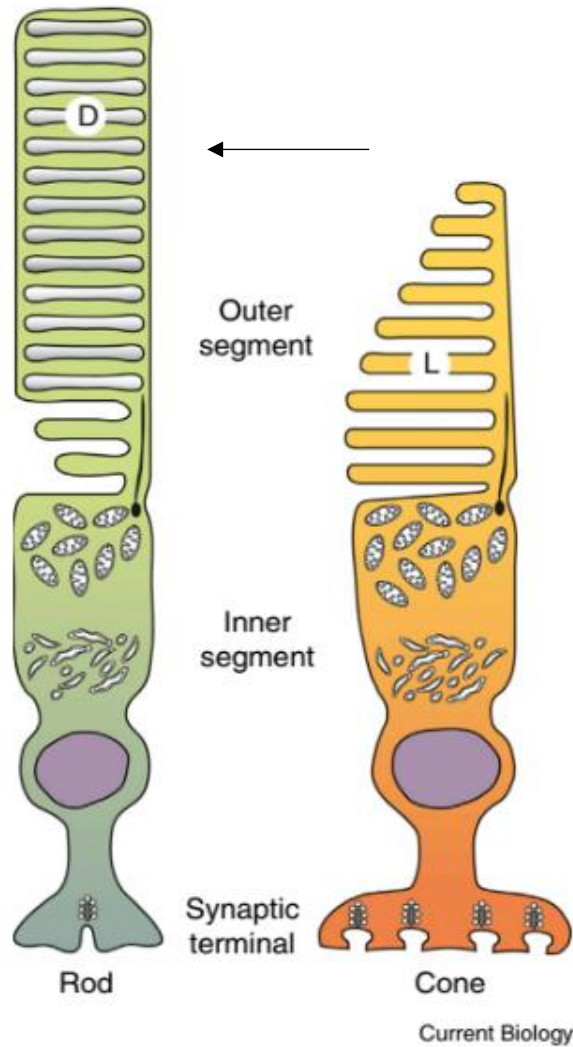


Figure 1. Diversity of simple eyes in planktonic larvae. Schematic drawings of simple eyes from marine invertebrate larvae. Rhabdomeric photoreceptors are shown in yellow, ciliary photoreceptors in blue, lenses in grey and pigment granules in black.



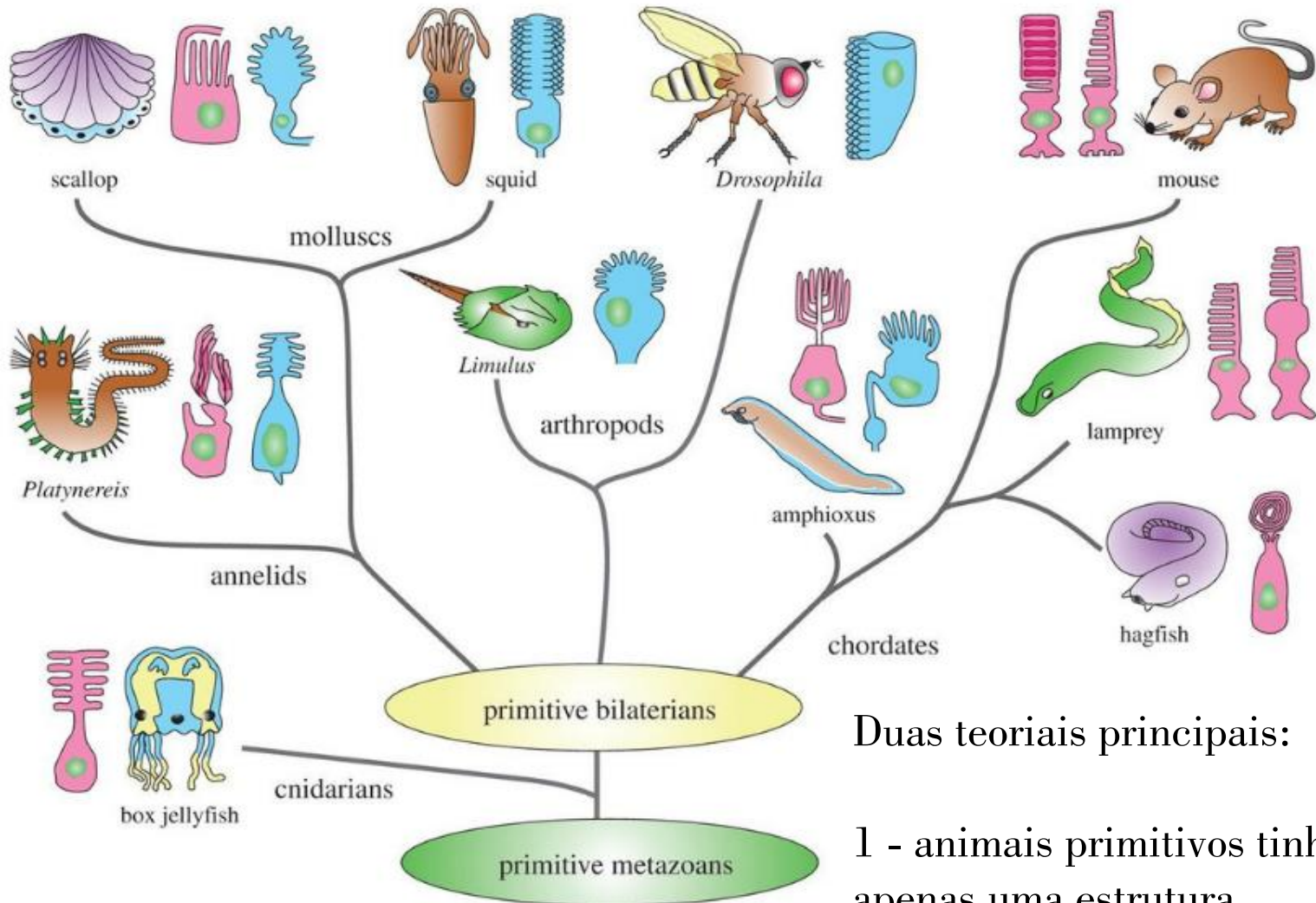
Cones e bastonetes



cone é fisiologicamente muito mais parecido com um bastonete

Figure 2. The morphology of rods (left) and cones (right) in modern jawed vertebrates. Note the outer segment lamellae (L) and internalised cytosolic disks (D) characteristic of cones and rods, respectively. After [12].

Histórico das células fotorreceptoras



Duas teorias principais:

1 - animais primitivos tinham apenas uma estrutura

2 – presença de estruturas ciliares, rabdoméricas e células pigmentares

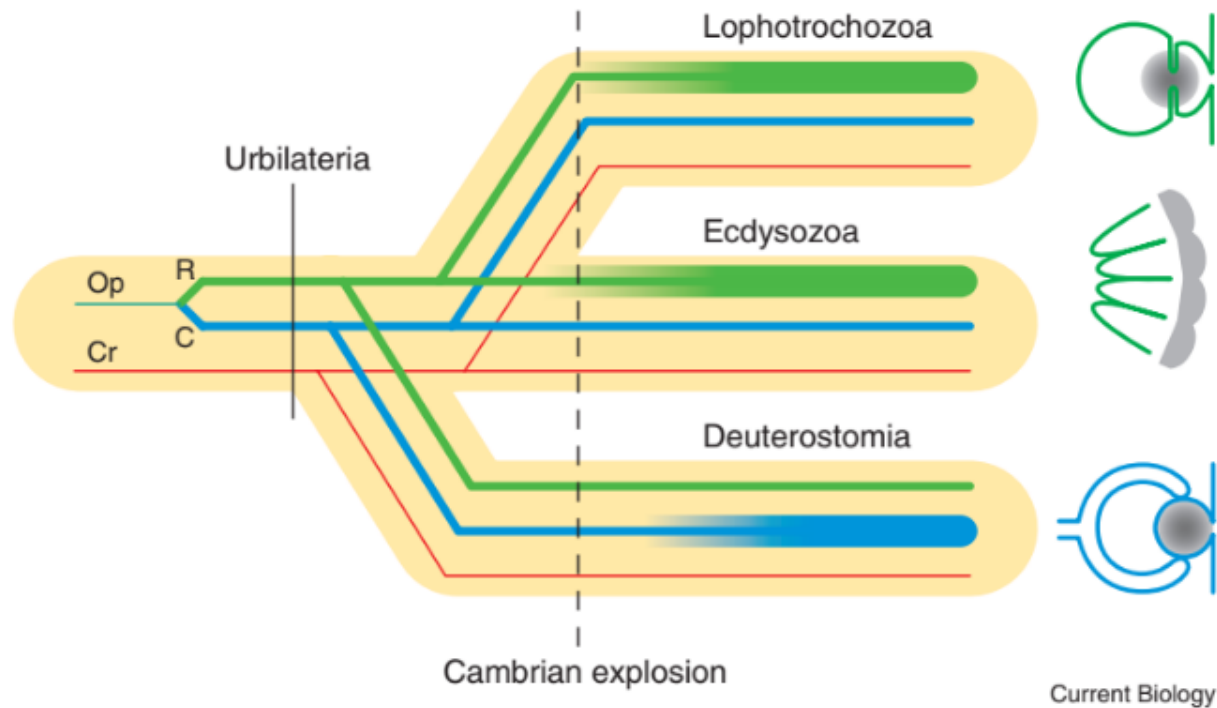


Figure 1. A phylogenetic tree of photoreceptive systems in the main branches of Bilateria.

Então porque considerar uma origem diferente?

Origem independente

MORFOLÓGICO

Origem comum - ressalvas

FISIOLOGICO

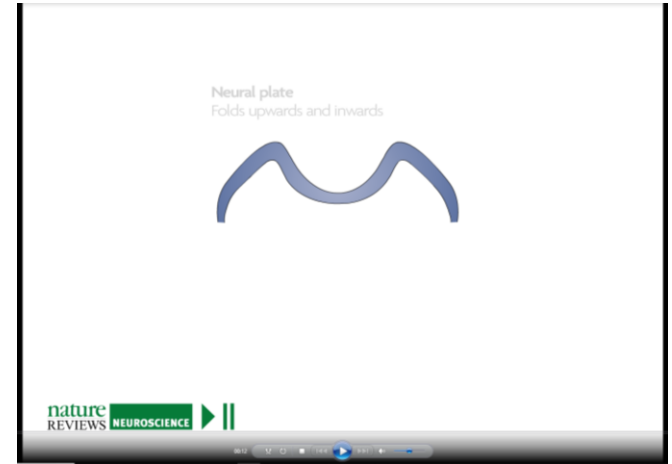
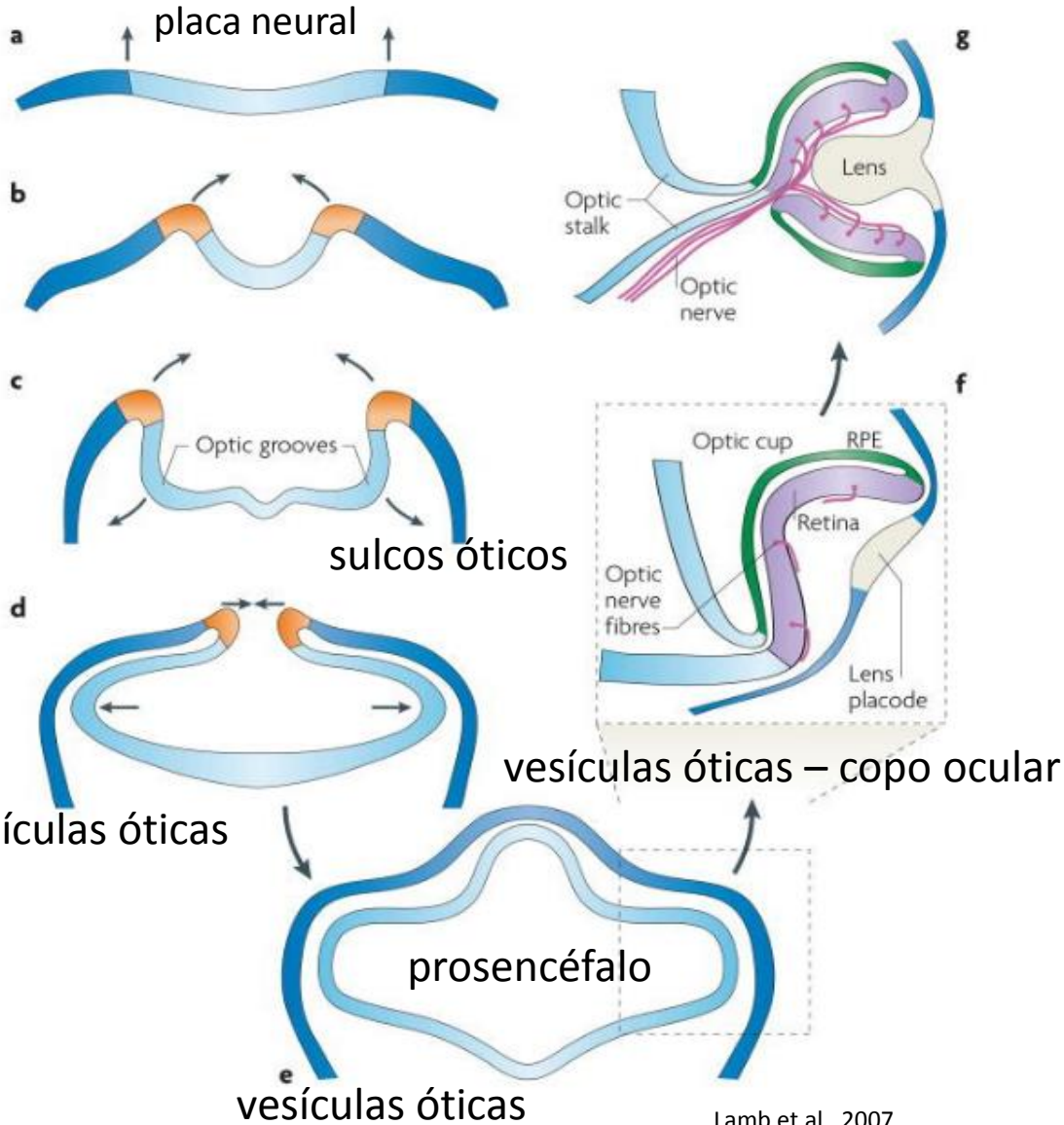
?

EMBRIONÁRIO



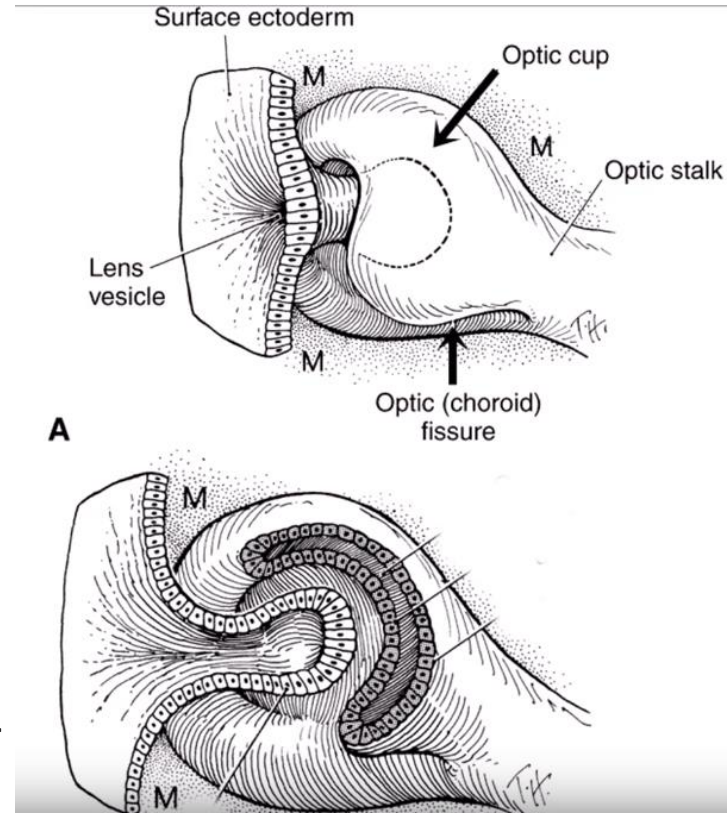
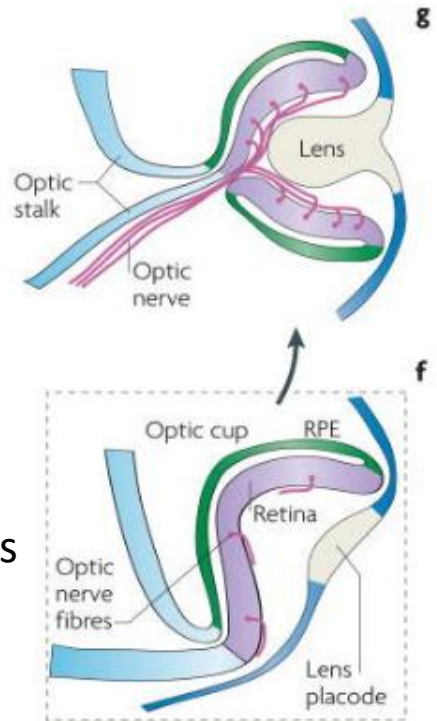
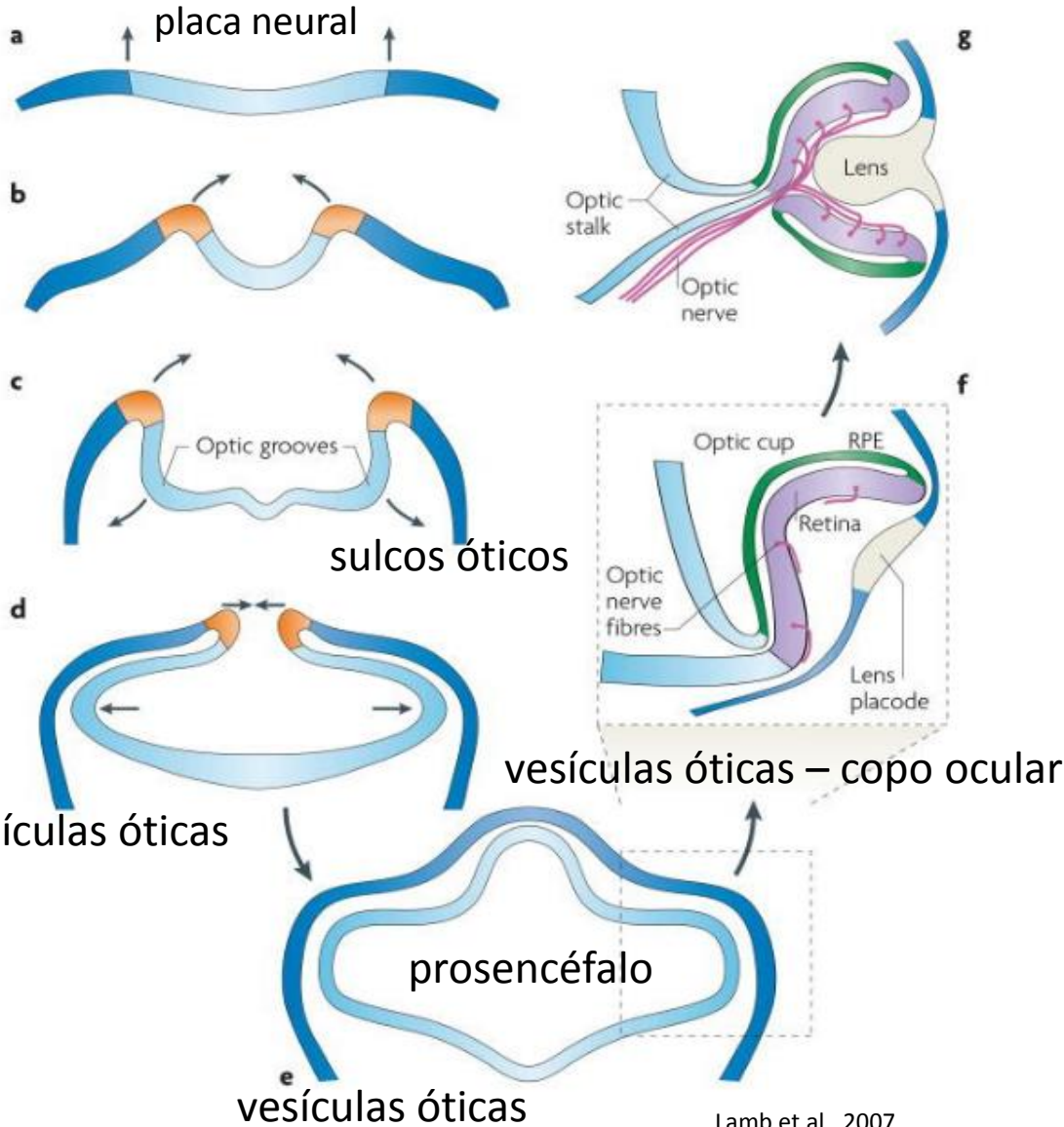
EMBRIÓNARIO

