



Thyroid hormones, T₃ and T₄, in the brain

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Thyroid hormones (THs) are essential for fetal and post-natal nervous system development and also play an important role in the maintenance of adult brain function. Of the two major THs, T₄ (3,5,3',5'-tetraiodo-L-thyronine) is classically viewed as a pro-hormone that must be converted to T₃ (3,5,3'-tri-iodo-L-thyronine) via tissue-level deiodinases for biological activity. THs primarily mediate their effects by binding to thyroid hormone receptor (TR) isoforms, predominantly TR α 1 and TR β 1, which are expressed in different tissues and exhibit distinctive roles in endocrinology. Notably, the ability to respond to T₄ and to T₃ differs for the two TR isoforms, with TR α 1 generally more responsive to T₄ than TR β 1. TR α 1 is also the most abundantly expressed TR isoform in the brain, encompassing 70–80% of all TR expression in this tissue. Conversion of T₄ into T₃ via deiodinase 2 in astrocytes has been classically viewed as critical for generating local T₃ for neurons. However, deiodinase-deficient mice do not exhibit obvious defects in brain development or function. Considering that TR α 1 is well-established as the predominant isoform in brain, and that TR α 1 responds to both T₃ and T₄, we suggest T₄ may play a more active role in brain physiology than has been previously accepted.

Keywords: T₄ thyronine, T₃ thyronine, thyroid hormone receptor, brain, coregulator, deiodinase 2

INTRODUCTION

Thyroid hormones (THs) are synthesized by the thyroid gland and are critical regulatory molecules with important roles in vertebrate physiology and development, including fetal and post-natal nervous system development and the maintenance of adult brain function (1, 2). The TH requirement for development is most apparent in the central nervous system (CNS) where severe TH deficiency in fetal and neonatal periods results in cretinism, a disease characterized by mental retardation, deafness, and ataxia; these consequences are irreversible if not treated soon after birth (3–5). Additionally, untreated hypothyroidism in the adult is associated with severe intellectual defects, abnormal balance and defects in fine motor skills, spasticity, and deafness (6). Correcting TH deficiencies is critical for normal brain development and function.

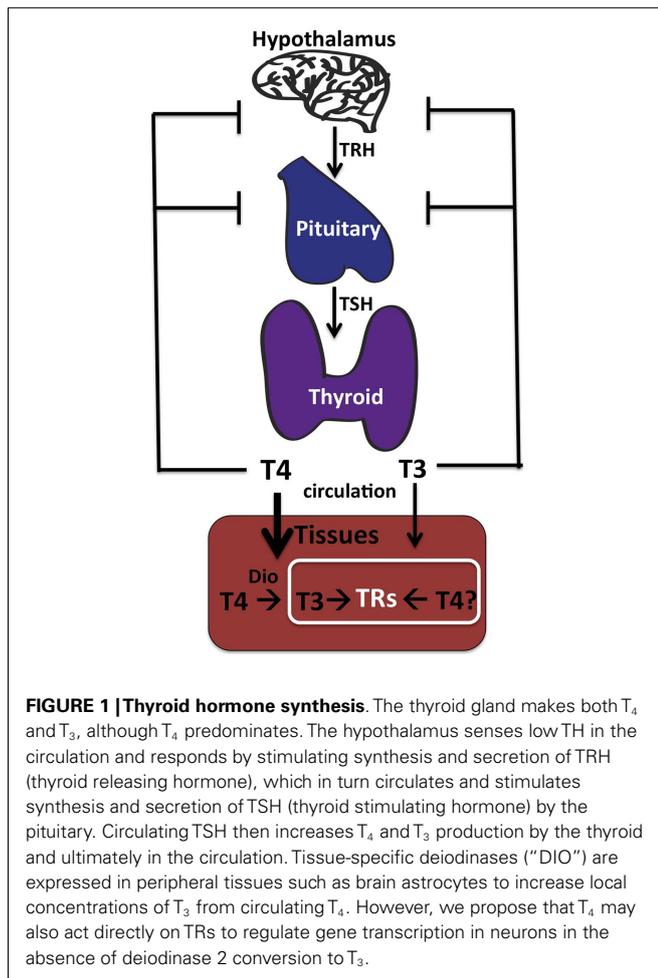
Thyroid hormones mediate CNS effects primarily through thyroid hormone receptors (TRs), members of the nuclear hormone receptor family (4, 7, 8). TRs bind to the DNA regulatory regions of target genes to activate or repress transcription through interactions with accessory proteins known as coregulators. There are two major THs, which bind to and activate TRs: T₃ (3,5,3'-triiodo-L-thyronine) and T₄ (3,5,3',5'-tetraiodo-L-thyronine, also known as thyroxine). T₄ differs from T₃ by an additional iodine located at the 5'-position of the first thyroxine ring. T₃ has been assumed to be the active form of TH, as T₃ binds to TRs with a greater affinity than T₄. In this model, T₄ is thought to simply act as a pro-hormone, existing only to be circulated in the serum and converted at the tissue-level to T₃ through an enzymatic reaction involving the removal of the 5'-iodine atom from T₄ by local deiodinases (9, 10). Nonetheless, it is notable that most of the TH produced under normal conditions in the thyroid is secreted in the form of T₄ and steady-state serum concentrations of T₄ are

