

Macronutrient analysis

| Food component | Available method of analysis | Limitation | Application |
|------------------|---|--|--|
| Water (moisture) | Air oven* | Caramelization of sugars, degradation of unsaturated fat, loss of volatiles | This method is applicable to all foods at 60°C. At 100°C, it is applicable to all foods except those rich in fat and sugar |
| | Vacuum oven* | Loss of volatiles | |
| | Freeze-drying* | Slow. Care must be taken to avoid residual water in samples | Applicable to most foods |
| | Microwave oven | Charring | Applicable to medium- or high-moisture foods only |
| | Dean and Stark distillation | Safety of solvents used | Applicable to foods high in volatiles* |
| | Karl Fisher | | Applicable to low-moisture, hygroscopic foods |
| | Physical methods (NMR, NIR) | High cost and needs calibration for each food group | NMR is applicable to most foods. NIR is only established for cereals and some other foods |
| | Chromatography (GLC, GSC) | High cost | GLC is applicable to meat and meat products only. GSC is applicable only to some meat products |
| Total fat | Continuous extraction (single solvent, also called Soxhlet) | Time consuming. Extracts cannot be used for fatty acid studies. Incomplete extraction from many foods (dry analytical samples). Non-comparable | Applicable to low-moisture foods and non-cereal foods |

| | | | |
|-------------------------|---|--|--|
| | | value for cereals | |
| | Acid hydrolysis | Some hydrolysis of lipids. Extracts cannot be used for fatty acid studies | Applicable to all foods except dairy and high-sugar products |
| | Acid hydrolysis and capillary GLC | High cost. This method is NLEA-compliant | Applicable for most foods |
| | Mixed solvent extraction * | Complete extraction from most foods. Extract often needs clean-up | Applicable to most foods and extract can be used for fatty acid analysis |
| | Alkaline hydrolysis | | Validated for dairy foods only |
| | NIR | High cost. Requires extensive calibration against other methods | Established only for cereals |
| Fatty acids | HPLC | High cost | |
| | GLC * | Moderate to high cost | Applicable to all foods. If used for <i>trans</i> fatty acids, capillary techniques are required |
| | Infrared absorption (for trans fatty acids) | High cost. Some interference | Applicable to all foods |
| Total nitrogen /protein | Kjeldahl (for total nitrogen) * | Minor interference from inorganic nitrogen. Toxic wastes | Applicable to all foods |
| | Dumas (for total nitrogen) * | Limitations are high cost, the inclusion of inorganic nitrogen and analytical portion size | Applicable to all foods |
| | Radiochemical methods (for total nitrogen) | Very high cost of instrumentation | Applicable to most foods |
| | Formol titration; Biuret; Folin's reagent (for protein) | Specificity | Applicable to dairy products only |
| | Alkaline distillation (for protein) | Specificity | Applicable to cereals only |
| | Dye-binding (for protein) | Specificity | Applicable only to specific foods, and some cereals |

| | | | |
|--|---|--|--|
| | | | and legumes |
| | NIR (for protein) | High cost. Number of calibration sample | Applicable to some foods |
| Amino acids (AA) | GLC (preceded by acid hydrolysis for most AA. Alkaline hydrolysis required for tryptophan. Special hydrolysis conditions required for sulphur AA and acid-sensitive AA.) | Moderate to high cost. Choice of derivative is critical. AA need to be derivatized prior to chromatography | Applicable to most foods |
| | HPLC* (Preceded by acid hydrolysis for most AA. Alkaline hydrolysis required for tryptophan. Special hydrolysis conditions required for sulphur AA and acid-sensitive AA. AA usually derivatized prior to chromatography) | High cost | Applicable to all foods |
| | Ion-exchange chromatography* (Preceded by acid hydrolysis for most AA. Alkaline hydrolysis required for tryptophan. Special hydrolysis conditions required for sulphur AA and acid-sensitive AA.) | High cost. Hydrolytic losses of more labile AAs and slow release of branched chain AAs | Applicable to all foods |
| | LC-MS | High cost | Applicable to all foods |
| | Colorimetry (Tryptophan and S containing AA, lysine) | Not sensitive enough | Applicable to all foods |
| | Microbiological assays | Tedious, time-consuming, non reproducibility | Applicable to all foods |
| Alcohol | Distillation* | Interference with volatiles | Applicable to all foods |
| | GLC* | | Applicable to all foods |
| | Specific enzyme method* | | Applicable to all foods |
| Sugars, total (mono and disaccharides) | Density | Accurate for sucrose | Applicable to sugar solutions |
| | Refractive index | Empirical calibration required | Applicable to sugar solution |
| | Polarimetry | Close attention to standardized methods is essential | Applicable to single sugars or simple mixture only |

| | | | |
|------------------|---|--|--|
| | Reductiometric | Non-reducing sugars, sucrose and invert sugar mixtures | Applicable to reducing sugars |
| | Colorimetric | Specificity | Applicable to single sugars and simple mixtures |
| | Specific enzyme method * | Reagents can be expensive | Applicable to glucose and complex mixtures |
| | GLC | Need for derivatives | Can be applied to complex mixture |
| | HPLC * | Moderate to high cost. Choice of columns, detectors are crucial | Can be applied to complex mixture |
| Polyols | Specific enzymatic method | Specificity of enzymes | Limited to a few polyols only |
| | HPLC* | Moderate to high cost. Lack of standardized procedures; choice of column | Can be applied to complex mixture |
| | Microbiology | Acyclic polyol only | All foods |
| Oligosaccharides | Specific enzymatic procedures | Moderate to high cost | Applied for selective hydrolysis and separation |
| | GLC | Moderate to high cost. Choice of column | Can be applied to complex mixture |
| | HPLC | Moderate to high cost | Complex mixtures |
| Starch | Polarimetry | Needs very careful calibration | Applicable only to some cereal foods |
| | Dilute acid hydrolysis using a general sugar method | Interference from any NSP present. | Applicable to highly refined foods that are low in NSP |
| | Dilute acid hydrolysis and glucose-specific method | Presence of β -glucans | Applicable only to foods low in β -glucans |
| | Enzymatic hydrolysis and glucose specific method* | Choice of enzymes and conditions | Applicable to all foods |