Vertebrados

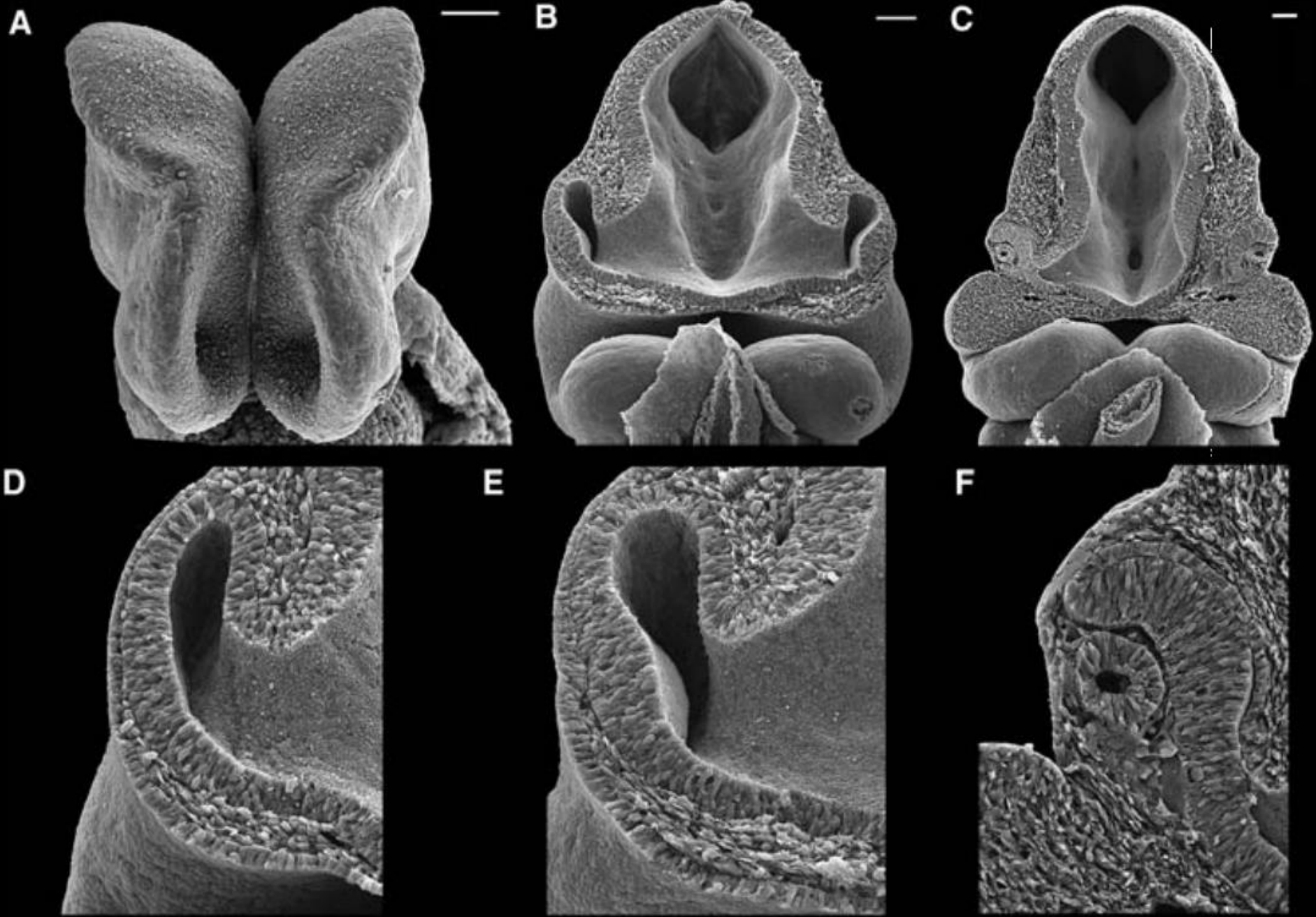


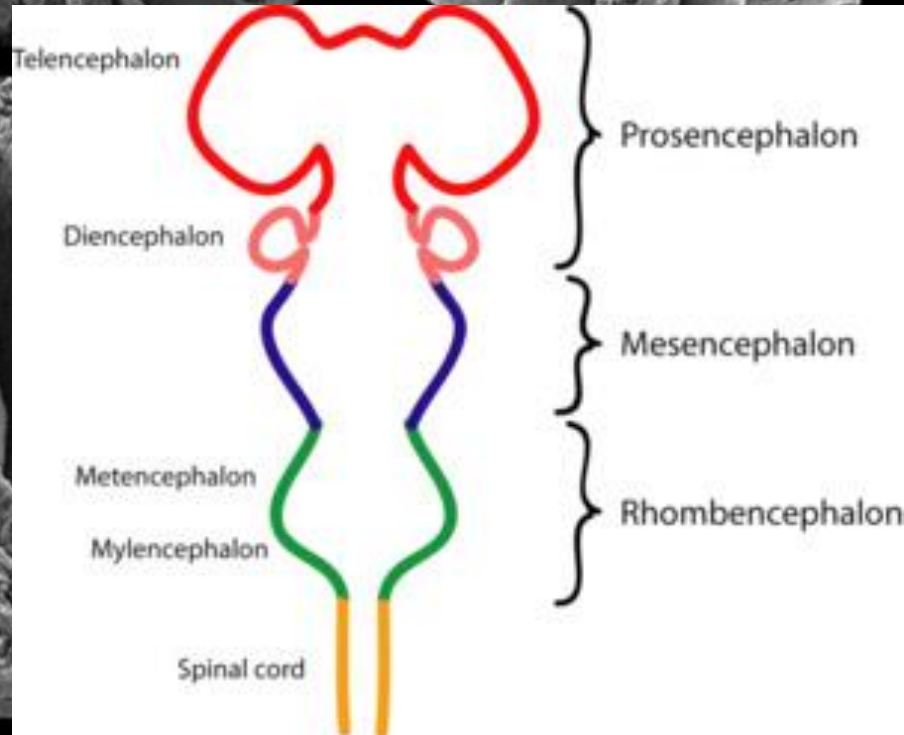
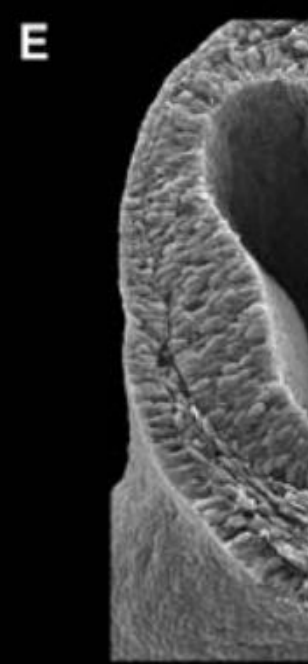
<https://www.youtube.com/watch?v=C4fPBYvx17Y>

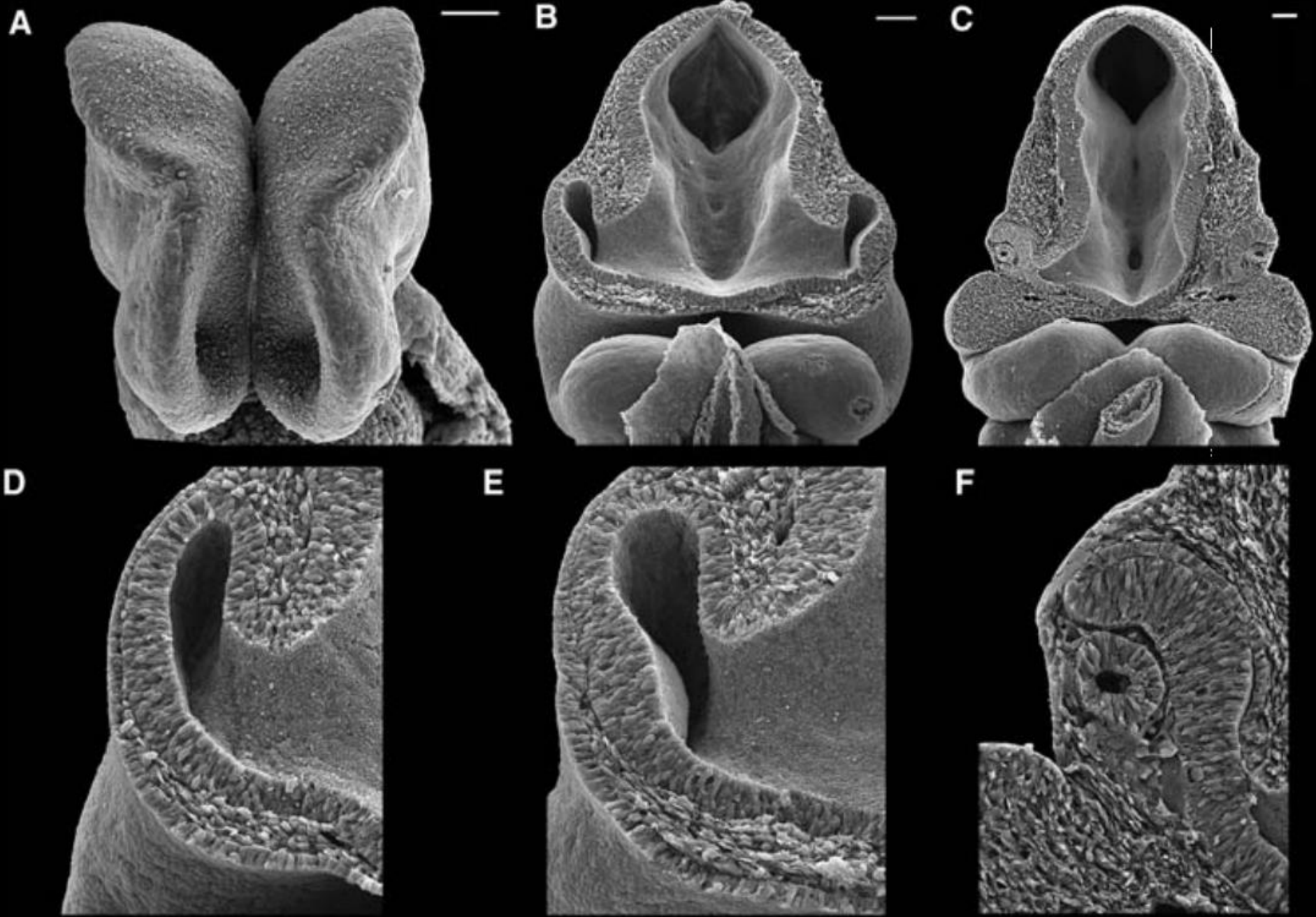
Vertebrados

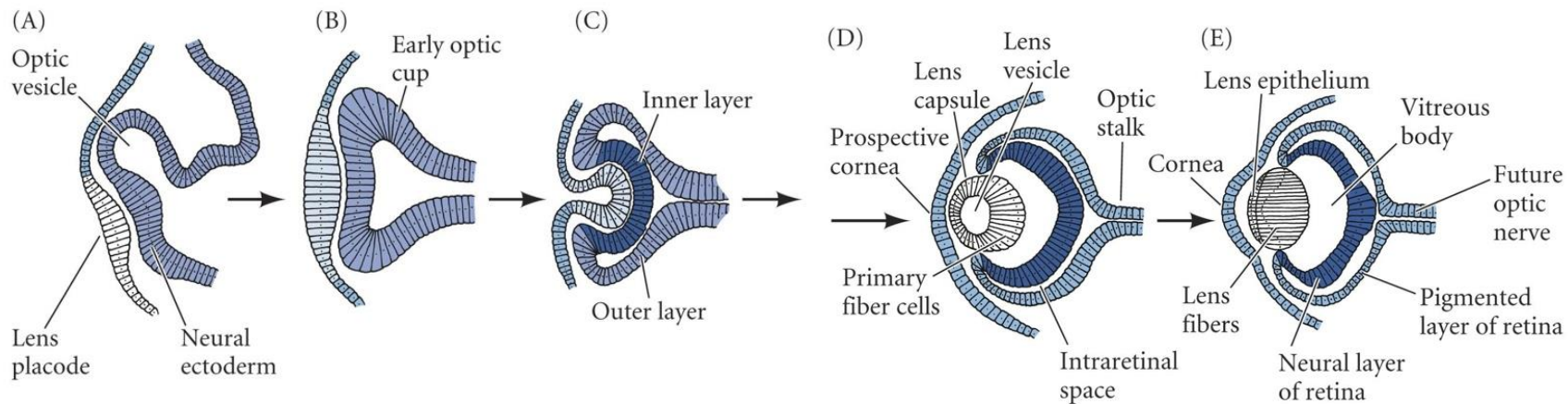


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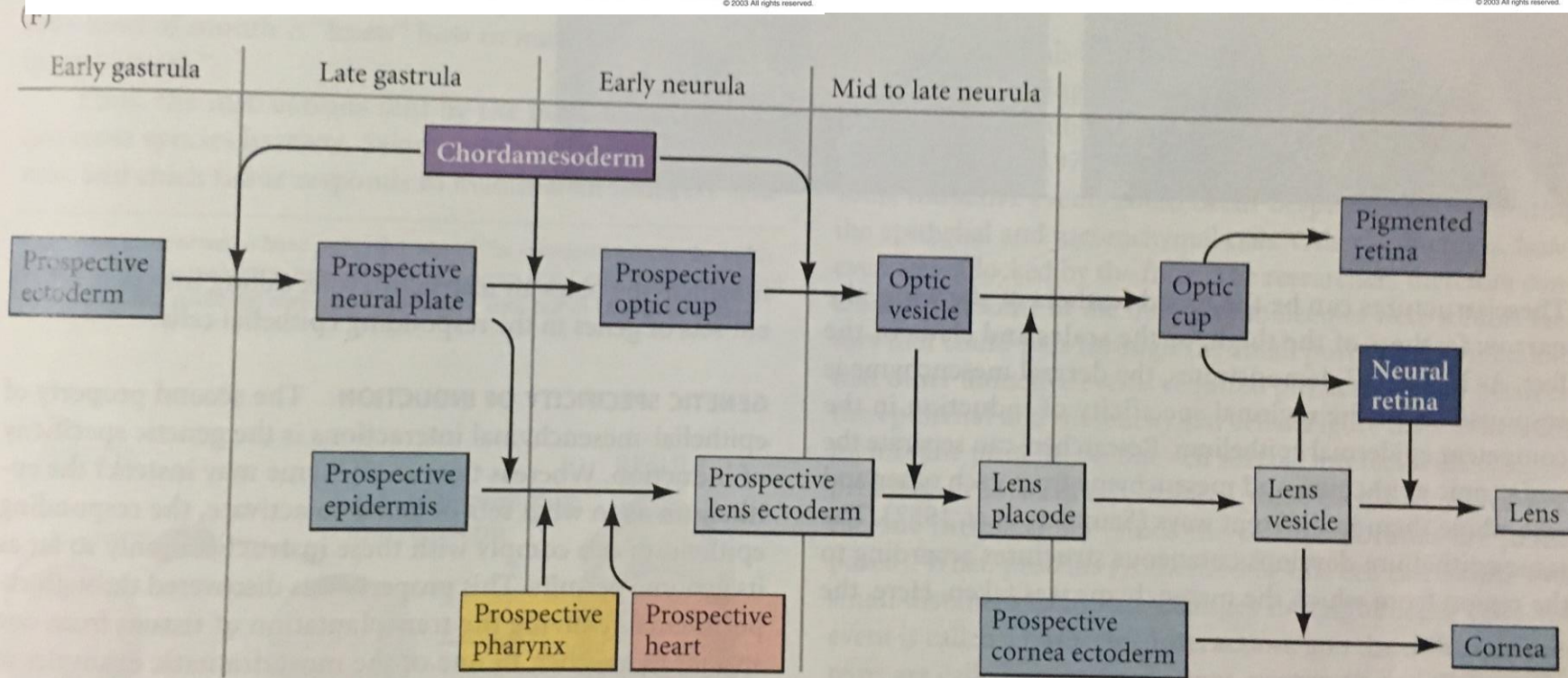






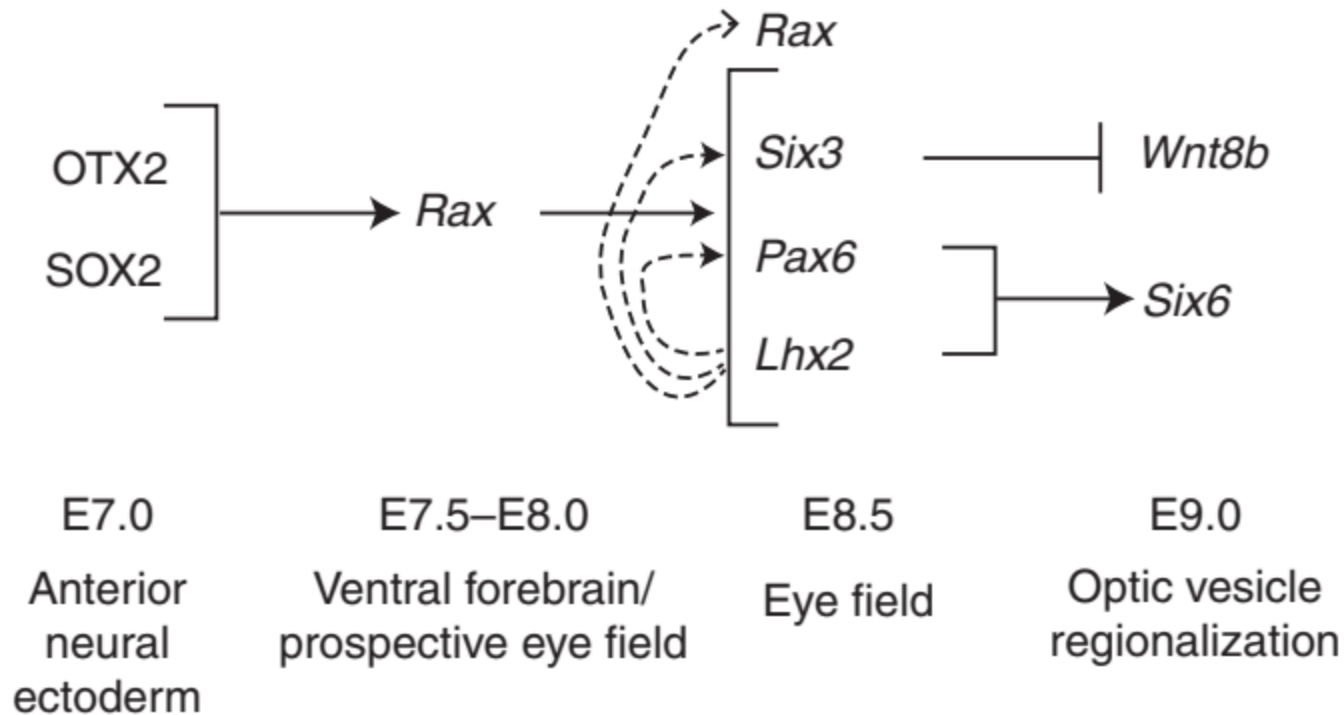
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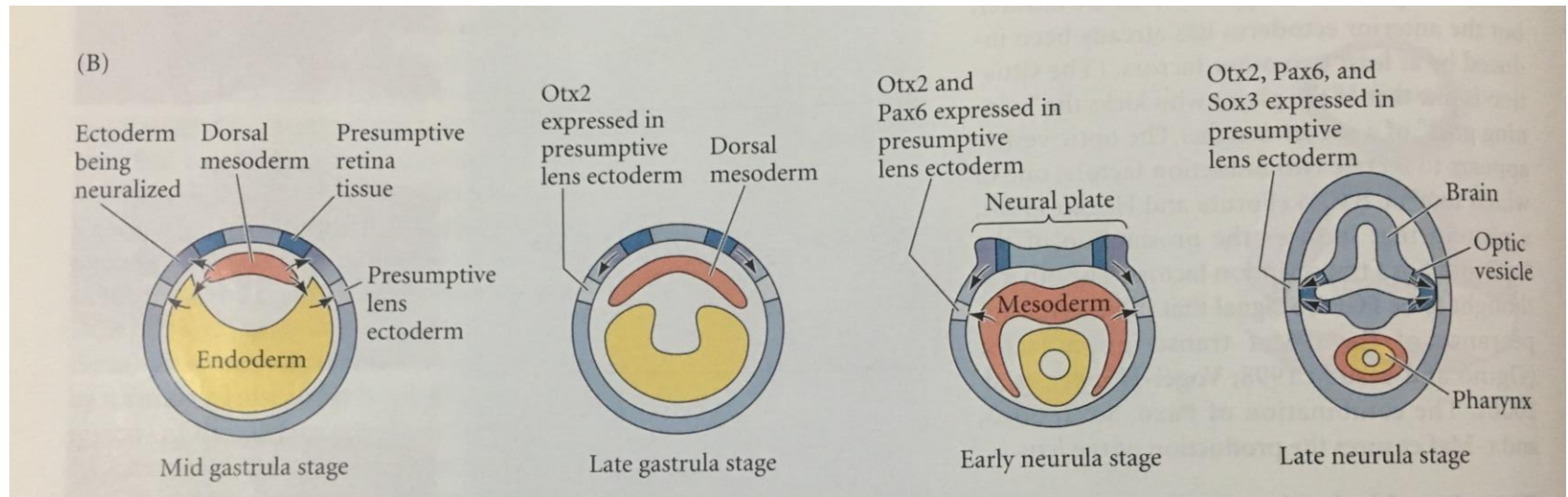
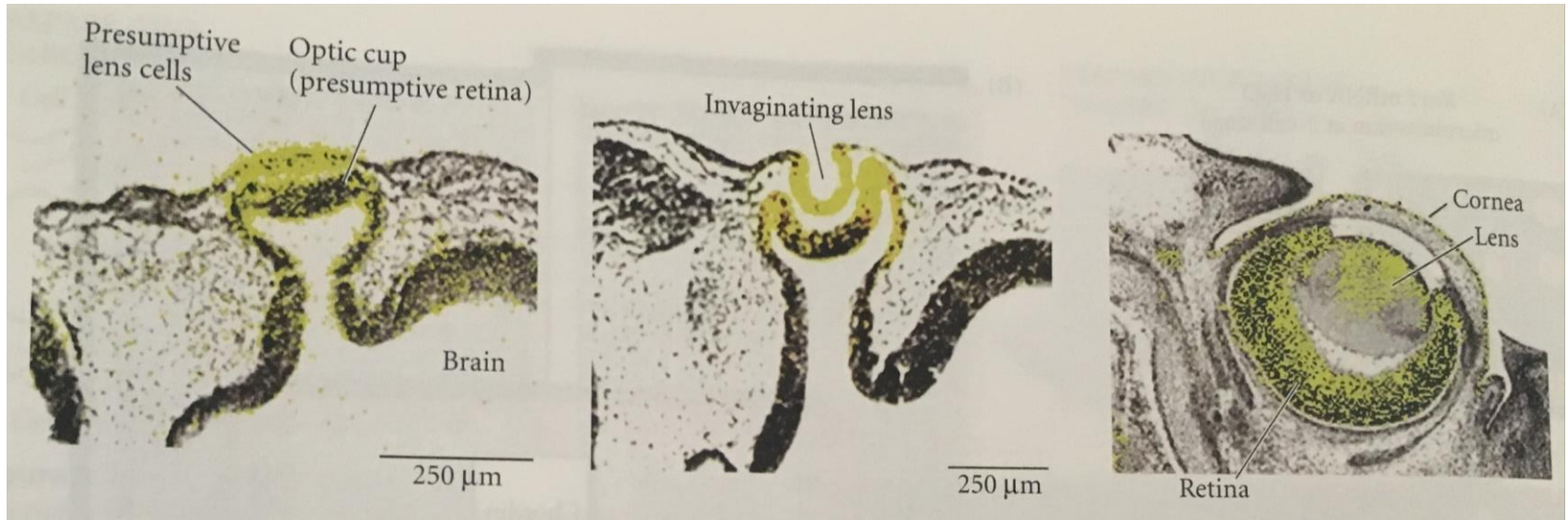