many fold greater than those of T₃ (11–14). Notably, iodine intake is important for the maintenance of both of these TH levels in circulation. In fact, during gestation and lactation in females, double the normal iodine intake is required to maintain adequate T₃ and T₄ in circulation to ensure normal fetal development (15, 16). Under conditions of low iodine intake, the serum T₃/T₄ ratio is somewhat increased reflecting the reduced abundance of iodine atoms (16). Although the ready availability of dietary iodized salt has largely eliminated these iodine deficiencies for school children in most developed countries today, these advances are often not adequate for pregnant and lactating women (17).

Indeed the primary TH crossing the adult blood–brain barrier (BBB) is believed to be T₄; therefore, the adult brain may have access to sufficiently high levels of T₄ to allow for direct binding to and transcriptional activation of TRs (18, 19). In fact, we know that both T₄ and T₃ binding by TRs lead to very similar structural changes in the receptor (12). Several reports have also shown that T₄ exhibits non-genomic effects by interacting with integrin cell membrane receptors (20). These studies suggest that T₄ may exhibit a greater role in physiology than merely acting as a pro-hormone. Therefore, the precise role of T₄ as a pro-hormone and whether T₄ might function directly as an active hormone in the CNS, remain incompletely answered questions.

T₄ SYNTHESIS, TRANSPORT, AND AVAILABILITY IN THE BRAIN

Determining the effective cellular concentrations of T₄ and T₃ in the brain, or in any tissue, is difficult due to the complexities of TH synthesis, transport, and regulation. Vertebrates have developed multiple mechanisms to ensure delivery of appropriate levels of TH to peripheral tissues such as the brain. These include regulation of secretion of THs from the thyroid into serum (21, 22),



control of free versus bound levels of THs determined by reversible binding to serum-binding proteins (22), cell-specific expression of TH cell membrane transporters (23, 24), and finally intracellular deiodination of T₄ to form T₃ [(22, 25); **Figure 1**].