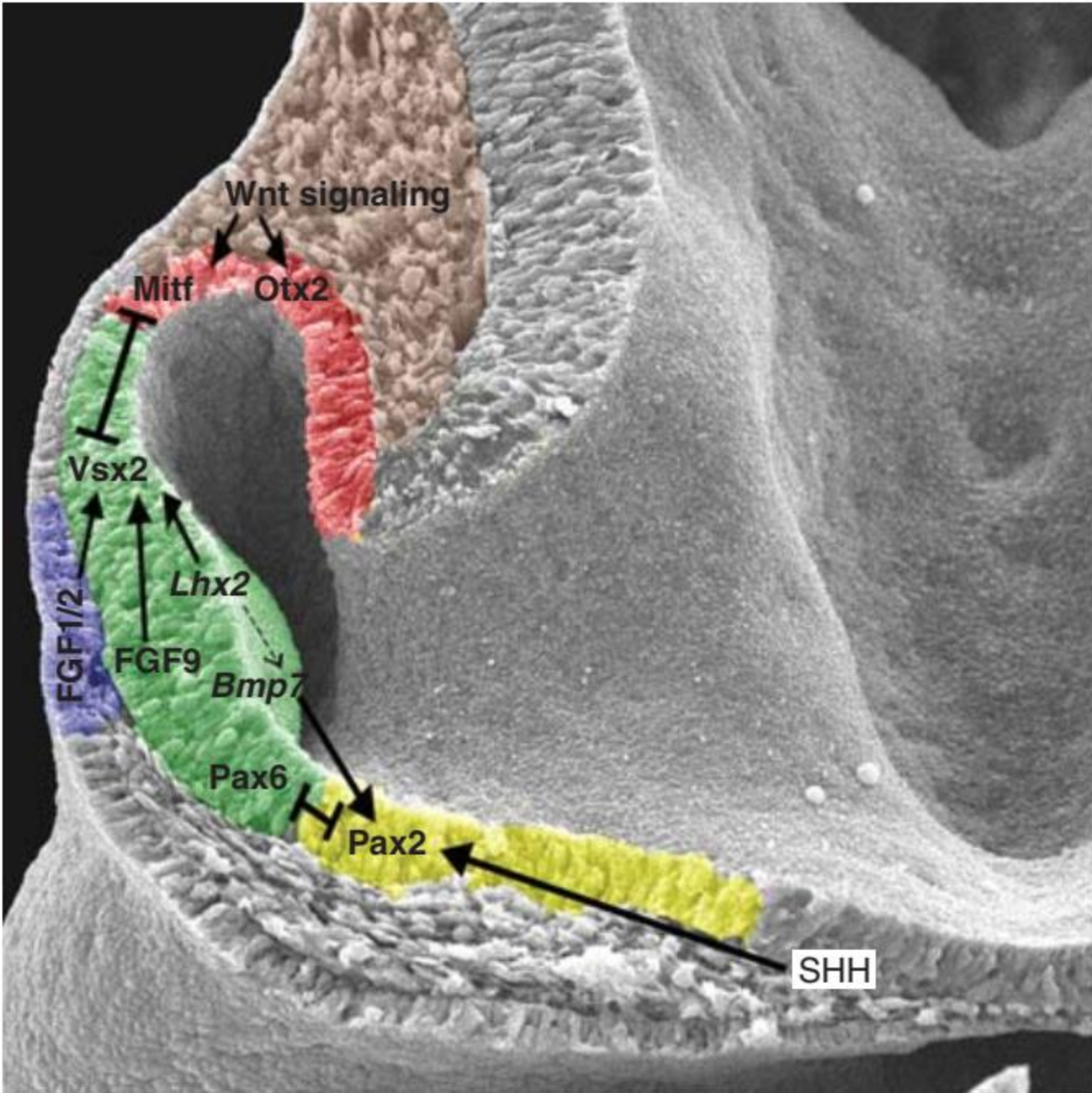
Expressão Genética

Fatores de transcrição



Indução progressiva





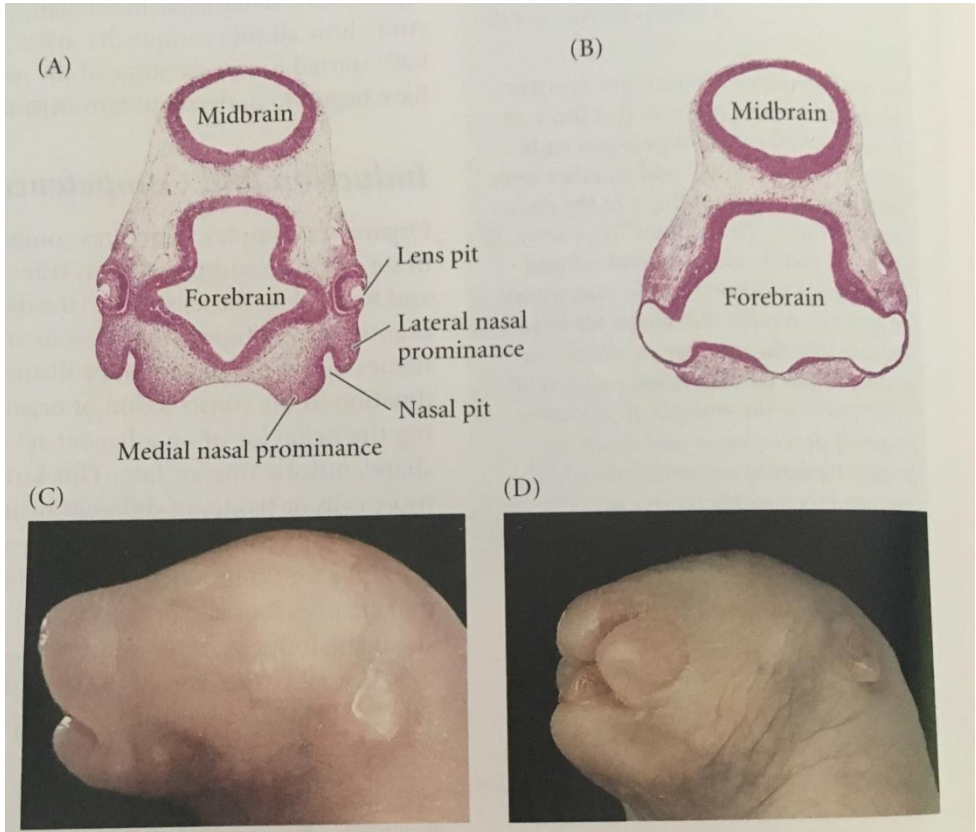
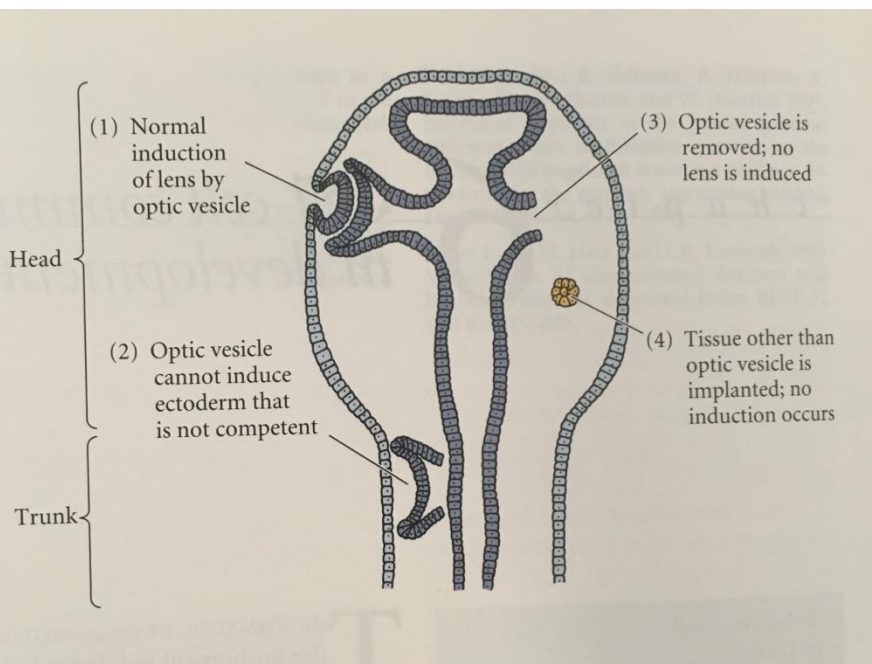
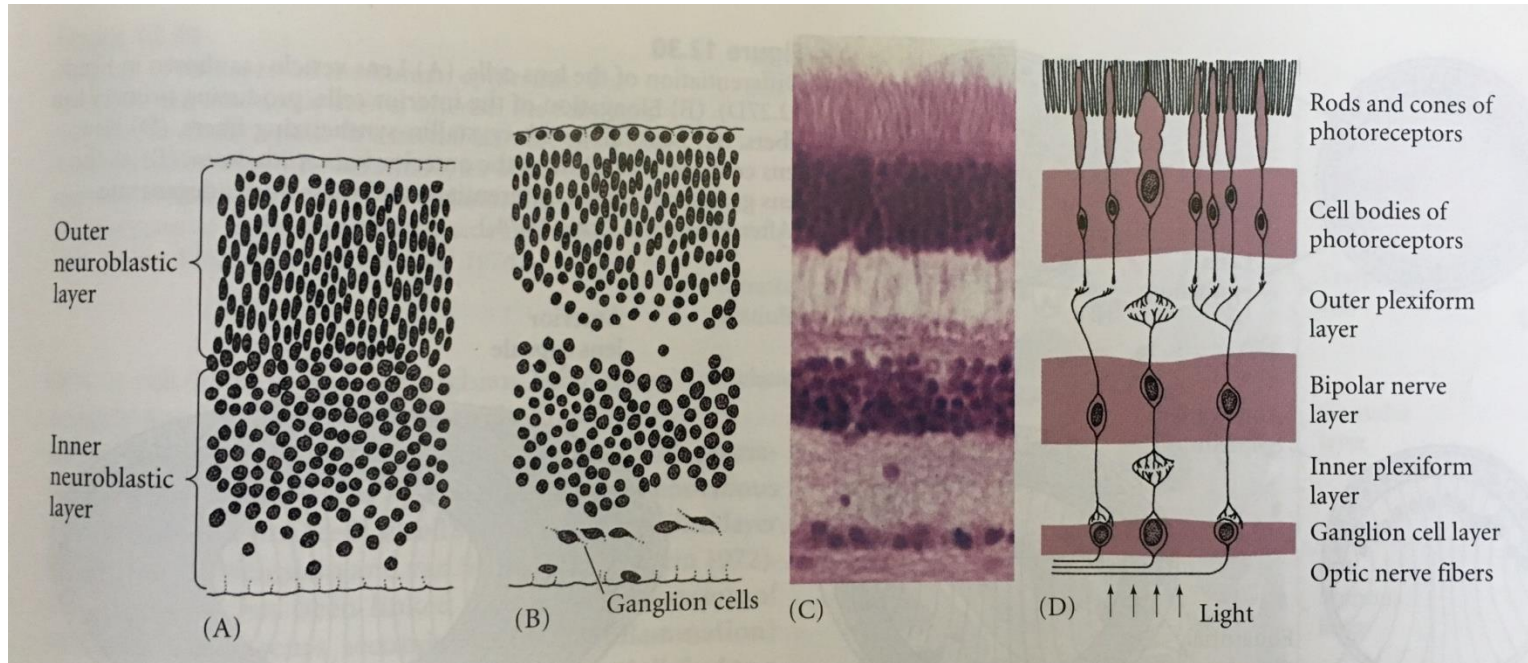
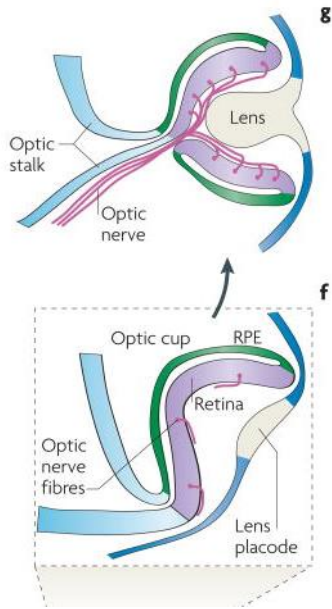
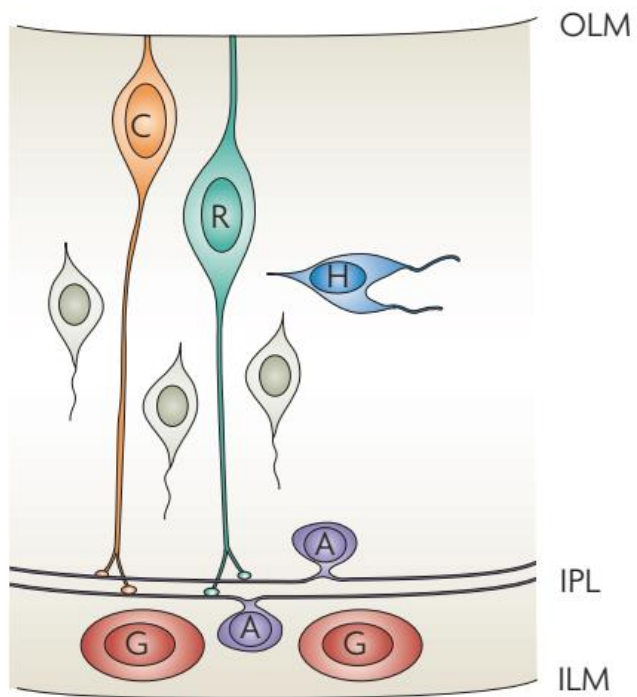


Figure 6.2 Induction of optic and nasal structures by Pax6 in the rat embryo. (A, B) Histology of wild-type (A) and homozygous *Pax6* mutant (B) embryos at day 12 of gestation shows induction of lenses and retinal development in the wild-type embryo, but not in the mutant. Similarly, neither the nasal pit nor the medial nasal prominence is induced in the mutant rats. (C) Newborn wild-type rats show prominent nose as well as (closed) eyes. (D) Newborn *Pax6* mutant rats show neither eyes nor nose. (From Fujiwara et al. 1994; photographs courtesy of M. Fujiwara.)

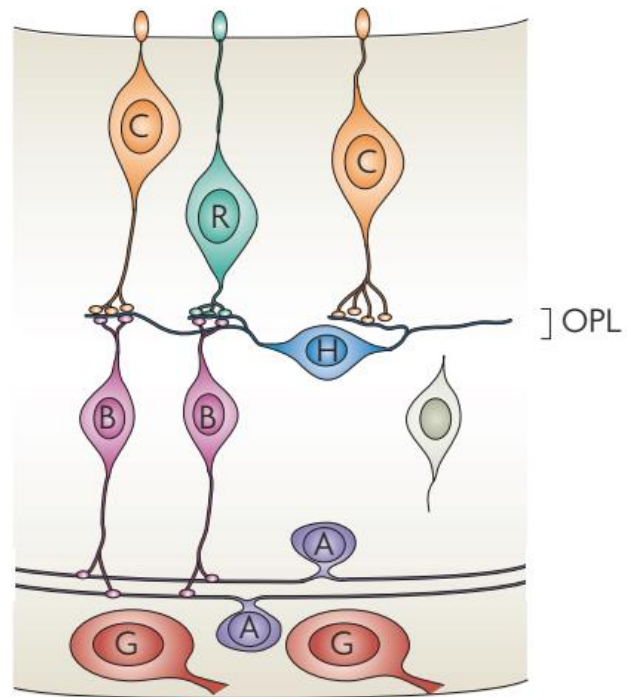
Formação da retina e células fotorreceptoras



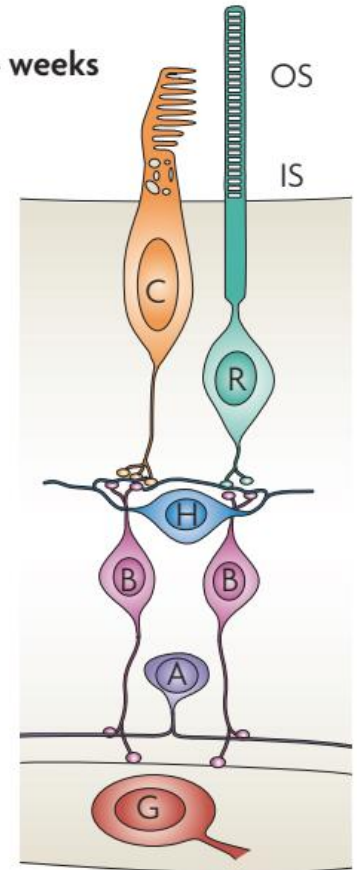
c At birth



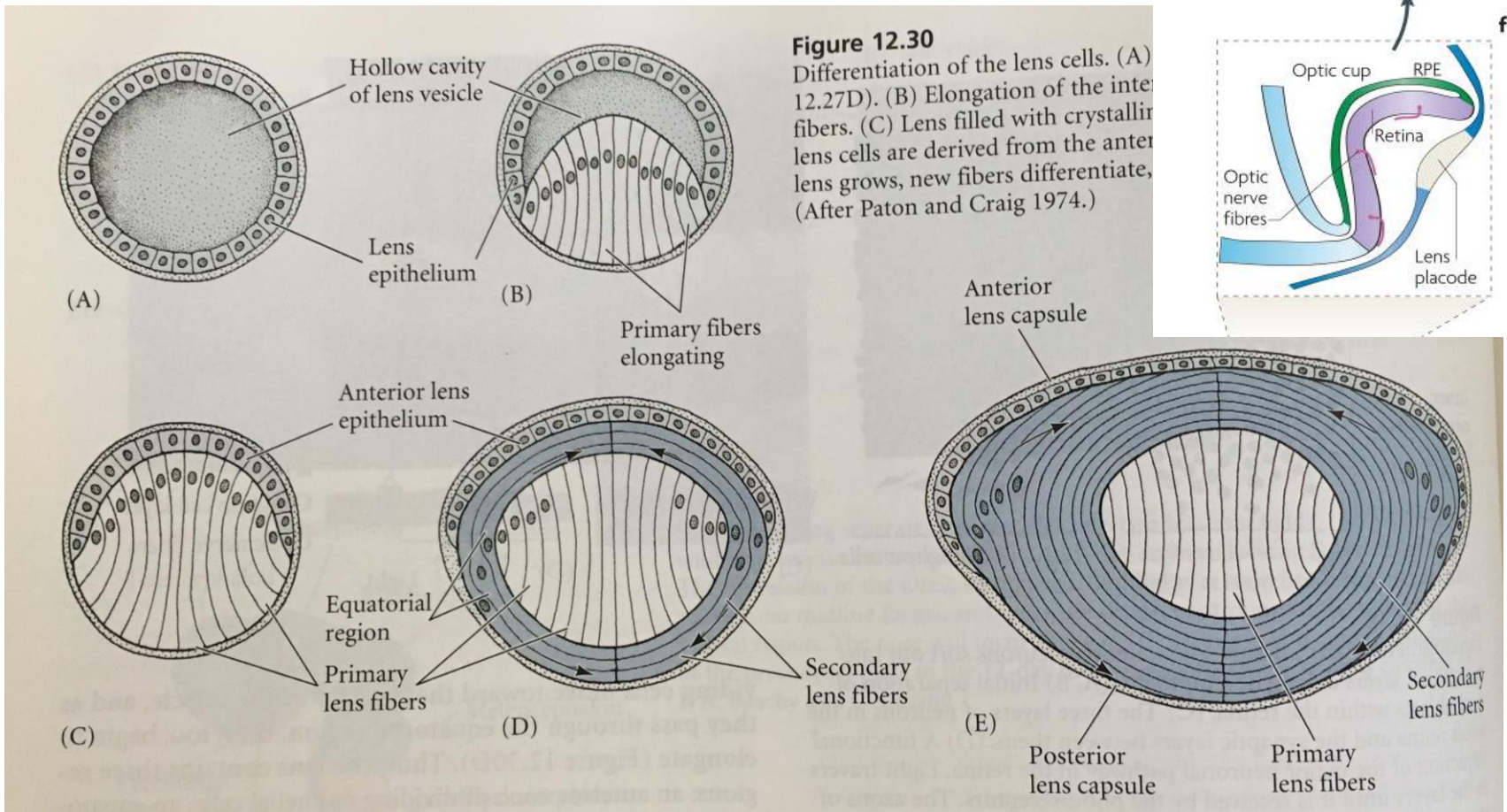
d 2 weeks



e 4 weeks



Formação da lente



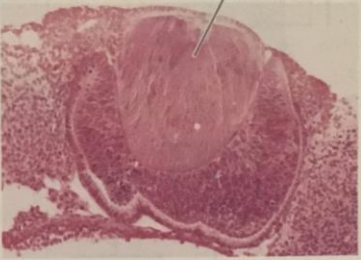
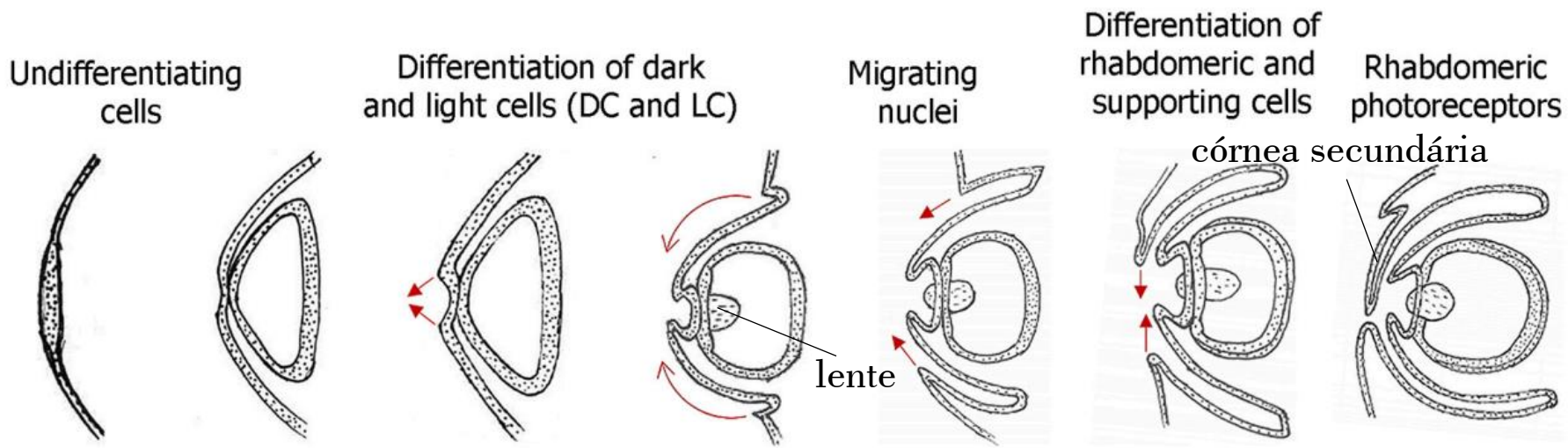
Optic vesicles	Surface ectoderm	Lens induction	
Wild-type	Wild-type	Yes	
<i>Pax6</i> ⁻ / <i>Pax6</i> ⁻	Wild-type		?
Wild-type	<i>Pax6</i> ⁻ / <i>Pax6</i> ⁻		?
<i>Pax6</i> ⁻ / <i>Pax6</i> ⁻	<i>Pax6</i> ⁻ / <i>Pax6</i> ⁻		?

Figure 6.3

Recombination experiments show that the induction deficiency of *Pax6*-deficient rats is caused by the inability of the surface ectoderm to respond to the optic vesicle. (Photographs courtesy of M. Fujiwara.)

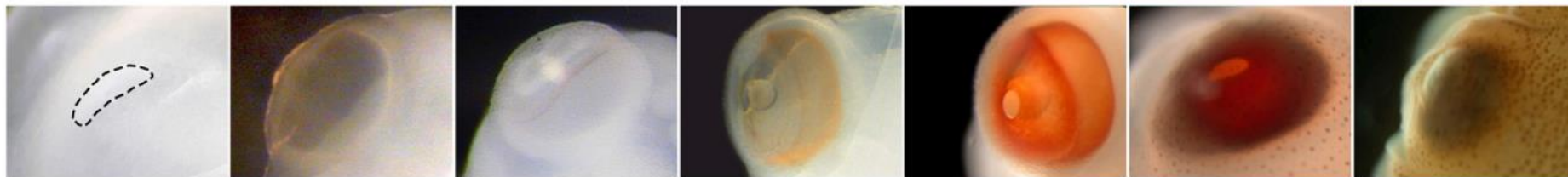
Cefalópodes

4 dobras ectodérmicas sucessivas



First reaction to light

Habituation and memorization



stage 18

stage 21

stage 23

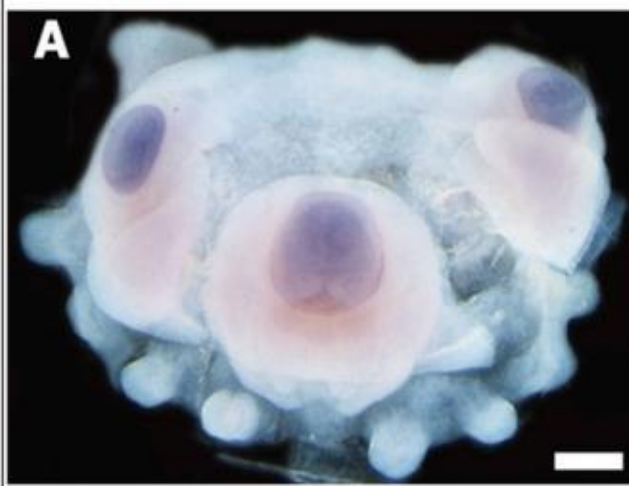
stage 25

stage 26

stage 28

stage 30

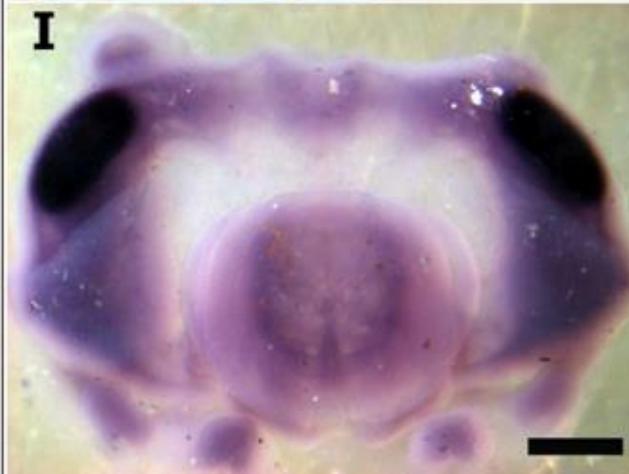
Sof-eya



Sof-dac

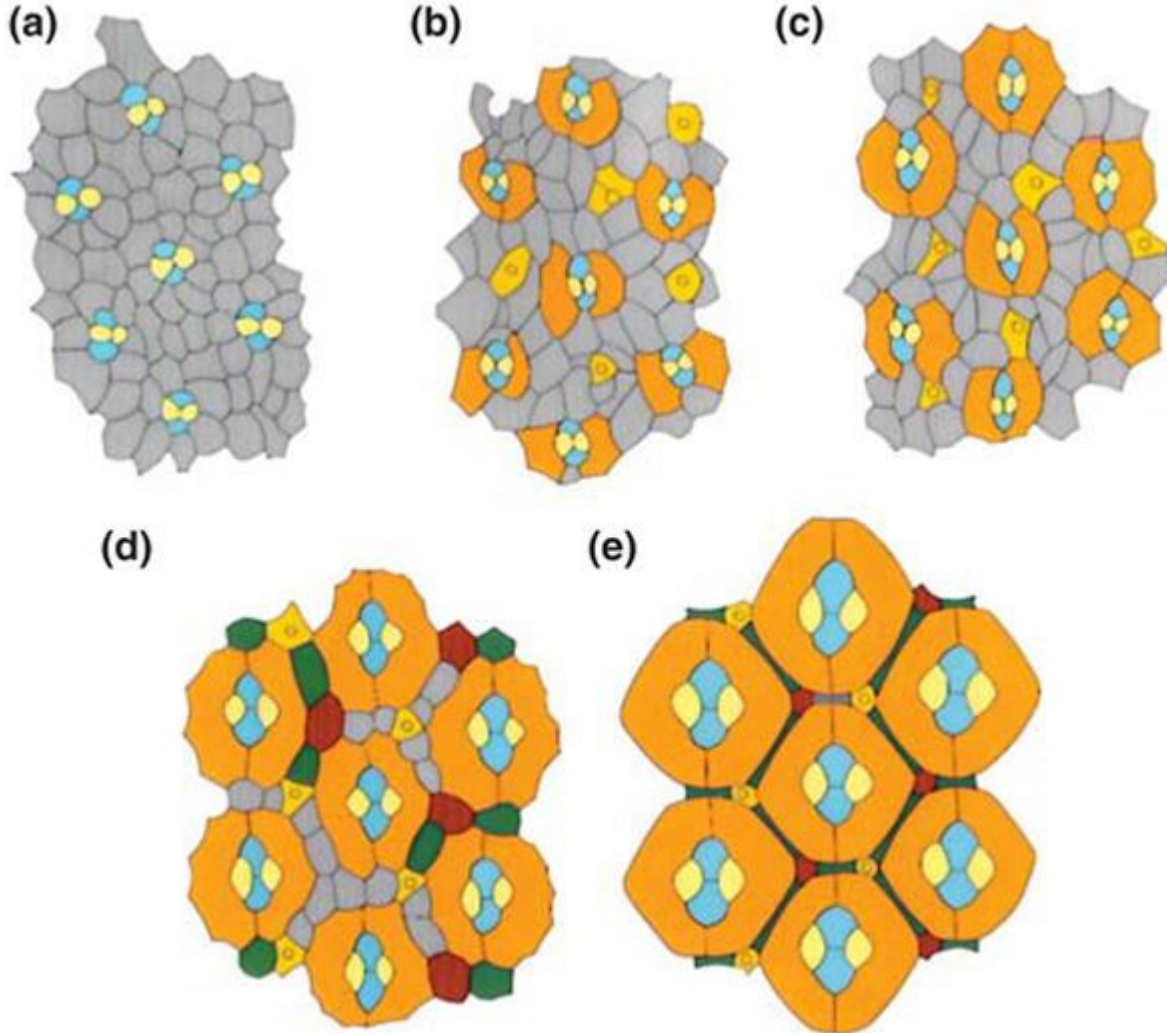


Sof-otx

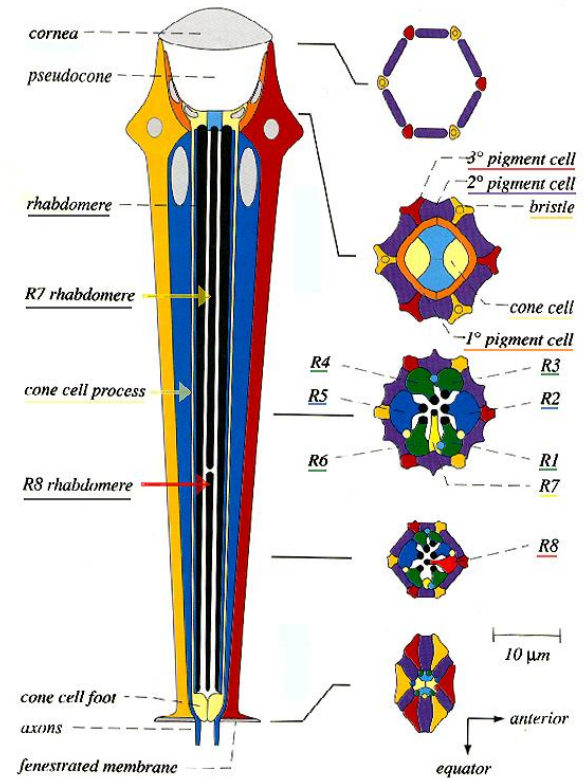


diferenciação dos neurônios fotorreceptores R8

- 20 células arranjadas
- 8 dessas células são fotorreceptores
- restantes são lentes