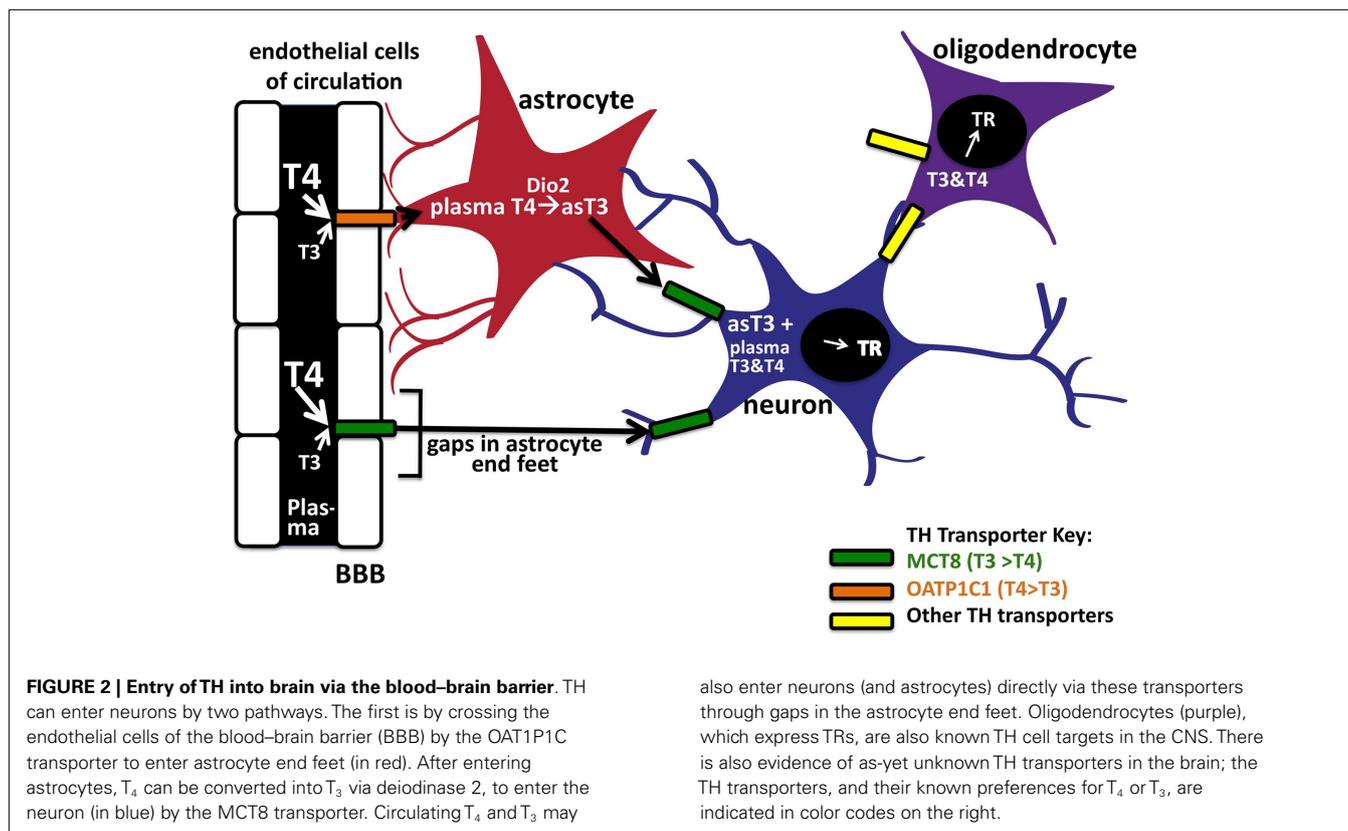
Transplacental TH transfer from maternal to fetal circulation is particularly important in vertebrate CNS development [reviewed by Ref. (26)] to ensure appropriate levels of TH are available to the fetus throughout development (16). Throughout the first trimester when TH levels are solely obtained through maternal transfer, free T₄ levels are high in the fetus, similar to levels of biologically active T₄ in adults, whereas fetal concentrations of T₃ are at least 10× lower than T₄ (16). Notably, T₃ levels in the fetal cerebral cortex increase somewhat between 12 and 20 weeks PMA (post-menstrual age) when placental deiodinase 2 levels increase (see below), although maternal serum levels of T₃ are still low. Both T₄ and T₃ in the fetus continue to be transferred from maternal origins through the placenta until half-way through pregnancy when endogenous THs are produced by the fetal thyroid. However, because fetal T₄ synthesis is elevated over that of T₃ for several weeks at this time, it is possible that an additional window in development exists where fetal circulating T₄ is quite high and may act as an active hormone with TRs (16).