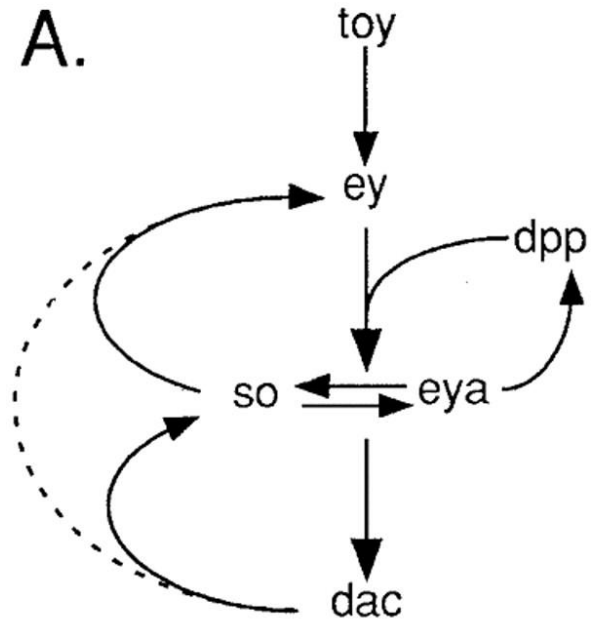


The Drosophila Adult Ommatidium

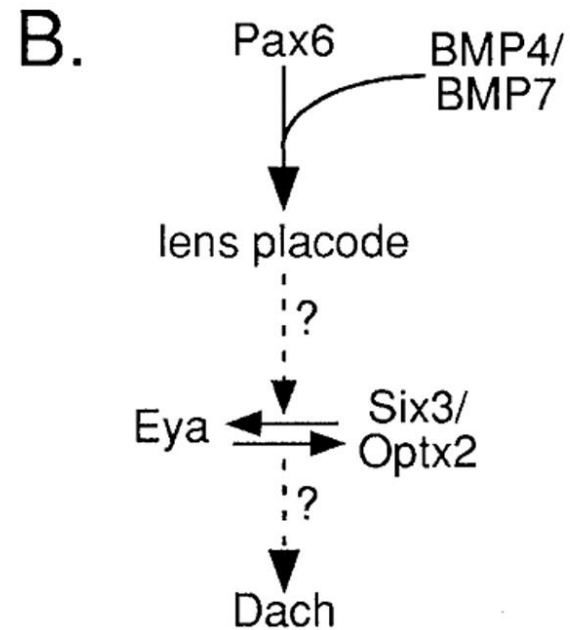


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Expressão Genética

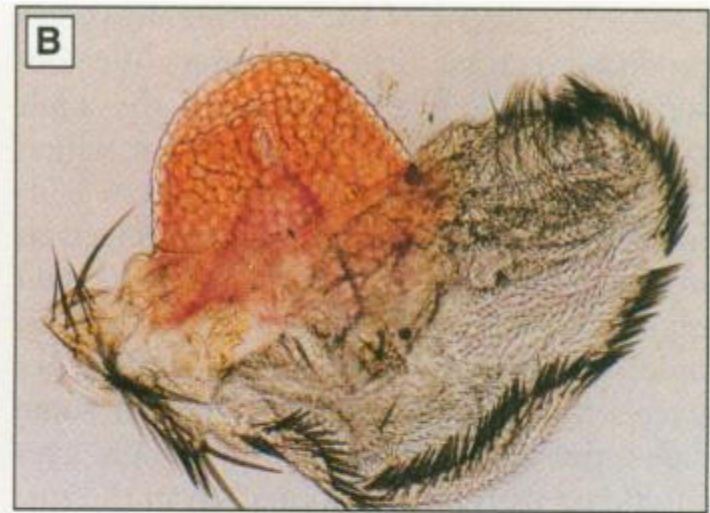


Drosophila

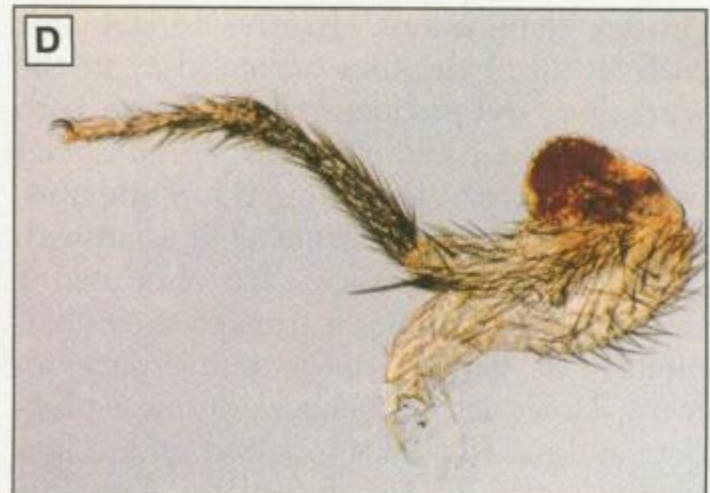


Mouse/Human

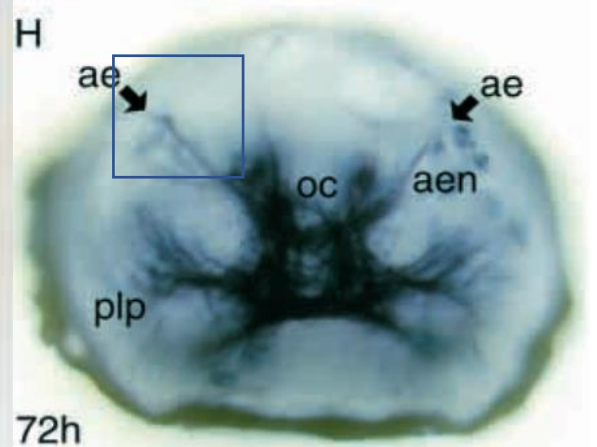
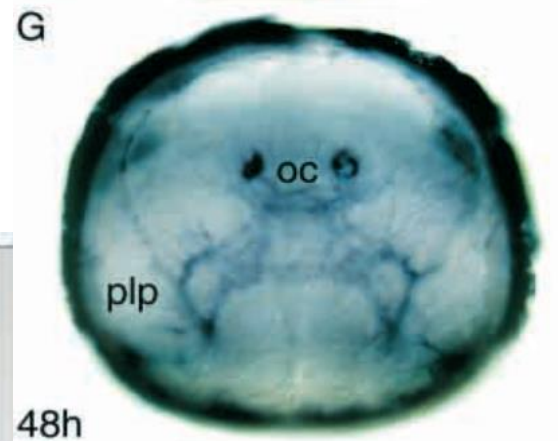
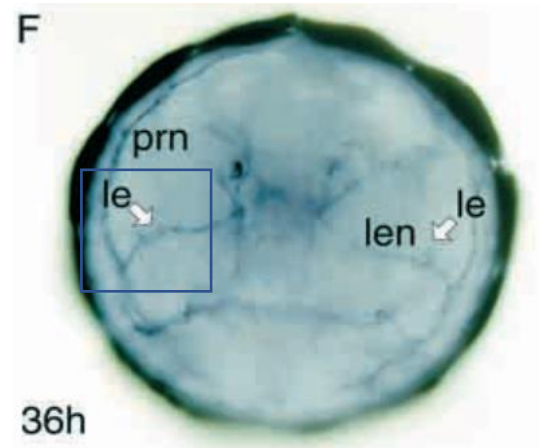
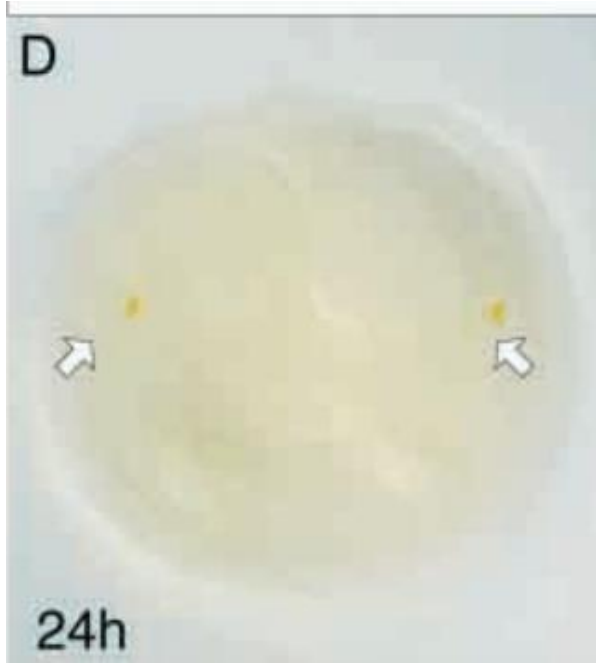
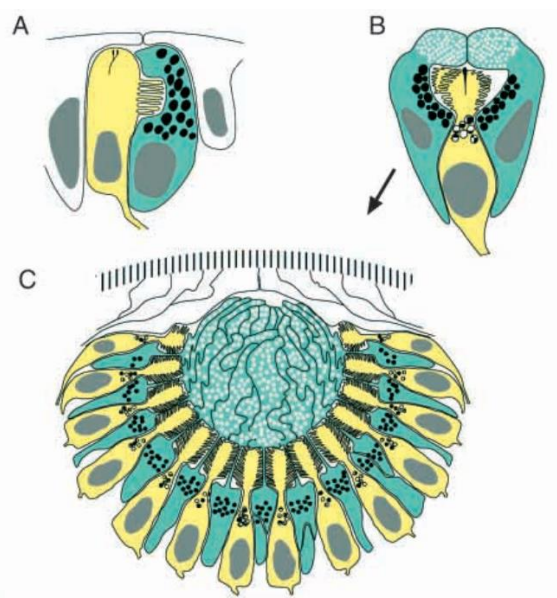
Expressão Genética



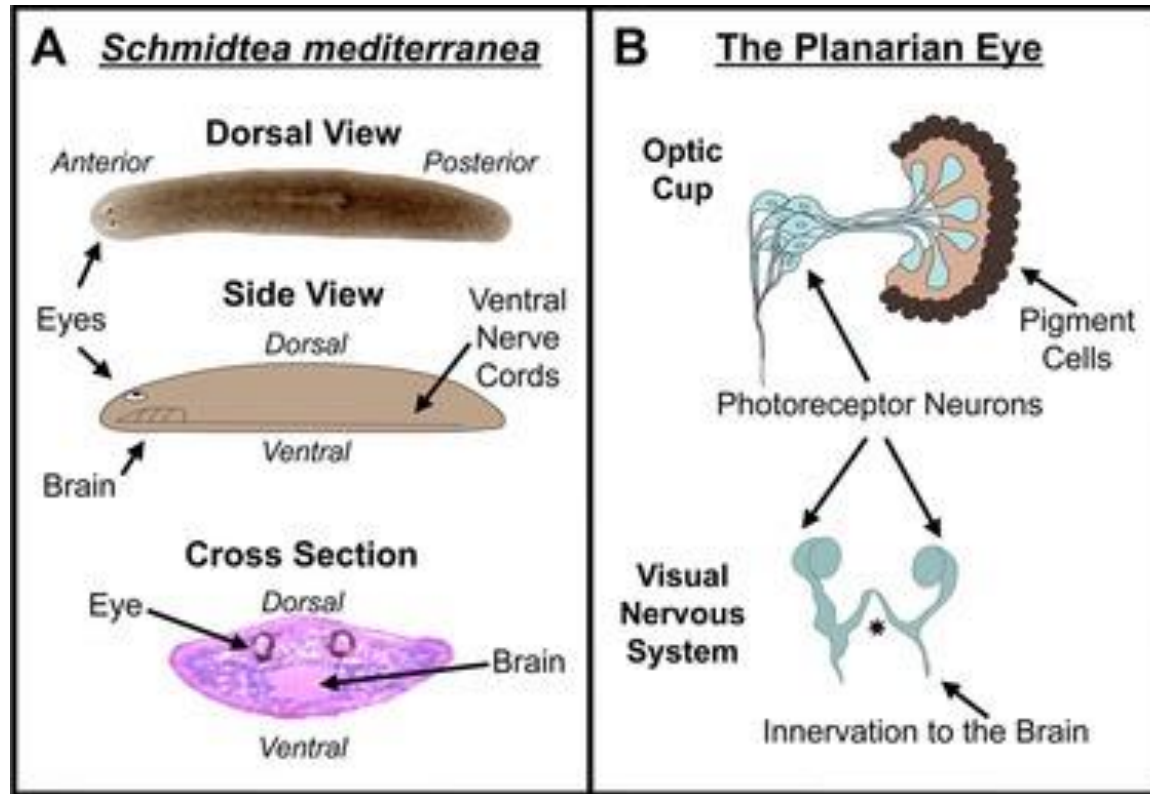
Eye less - ey



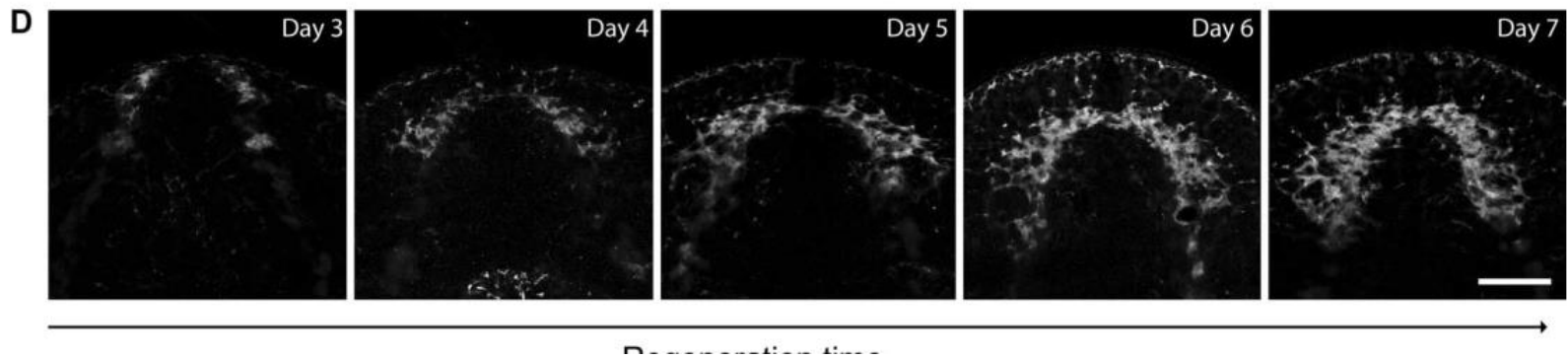
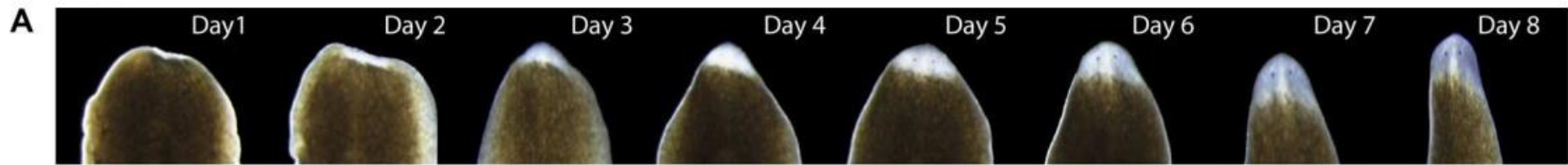
Poliqueta

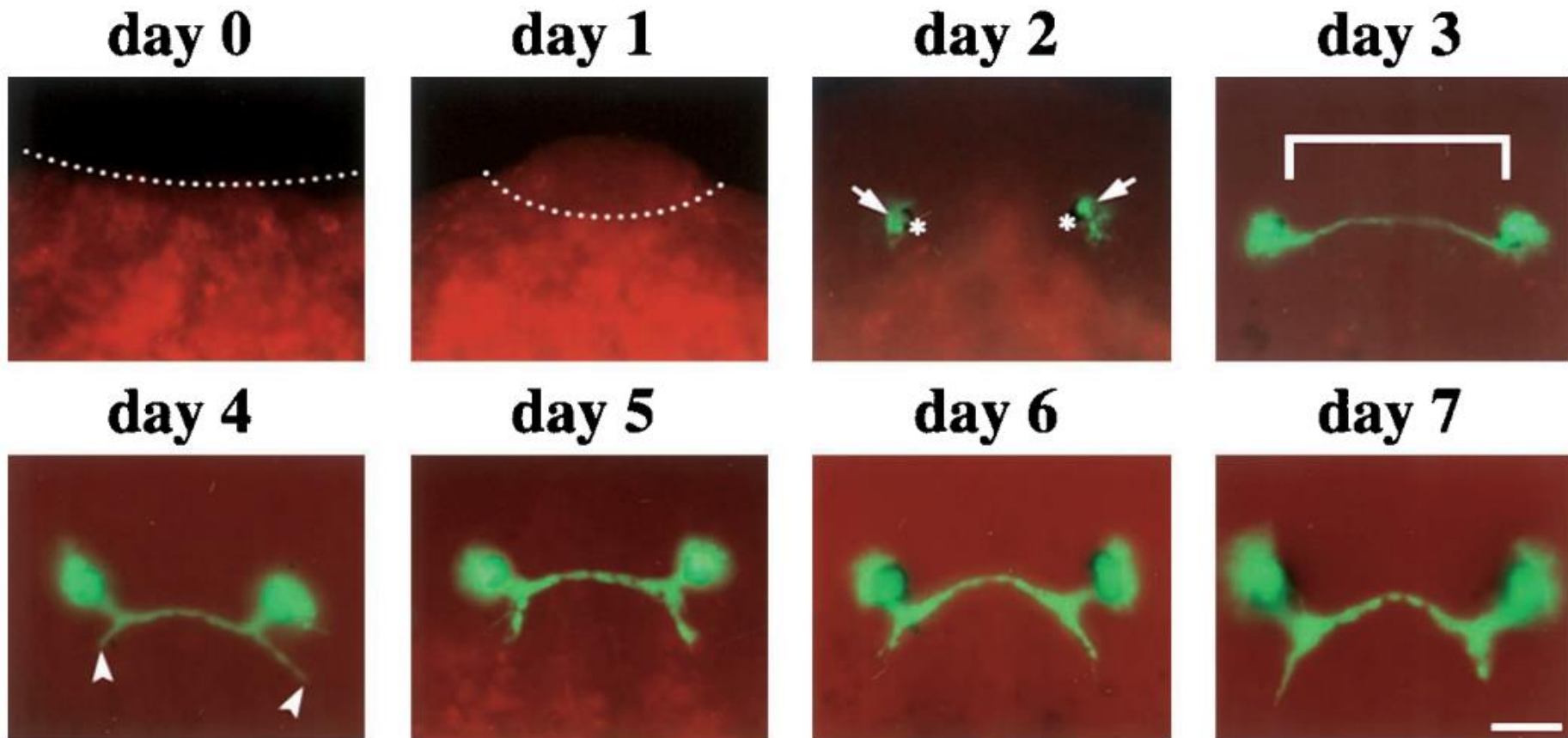


Planária

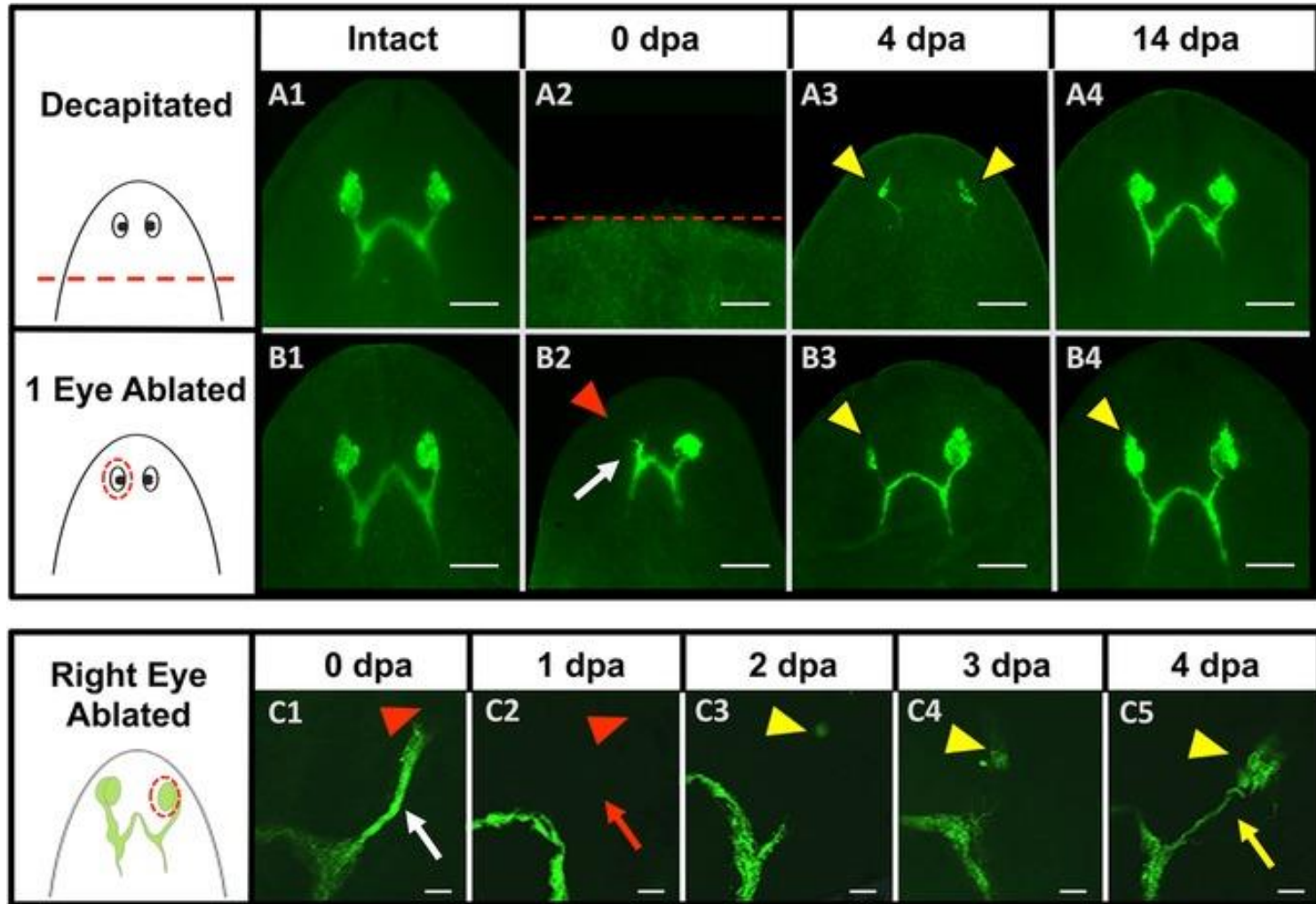


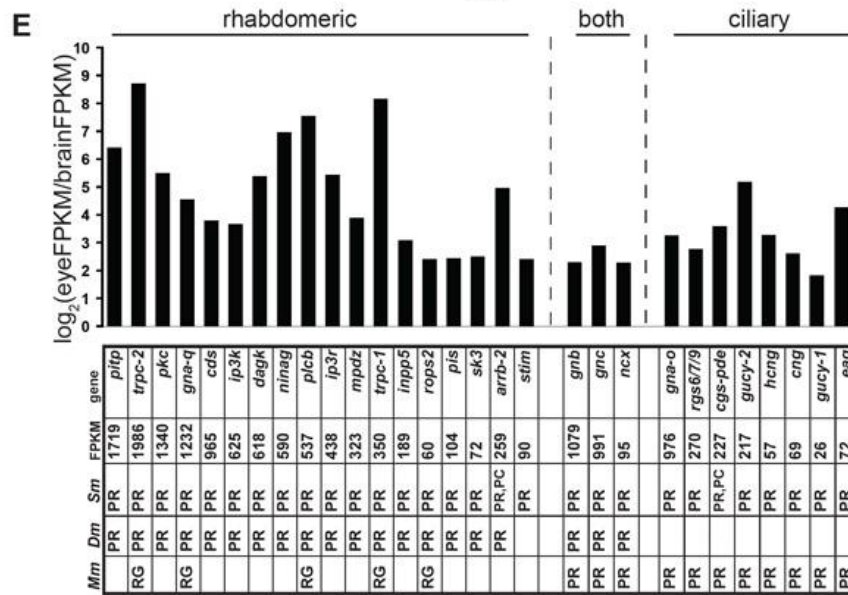
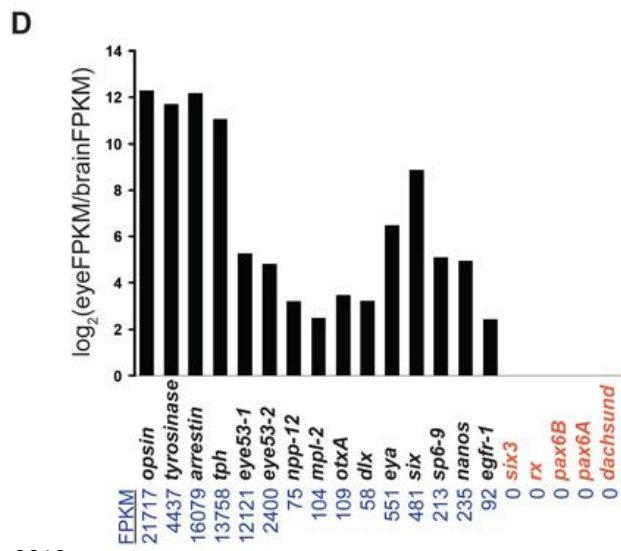
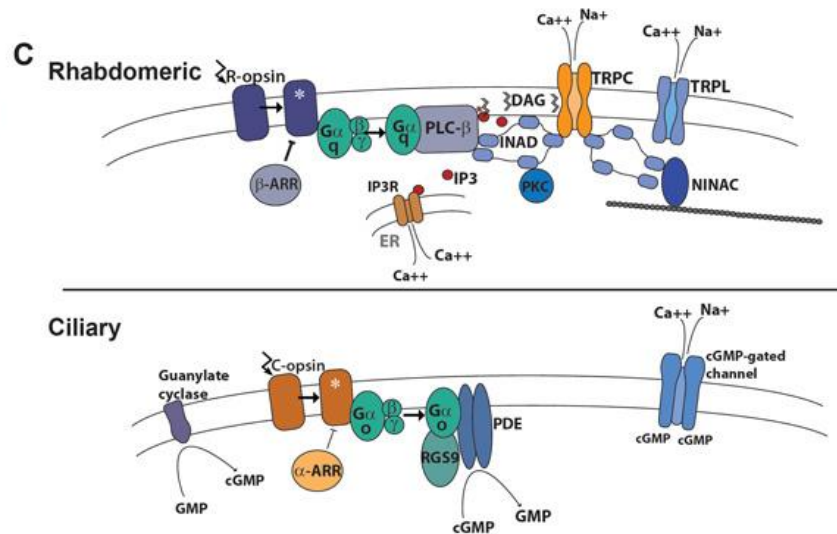
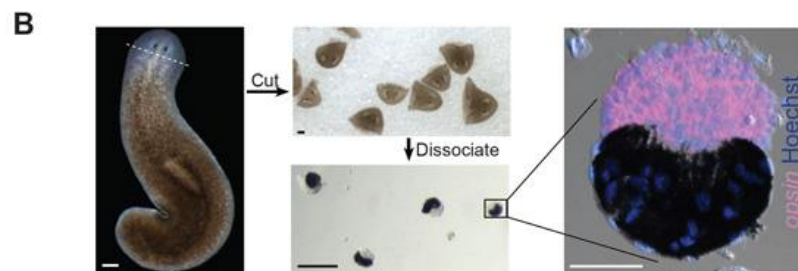
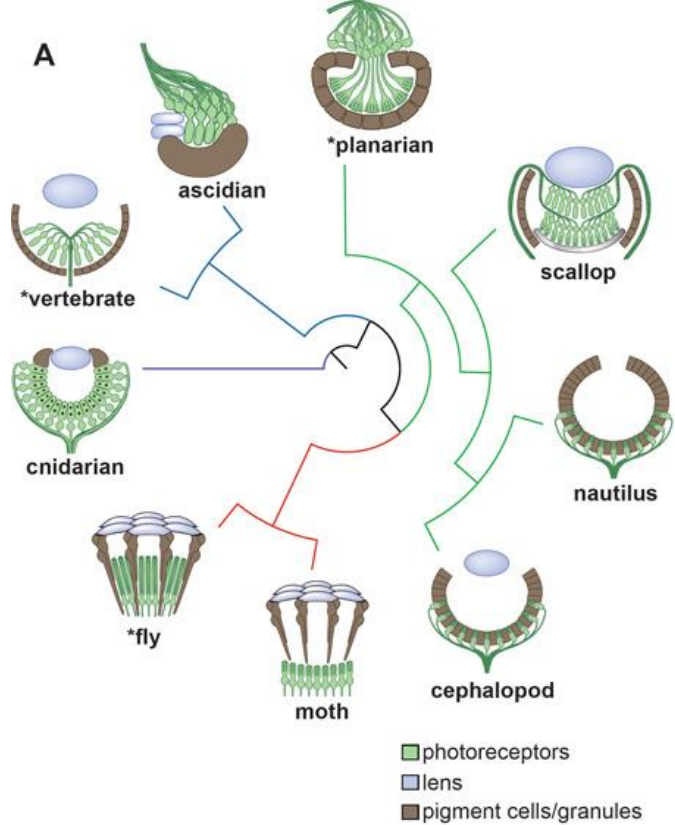
Planária





Inoue et al., 2004





Origem independente

MORFOLÓGICO

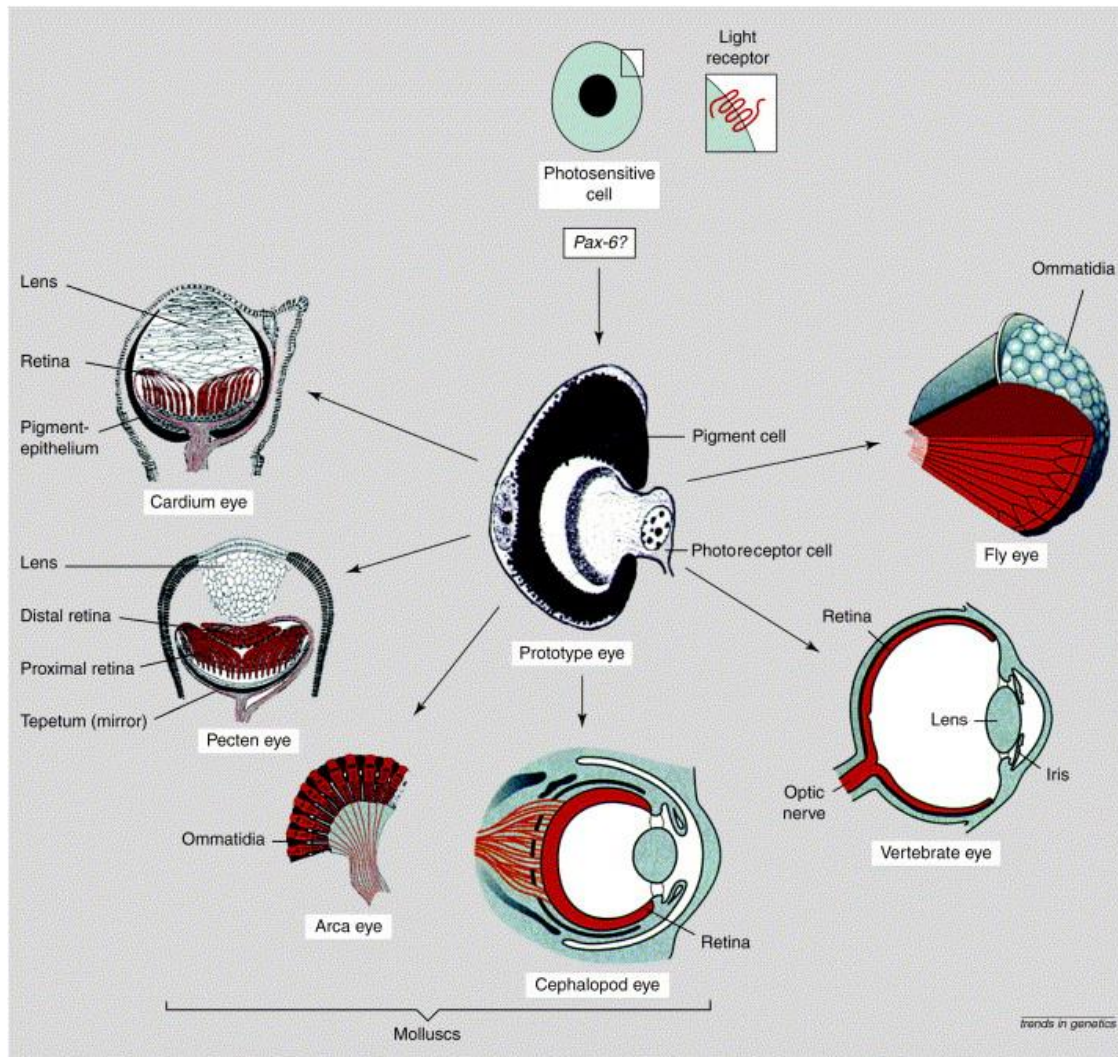
Origem comum - ressalvas

FISIOLOGICO

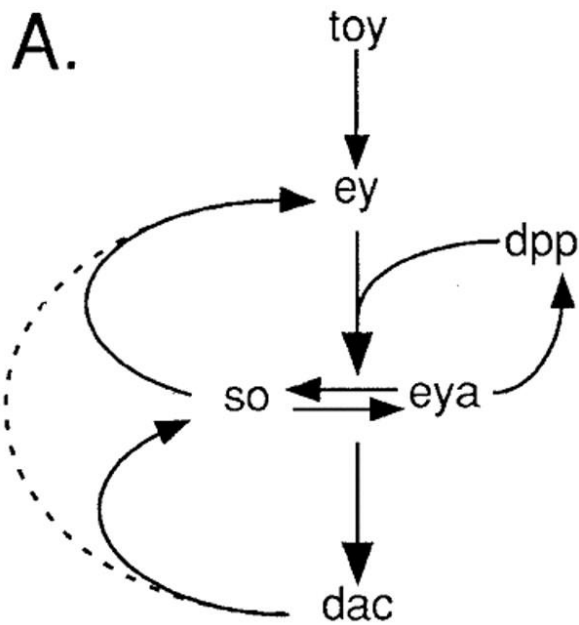
Estruturas diferentes

EMBRIONÁRIO

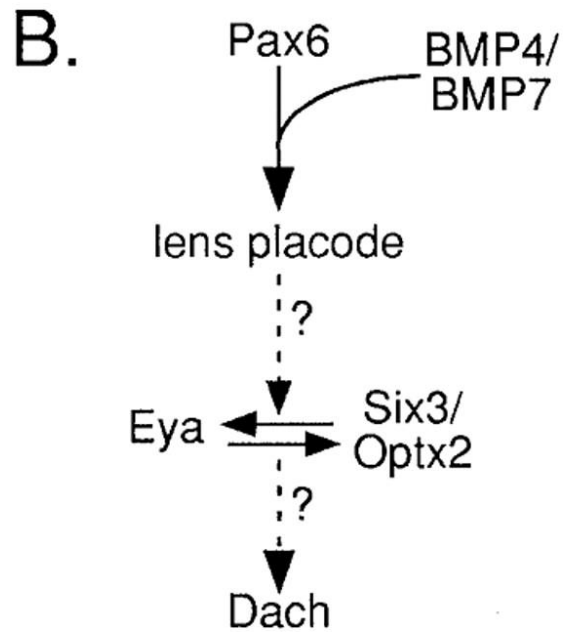
O que possuem em comum?



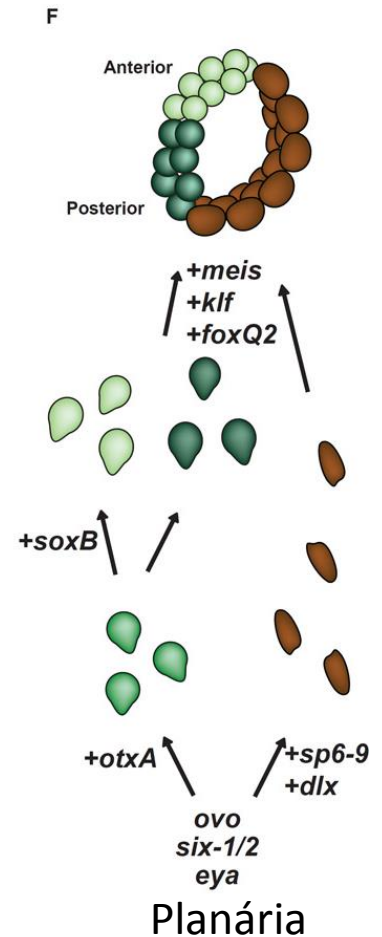
Não apenas Pax6



Drosophila



Mouse/Human



Planária

Origem independente

MORFOLÓGICO

Origem comum - ressalvas

FISIOLOGICO

Estruturas diferentes

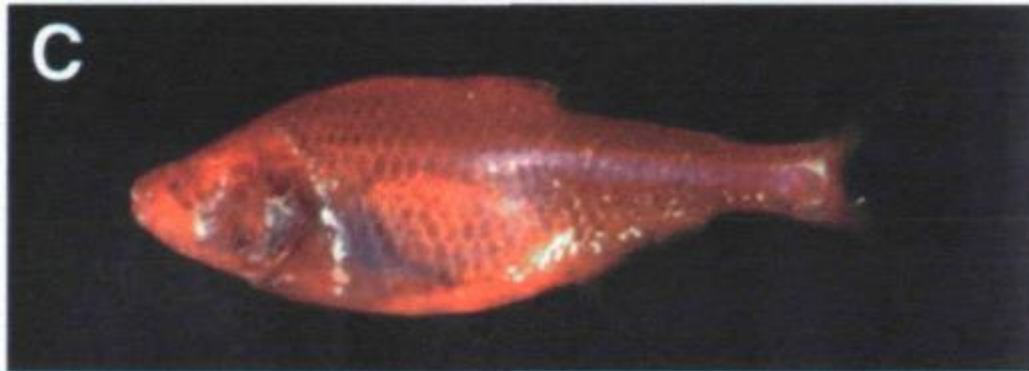
Mesmo fatores de transcrição envolvidos

EMBRIONÁRIO



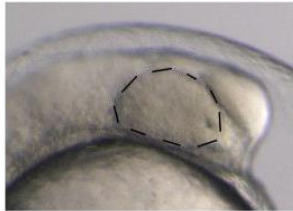
Jeffery, 2005

PERDA DA VISÃO ?

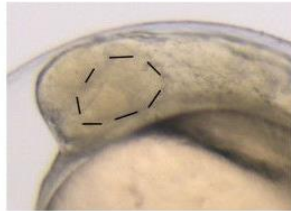


Astyanax mexicanus

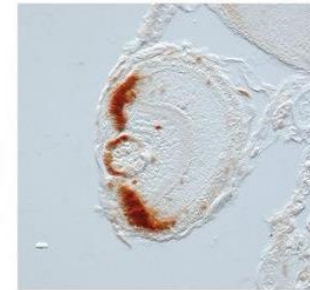
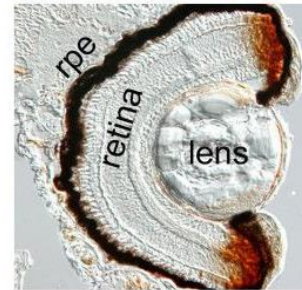
surface



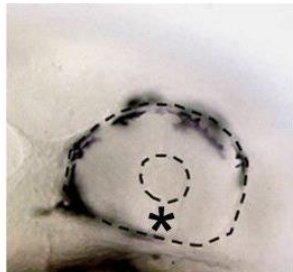
cave



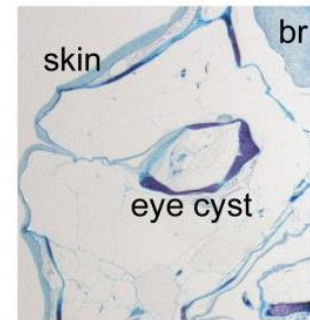
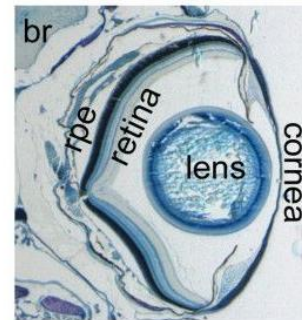
18hpf,
embryonic
eye vesicle



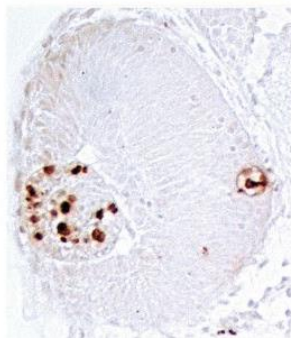
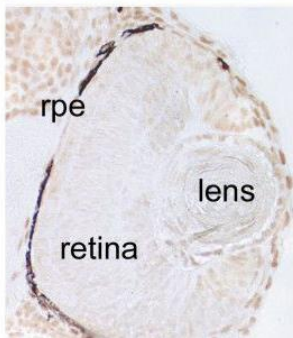
5dpf,
progenitor
proliferation
(PCNA)



24hpf,
larval eye
morphology



adult,
eye
morphology

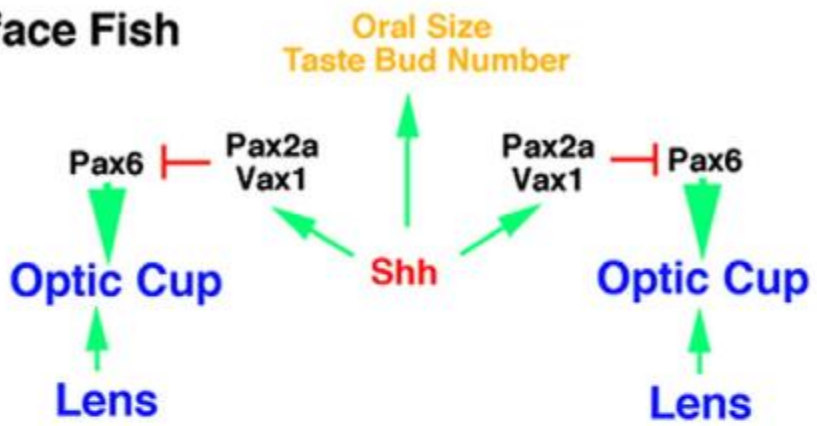


48hpf,
lens and retina
apoptosis
(TUNEL)



adult,
head
morphology

A Surface Fish



B Cavefish

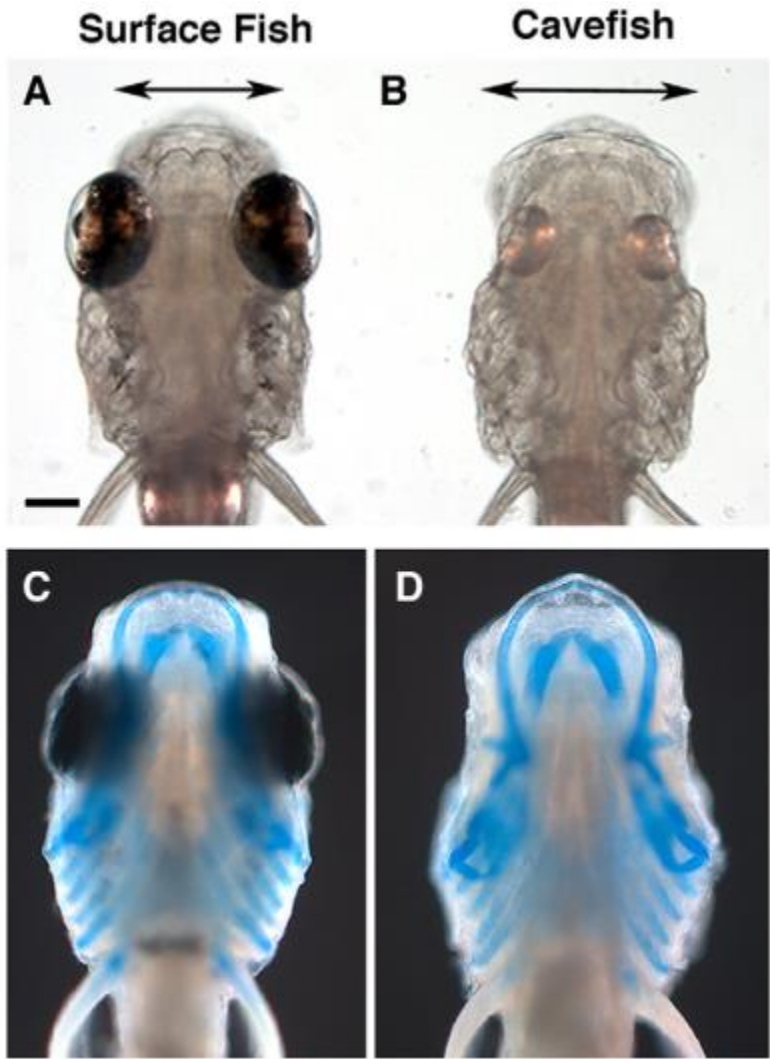
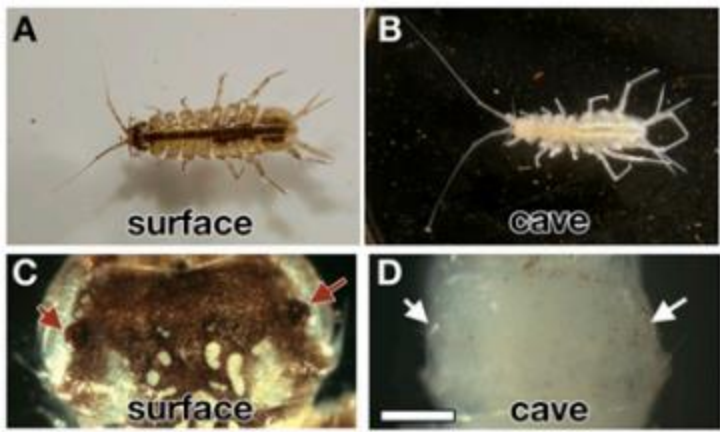


Fig. 7. The relationship between Shh signaling, oral-pharyngeal constructive traits, and eye degeneration in *Astyanax* surface fish (A) and cavefish (B) indicating the effects of Shh signaling on oral-pharyngeal, lens, and optic cup development. Letter size indicates relative increase or decrease in cavefish compared to surface fish. See text for other details.