Outer ring 5′-monodeiodination via cell-specific deiodinases converts a small fraction of the normal serum T₄ pool to T₃ (10, 22). Deiodinase 2 is the primary enzyme responsible for intracellular conversion of T₄ into T₃ in most local tissues including brain, whereas deiodinase 1 is found primarily in the liver (25, 27). Deiodinase 2 is only expressed in selected cell types within the CNS: astrocytes and tanycytes. These are both glial cell-derived and are located in the hypothalamus (28–30). The other deiodinase enzyme expressed in the CNS is deiodinase 3, selectively expressed in neurons. Deiodinase 3 inactivates both T₄ and T₃ by inner ring deiodination to rT₃ and T₂ so as to down-regulate local TH concentrations and protect the neuron from supraphysiological levels of TH. Currently it is believed that astrocytes generate active T₃ from circulating pro-hormone, T₄, whereas neurons degrade both T₄ and T₃ to inactive rT₃ and T₂, respectively, and thereby regulate local TH availability within the brain. When levels of TH are low, deiodinase 2 levels in brain increase and contrastingly when there are high levels of TH, deiodinase 3 levels increase (19, 30, 31). This balancing act protects the brain from the detrimental effects of hyper- or hypothyroidism.

T₃ concentrations equilibrate rapidly in peripheral tissues such as the liver and kidney but appear to take longer to equilibrate in the brain. In general, TH concentrations in the CNS are approximately 20% that of serum concentrations (32); this is likely due to the added complexity of TH transport across the BBB, which is comprised of the endothelial cells of brain capillaries surrounded by astrocyte end feet. To enter the brain, the THs cross the BBB of the choroid plexus via the MCT8 or OATP1C1 TH transporters. T₄ is thought to predominately enter the CNS in preference to T₃ as the majority of BBB TH transporters exhibit greater affinities for T₄ transport [(19, 33); **Figure 2**]. As mentioned above, after T₄ is taken up into astrocytes likely by OATP1C1, deiodinase 2 can in turn convert it locally to T₃. Finally, the astrocyte-generated T₃ can enter neuronal cells via the MCT8 transporter to bind and activate TRs. Therefore, it is intriguing that the T₄-activating deiodinase is not expressed in the neurons themselves, where the relevant TRs are located, but in the astrocytes. T₄ and/or T₃ also enter the CNS directly via gaps in the end feet of the astrocytes, which do not completely cover the capillaries in contact with the interstitial spinal fluid (34).

DIFFERENT TR ISOFORMS DIFFER IN THEIR ABILITY TO BIND TO T₄

Thyroid hormones bind TRs, ligand-regulated transcription factors, which bind to specific target DNA sequences and repress or activate target genes through the recruitment and release of accessory proteins. TRs contact their DNA-binding elements as protein dimers, heterodimerizing with another member of the nuclear receptor family, RXRs (primarily Retinoid X Receptors), or homodimerizing with themselves (35–39). TRs exhibit bimodal regulation, typically binding corepressors to repress transcription of target genes in the absence of TH, but releasing corepressors and recruiting coactivators to activate transcription of these “positive response” target genes in the presence of TH (40, 41). These corepressor and coactivator proteins alter the chromatin template or interact with the general transcription machinery to produce the appropriate transcriptional outputs. However, many TR target



genes display the opposite properties in that they are expressed in the absence of TH and are repressed in the presence of TH; the molecular mechanisms involved in this “negative response” is not well-understood.

Thyroid hormone receptors are encoded by two distinct genetic loci, denoted THRA and THRB, which are each expressed as alternatively spliced mRNAs to create additional receptor diversity [reviewed in Ref. (42)]. Two of the major TR isoforms are referred to as TR α 1 and TR β 1; both bind TH and yet exhibit distinct biological roles [reviewed in Ref. (43)]. TR α 1 is expressed early in embryonic development and then widely in adults whereas TR β 1 is expressed later in embryonic development and exhibits a more restricted tissue-expression pattern in adults (31, 44–49). Genetic disruption in mice of TR α 1 or TR β 1 indicates that these isoforms have somewhat overlapping, yet distinct roles in normal physiology (45–47, 49, 50).