Asellus aquaticus

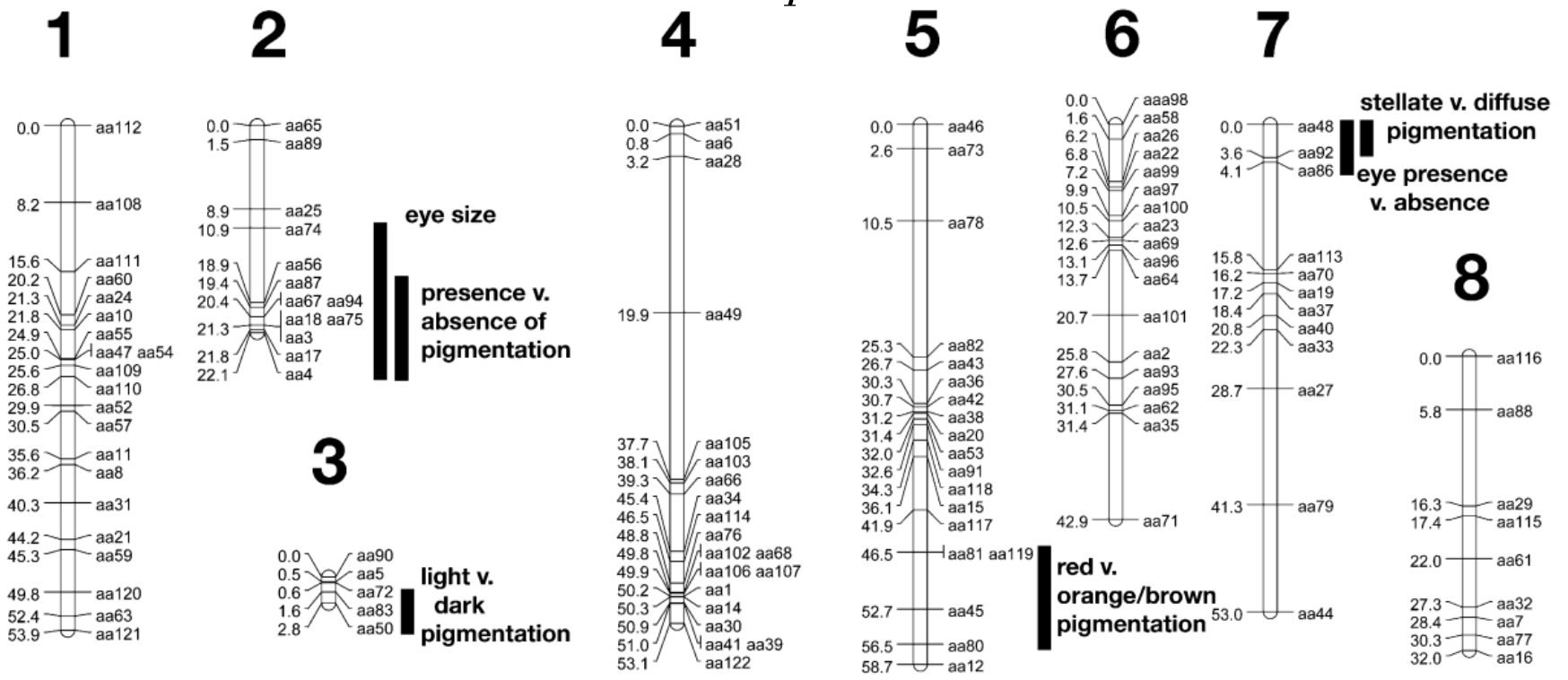
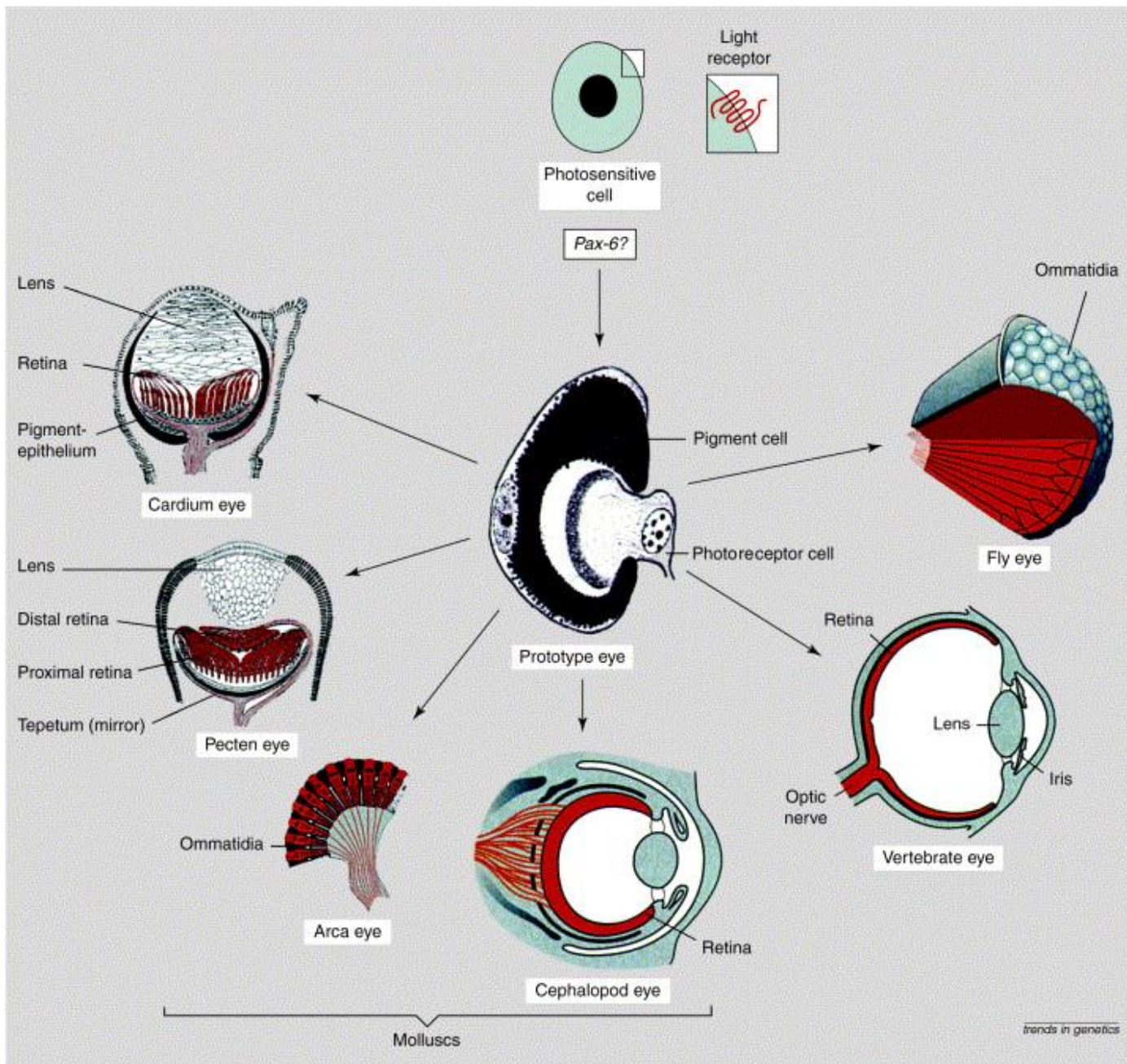


Fig. 2. Linkage map of *A. aquaticus*. Linkage group (LG) number is listed above each linkage group diagram. Placement in centimorgans is to the left and marker name to the right. The 1.5 LOD support intervals for each trait are shown with black vertical bars.

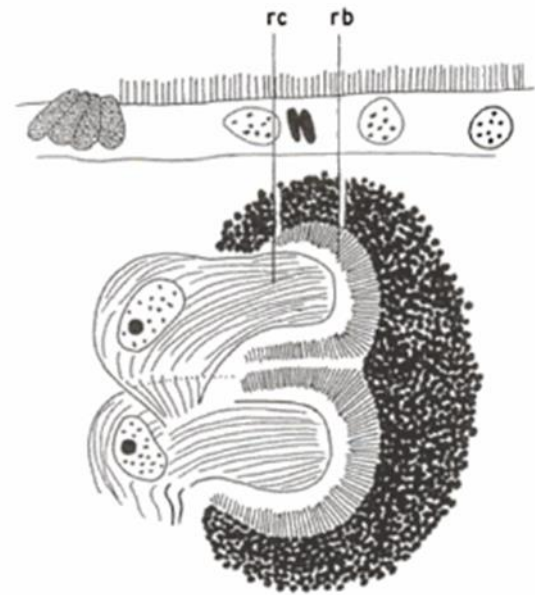
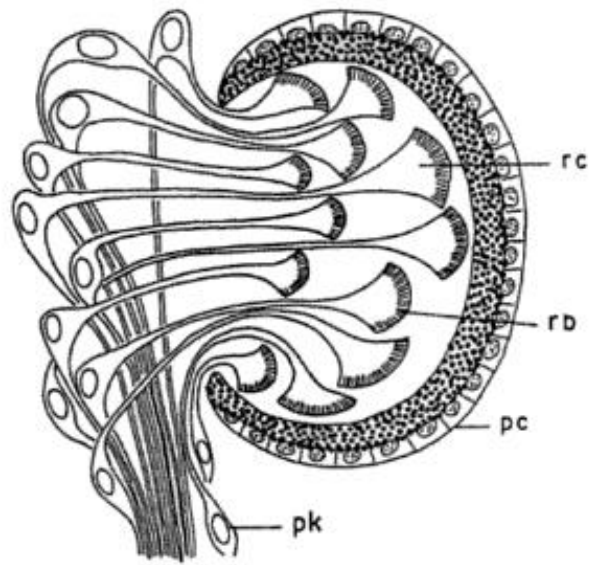
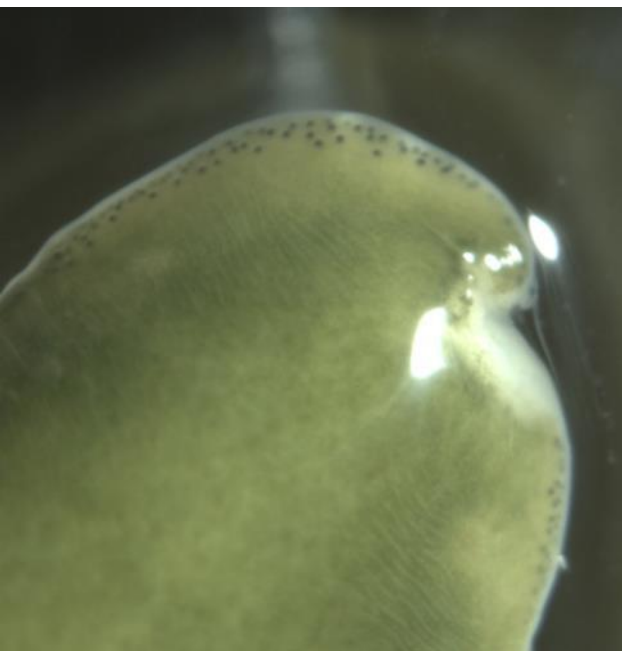
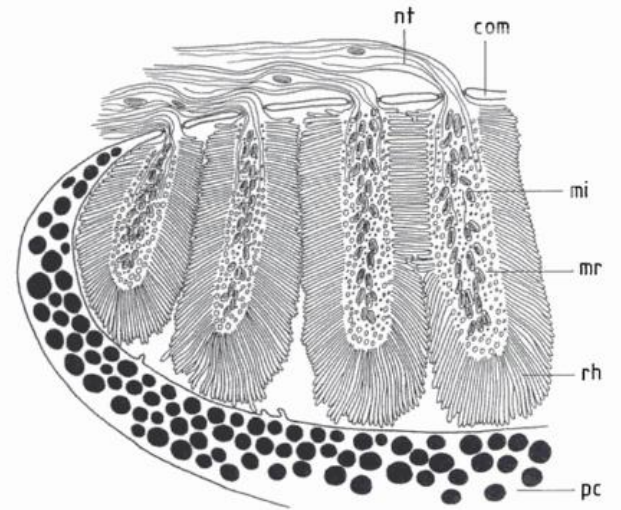
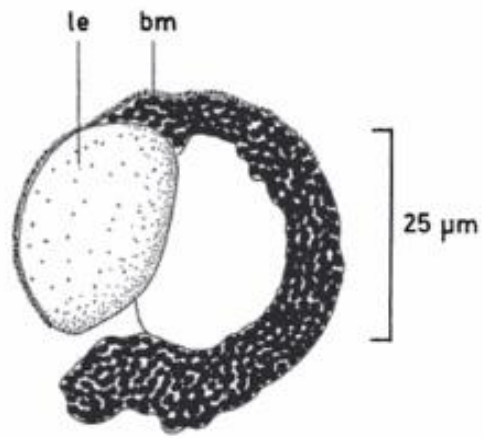
Locus de característica quantitativa QTL

FROM LIGHT TO DARKNESS:

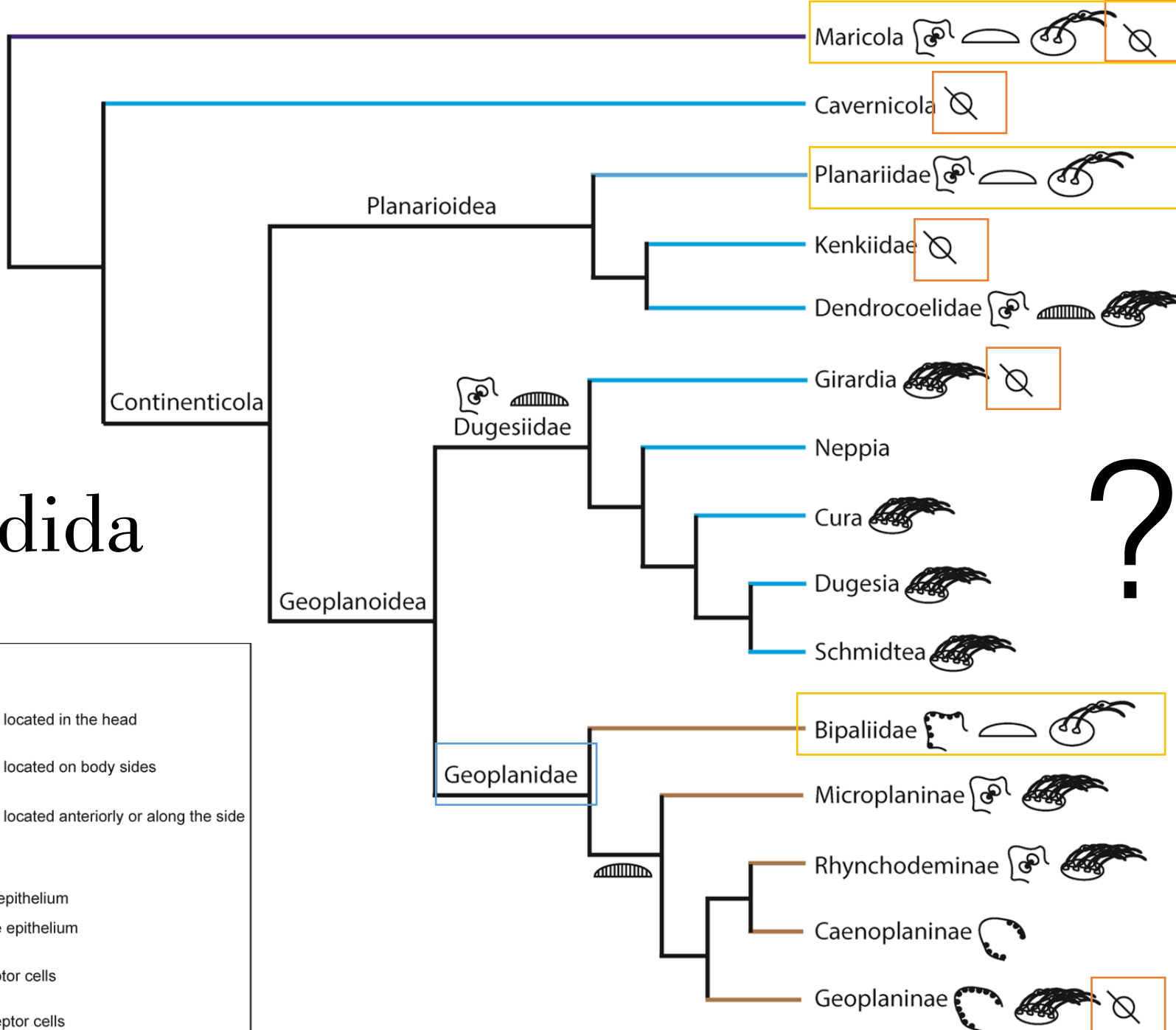
*evolutionary history of the photosensory
system in planarians*







Tricladida



	Two eyes
	Numerous eyes located in the head
	Numerous eyes located on body sides
	Numerous eyes located anteriorly or along the side
	Eye absence
	Unicellular eye epithelium
	Multicellular eye epithelium
	Few photoreceptor cells
	Many photoreceptor cells



Perda de olhos



Girardia tigrina



Girardia multiverticulata

Perda de olhos



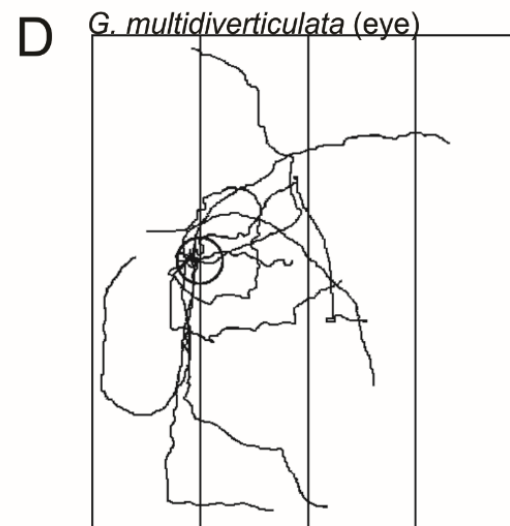
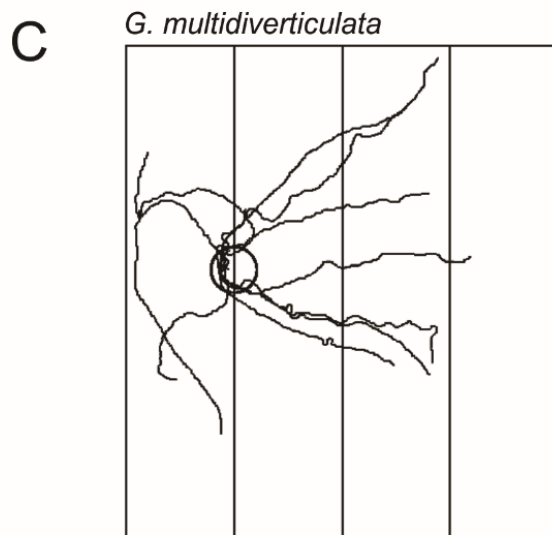
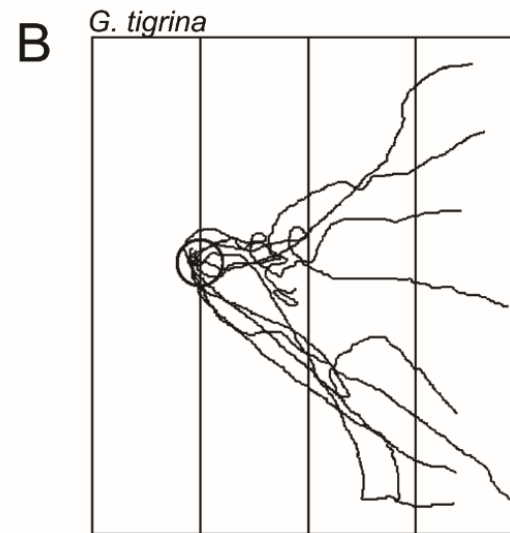
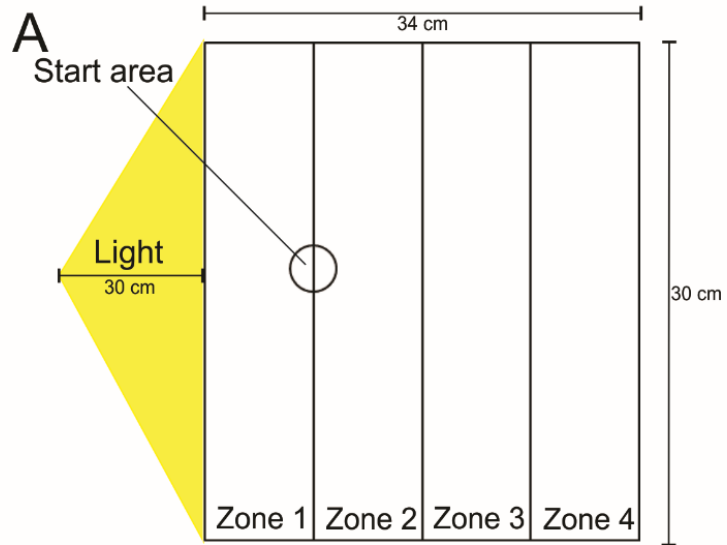
Girardia tigrina