These two different TR isoforms differ in their ability to respond to T₄, with TR α 1 generally exhibiting a much stronger response to T₄ than TR β 1. We suggest that different cell types may modulate their relative ability to respond to T₄ versus T₃ by altering the relative abundance of different coactivators and corepressors that have distinct responses to T₄ and T₃, raising the possibility that T₄ may be able to function as a direct-acting hormone agonist with TR α 1 (Amy C. Schroeder and Martin L. Privalsky, unpublished observations).

TR α 1 EXPRESSION IN THE BRAIN

Notably, TR α 1 encompasses 70–80% of all TR expression in the adult vertebrate brain (2) and TR α 1 is present in nearly all

neurons (51). Intriguingly, TR α 1 is also the predominating TR isoform early in fetal brain development (detected by 8.1 weeks and increasing until 13.9 weeks post-menstrual age). Critical roles in CNS development are known to be mediated by TR α 1 including TH-dependent oligodendrocyte differentiation (52). If TR α 1 is inactivated, the number of mature oligodendrocytes after T₃ treatment is decreased (52). The commitment of these cells as oligodendrocytes is therefore believed to be linked to cell-specific TR α 1 expression while the availability of TH regulates the timing of differentiation (52). In fact, maturation of several cell types in the brain in development may depend on specific windows of TR α 1 expression and involve a complicated interplay between TRs, THs, and coregulators (2). Additionally, TR α 1 is known to exhibit important roles in later stages of neurodevelopment and its expression persists in adult neurons. Therefore, it is interesting that expression of the TR α 1 isoform predominates in both fetal and in adult brain at the same times when free T₄ levels appear to be at biologically active levels (16), suggesting windows in brain development may exist where T₄ may act on TR α 1.

DEIODINASE 2-DEFICIENT MICE EXHIBIT NORMAL CNS DEVELOPMENT AND FUNCTION

As noted above, deiodinase 2 expression does not overlap TR receptor expression in the brain. Deiodinase 2 is expressed instead in astrocytes whereas the TRs are expressed in neurons along with deiodinase 3 [(28, 29); Figure 2]. The current theory therefore suggests astrocytes are involved with T₄ uptake from capillaries to subsequently generate a source of locally generated T₃. Conversion of T₄ into T₃ via deiodinase 2 in astrocytes has been estimated to

produce as much as 80% of the T₃ bound to the TRs in the brain (18), suggesting astrocyte deiodinase 2 is important for generating local T₃ concentrations. Therefore, many argue that deiodinase 2 likely plays a critical role in developing brain by providing the necessary amount of T₃. If this were in fact the case, one would predict the absence of deiodinase 2 would result in detrimental defects in CNS development similar to that seen in hypothyroidism.

However, the Galton lab produced a deiodinase 2-deficient and a deiodinase 2/deiodinase 1 dual-deficient mouse (KOs) without any evident defects in brain development or function (27, 53). The deiodinase KO mice demonstrated slightly elevated circulating T₄ and TSH levels, and normal thyroid-secretion of T₃ but no tissue-level production of T₃ from T₄ (27). Notably, these mice did not display any signs of hypothyroidism and have no gross physiological or behavioral abnormalities (27). The deiodinase KO was also combined with an MCT8 TH transporter knockout (54, 55); this combination resulted in minor neuronal defects mostly noted by decreased expression of genes in the neural cortex, which are usually positively regulated by T₃, however, most neural development and function was normal. KO mice studies suggest that T₃ transport into the brain and local conversion of T₄ to T₃ in the brain are not essential for normal brain function in mice, and suggest that CNS T₃-defects do not produce syndromes as severe as that seen in the hypothyroid mice (27).

Many suggest that there might be compensation in the deiodinase KO mice through the absorption of more T₃ directly from circulation via the MCT8 transporter in endothelial cells of the BBB, but it should be again noted that the parallel transporters such as OATP1C1 and OATP2 favor T₄ transport (56, 57) and it is unlikely that T₃ can be transported into the brain at rate equivalent to T₄ transport. We suggest that in the absence of available T₃, T₄ can act as an active TH in the brain working on, most likely, TR α 1. Interestingly, in the absence of deiodinase 1 and 2, positively regulated TH genes in the cerebral cortex remain unaffected but negatively regulated TH genes appear to be impaired in a way that parallel the hypothyroid mice (27, 58). Perhaps in the absence of deiodinase 2, T₄ can act as an active hormone in brain cells to activate positively regulated TH genes, but not to repress negatively regulated TH genes.