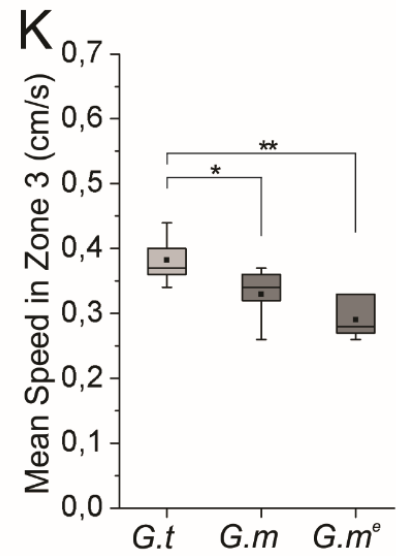
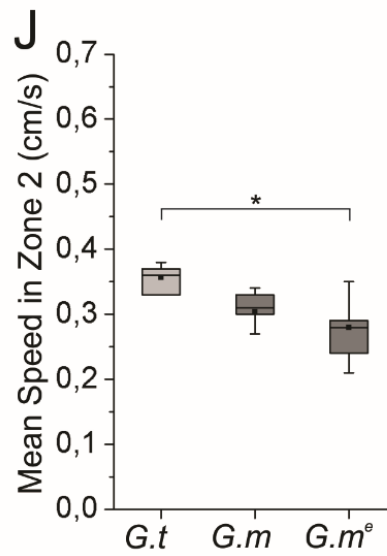
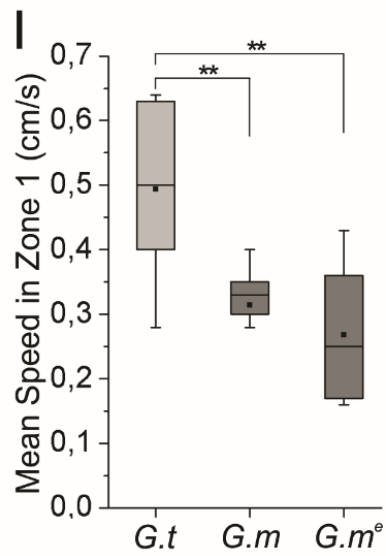
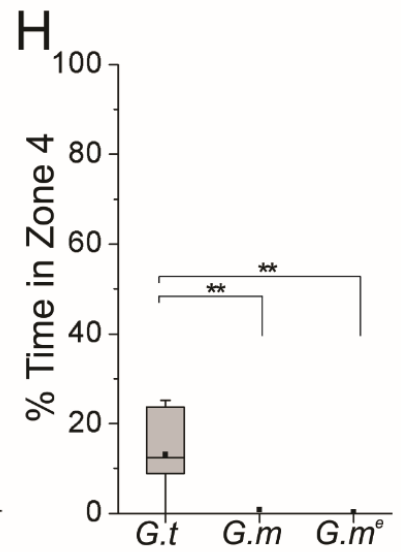
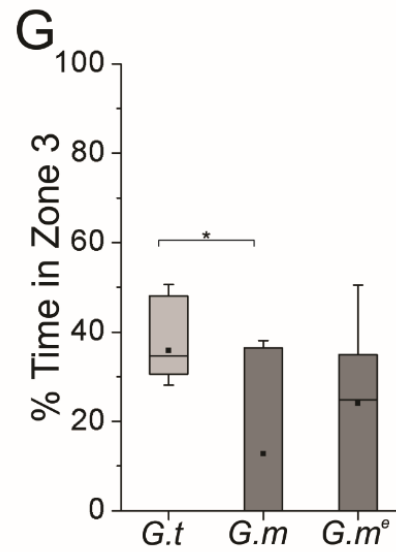
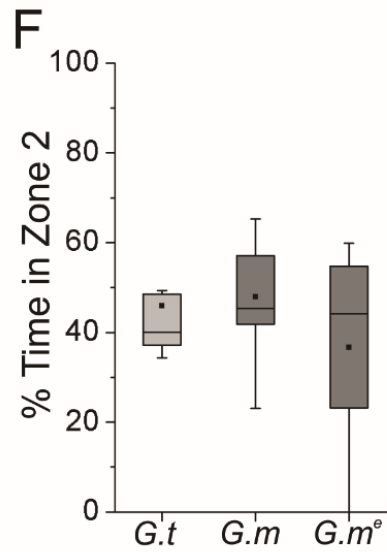
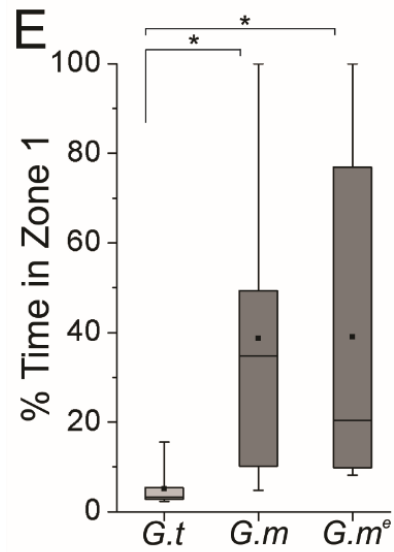


Girardia multiverticulata

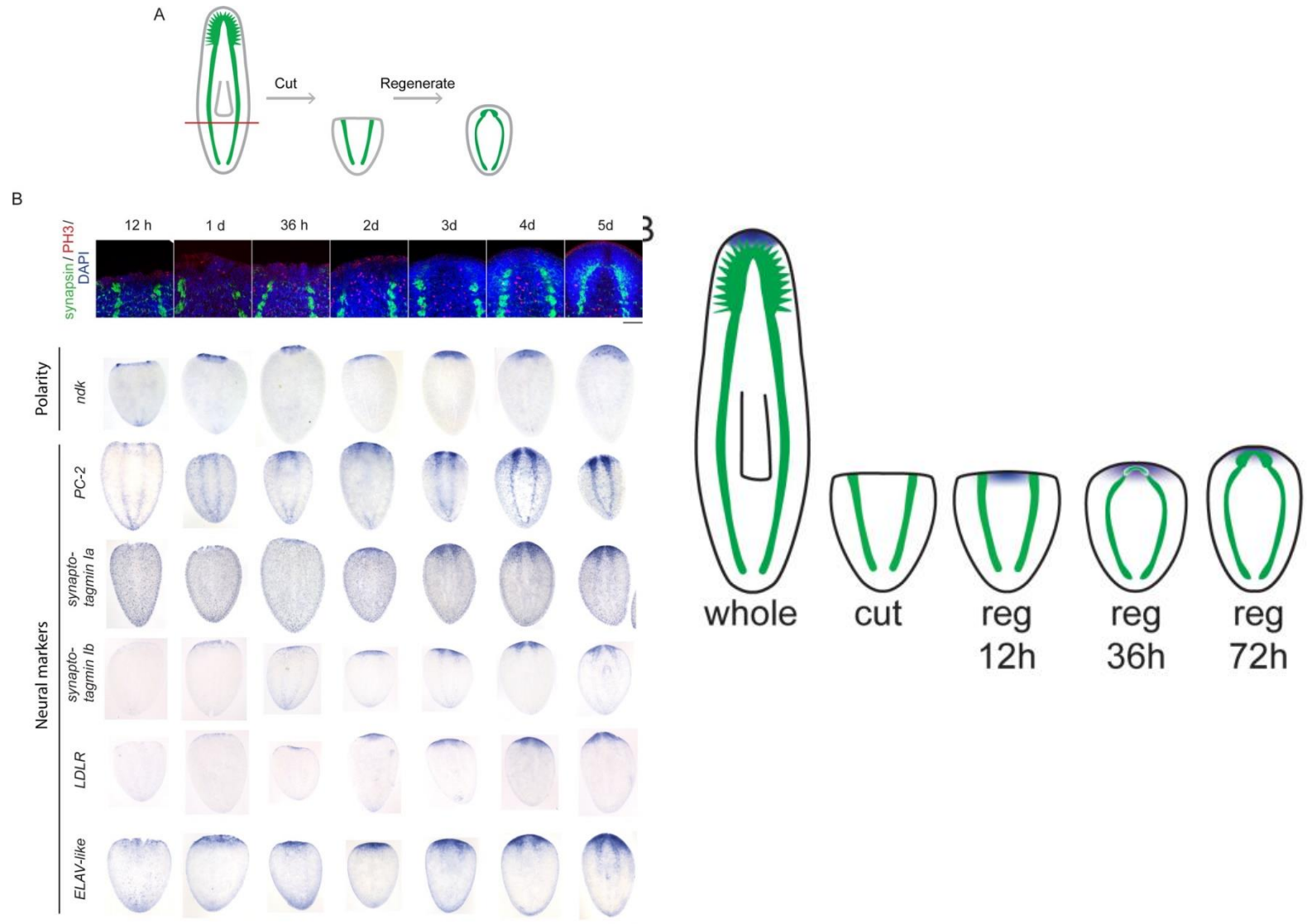


Girardia multiverticulata





Quais mecanismos moleculares envolvidos?

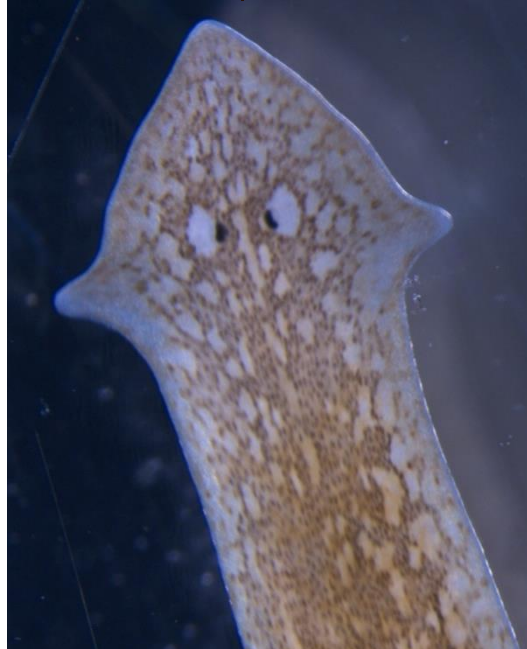
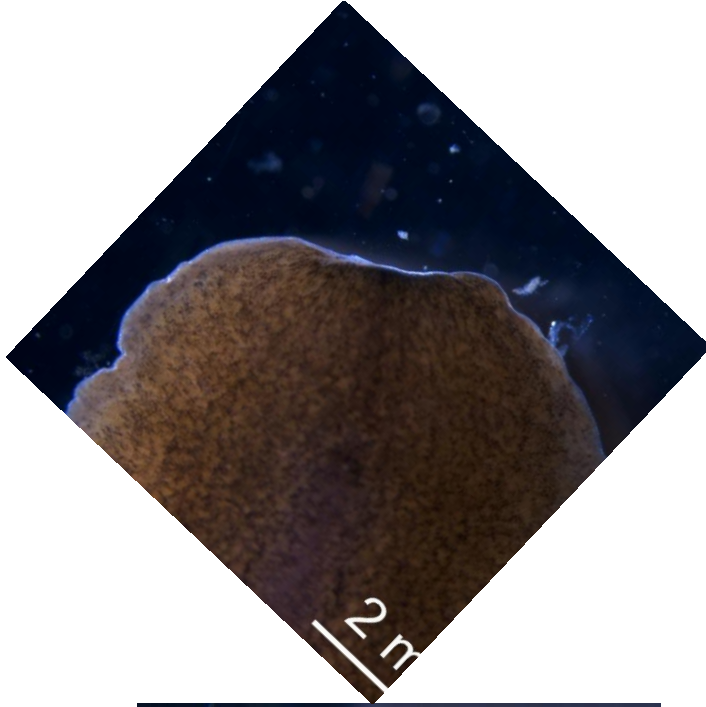


Prática

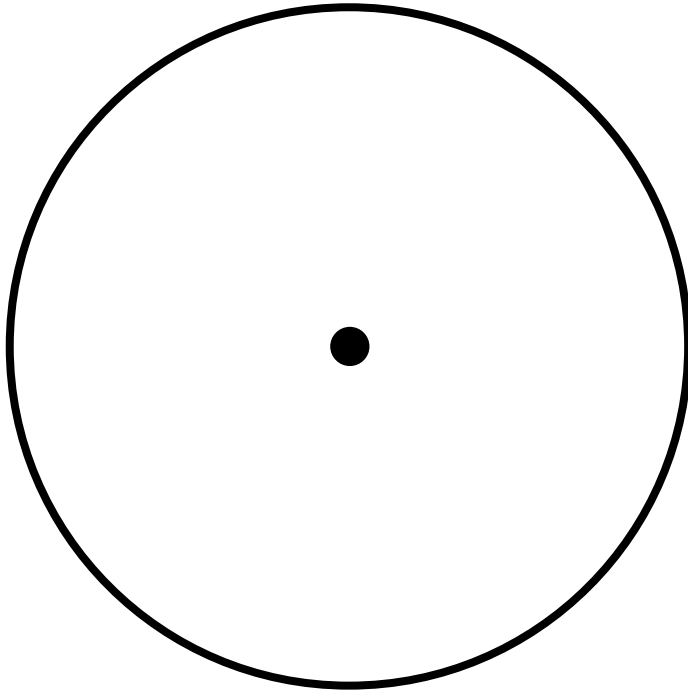
OBJETIVO

1) Compreender as etapas do desenvolvimento dos olhos das planárias

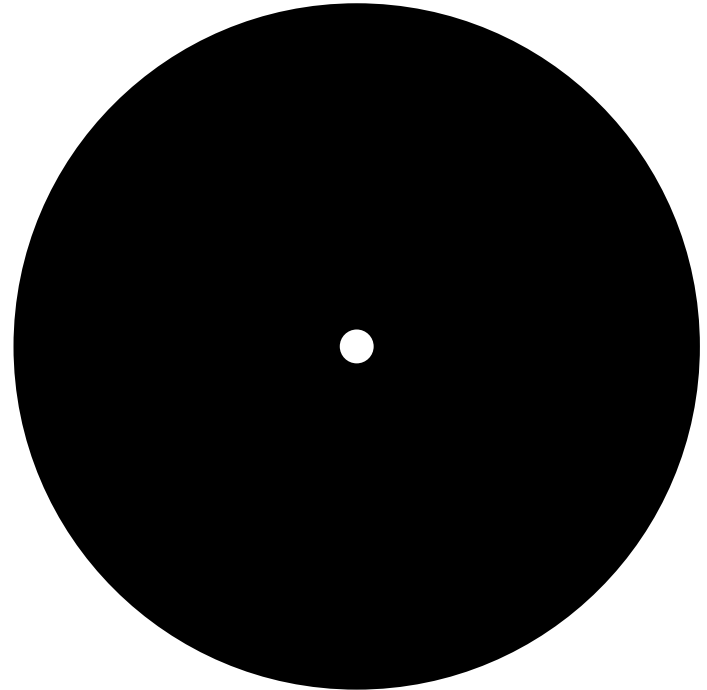
2) Realizar experimento simples de recuperação mecânica



Tratamento no claro

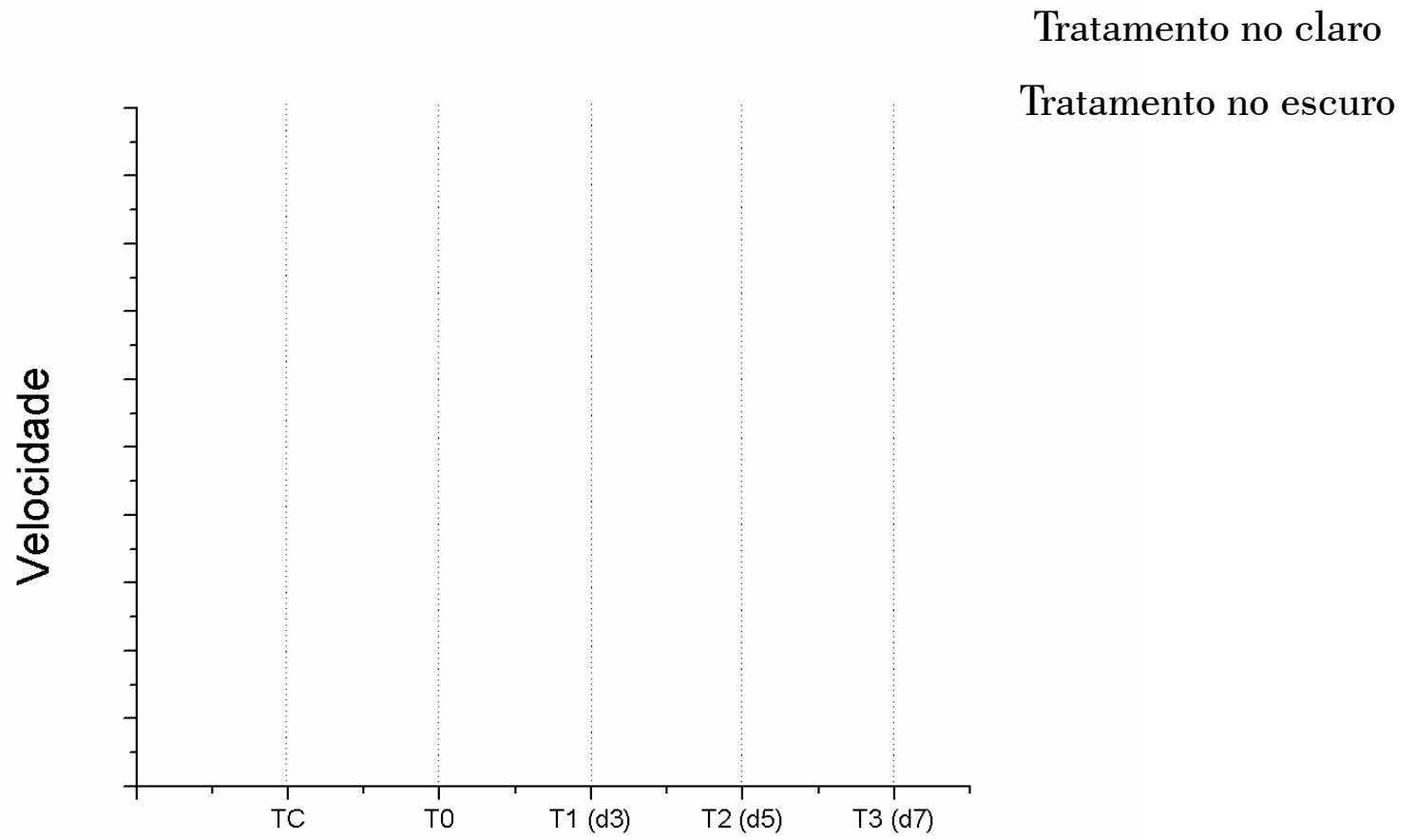


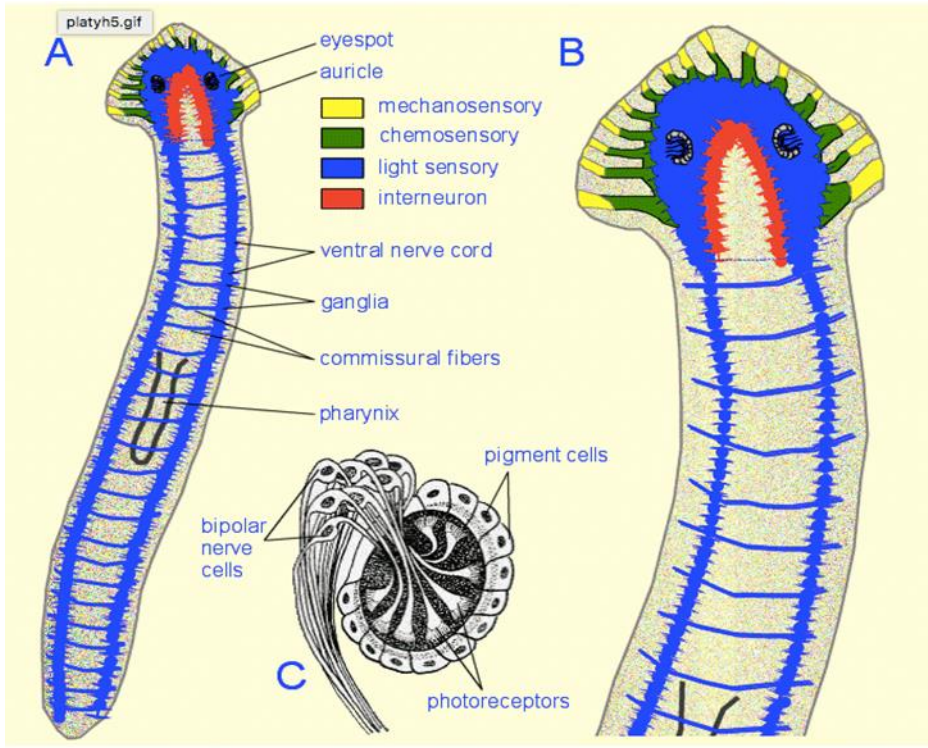
Tratamento no escuro



Por 1 minuto ou até permanecer na borda

$$Vm = \frac{Espaço(S)}{Tempo(t)}$$





Controle = 0 dias

Corte t1 = 3 dias

Corte t2 = 5 dias

Corte t3 = 7 dias