It should be noted that humans with MCT8 mutations display severe neurodevelopmental defects with psychomotor retardation and abnormal serum TH levels (57, 59). Contrastingly, MCT8 KO mice mimic the human MCT8 mutations in their thyroid phenotype but display no obvious brain developmental defects (57, 59). It is therefore possible that the need for locally produced T₃, and/or the presence of alternative T₃-specific transporters, differ in mice and in humans (55).

TR COREGULATORS AND THE BRAIN

T₄ efficiently recruits many coactivators to TR α 1, with certain well-established TR coactivators (SRC1 and TRAP220) exhibiting a T₄ response equal or near equal to that induced by T₃ (Amy C. Schroeder and Martin L. Privalsky, unpublished data). SRC1 mRNA is expressed in many tissues during development including the CNS (60). TRAP220 is also expressed in the developing brain and is thought to play a regulatory role in the process of cell proliferation and differentiation, in learning, and in memory formation

(61). The widespread expression of TRAP220 in the developing brain appears to parallel TR α 1 expression. Therefore, CNS development correlates with a high level of expression of TR α 1 together with TRAP220 and/or SRC1 and may provide an opportunity for T₄ to directly regulate gene transcription. CNS cell-specific differences in TR isoform and cofactor levels or function are likely to contribute to differences in T₄ hormone response and may suggest a means by which the T₄ sensitivity of a given CNS cell type can be regulated in response to internal or external signals.

A POSSIBLE DIRECT ROLE FOR T₄ IN BRAIN: ARE THERE CONTEXTS IN THE BRAIN IN WHICH T₄ IS A DIRECT-ACTING TR α 1 AGONIST?

Several recent studies have led to the view that T₄ exhibits non-genomic roles that do not require conversion to T₃ (20) but which have not challenged the general view that T₃, not T₄, is the only direct, biologically relevant agonist for nuclear TR function. Our own experiments indicate that TR α 1 has the potential to act as a dual sensor of both T₄ and T₃ (Amy C. Schroeder and Martin L. Privalsky, unpublished observations).

Although the effective concentration of T₄ in the brain is difficult to determine, it is plausible that T₄ levels are sufficient to induce activation of TR α 1-regulated genes in the brain even in the absence of T₃. We suggest that the normal mix of T₄ and T₃ in the brain may actually confer a mixed T₄/T₃ transcription response mediated primarily by TR α 1, together with a more pure T₃ response mediated primarily by TR β 1. Notably, mice in which both deiodinase 1 and 2 have been genetically ablated, and thus lack astrocyte deiodinase conversion of T₄ to T₃, display only very mild defects in their physiological with little to no neurological defects (27). If, as indicated by these knockouts, T₄ is not absolutely required in its traditional role as a pro-hormone, the dominance of T₄ to T₃ in the circulation and transport into the CNS may instead reflect a novel role of T₄ as a direct-acting hormone and this direct role may be helping to ameliorate the effects of the deiodinase knockouts in the CNS.

In conclusion, TH endocrinology in the CNS is tightly regulated at multiple tiers. Negative feedback loops in the hypothalamus and the pituitary control T₃ and T₄ output by the thyroid gland itself. Further, multiple phenomenon functions together to modulate the transport of circulating TH through the BBB, and multiple transporters act together to directly alter TH availability in the CNS itself. Additionally, conversion of intracellular T₄ into T₃ by deiodinase 2, inactivation of both T₃ and T₄ by deiodinase 3, and, the ability of different TR isoforms and different coregulators to respond directly to T₄ versus T₃ further regulate the CNS response to TH. Operating together, we propose these mechanisms serve to maintain proper endocrine homeostasis while permitting the CNS to respond to developmental and physiological needs.

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