| Dietary fibres | | | |
|----------------------------------|---|--|-------------------------|
| Total dietary fibre | AOAC method for dietary fibres (Prosky <i>et al.</i>) * - a enzymatic- gravimetric method | Time-consuming | Applicable to all foods |
| Non-starch polysaccharides (NSP) | Enzymatic hydrolysis and removal of starch. Acid hydrolysis of NSP, GLC, HPLC separation of component monosaccharides. Colorimetric analysis of monosaccharide (Englyst et al.) | Moderate to high cost. Resistant starch must be treated before hydrolysis. GLC requires preparation of derivatives. Gives only total values. This method is not robust | Applicable to all foods |
| Resistant starch | Enzymatic hydrolysis of starch before and after treatment with alkali or DMSO | Choice of enzymes and conditions | Applicable to all foods |

* recommended method

Inorganic material analysis

Applicable to all foods after defatting and drying, especially for food high in fat and/or water content

| Food component | Available method of analysis | Limitations |
|--|---|--|
| Total Ash | Dry ashing | Not suitable for mineral analysis of volatile minerals because of their partial loss |
| | Wet ashing | Small sample throughput |
| Cations | | |
| Na ⁺ , K ⁺ , Ca, Mg | Flame photometry | Interference |
| Na, K, Ca ⁺ , Mg ⁺ , Fe ⁺ , Cu ⁺ , Zn ⁺ , Mn ⁺ , Co ⁺ , Cr ⁺ | Atomic Absorption Spectrometry (AAS) with electrothermal furnace | Moderate to high cost. Interferences from anions; special suppression techniques |
| Se ⁺ | Hydride generation AAS | Moderate to high cost |
| | Fluorimetry | |
| all cations | Plasma emission spectrometry (= inductively coupled plasma spectroscopy ICP) ideally coupled with Mass spectrometry (MS)* | Very high cost. Matrix effects need to be controlled |
| K, Mg, Fe, Cu, Zn | Colorimetry | Extracting techniques. Difficult for K and Zn |
| Ca and Mg | Classical precipitation and titration | Size of analytical sample; skilled techniques |
| Anions | | |
| Phosphorus | Colorimetry | |
| | ICP-MS | Very expensive |
| Chloride | Titrimetric | |
| | Ion-specific electrode (ISE) | Interference |
| | ICP-MS | Very expensive |
| | Automated conductimetry | High cost |
| Iodine | Microdistillation | Laboratory contamination |
| | ISE | |
| | ICP-MS | Very expensive |
| | Alkaline dry-ashing | |
| | GLC | High cost |

FAO/INFOODS e-Learning Course on Food Composition Data

Lesson 3.3: Food Component Analysis

| | | |
|----------|------------------------------|----------------|
| Fluorine | Microdistillation | Time-consuming |
| | Ion-specific electrode (ISE) | |
| | Polarography | |
| Sulphur | Gravimetric | |
| | X-ray fluorescence | High cost |
| | ICP-MS | Very expensive |
| Nitrite | Colorimetry | |
| | Ion-specific electrode (ISE) | |
| Nitrate | HPLC | High cost |

Vitamin analysis

Applicable to all foods

| Food component | Available method of analysis | Limitations |
|------------------------|---------------------------------|---|
| Retinol | Colorimetry | Obsolete (Carr and Price, 1926). Low recoveries of retinoids |
| | HPLC * | Moderate to high cost. |
| Carotenoids | Open column chromatography | Identification of carotenoids. Lack of resolution of some geometrical isomers (lutein/zeaxanthin) and stereo-isomers (cis/trans). |
| | HPLC * | Moderate to high cost. Identification of carotenoids |
| Vitamin D | Bioassay | For low level only; animal facilities required |
| | Colorimetry | Lack of precision and sensitivity |
| | Gas-liquid chromatography (GLC) | New procedures under development |
| | HPLC* | High cost. Lipid interference; two stages, preparative followed by analytical separation needed for most foods |
| | Radio-immunoassay | High cost |
| Vitamin E | Colorimetry | Interfering compounds |
| | GLC | Derivation prior to chromatography required |
| | HPLC * | High cost. Extraction techniques |
| Vitamin K | Colorimetry | Lack of specificity |
| | Column chromatography, GC* | Moderate to high cost for GC |
| | HPLC* | High cost. Lipid interference |
| Vitamin C | Dye titration | Measure ascorbic acid only; pigments interfere; value lower as HPLC but comparable values for fresh fruits and vegetables |
| | Colorimetry | Measures inactive compounds also |
| | Fluorometry | Does not separate ascorbic and dehydroascorbic acid |
| | GLC | Derivatization prior to chromatography required |
| | HPLC * | High cost. Clean-up and separate detection of homologues add delays |
| Thiamin/ Riboflavin | Microbiological* | Time |
| | Fluorometry | |

FAO/INFOODS e-Learning Course on Food Composition Data

Lesson 3.3: Food Component Analysis

| | | |
|------------------|-----------------------------|--|
| | HPLC* | High cost |
| Niacin | Microbiological* | Time |
| | Colorimetry | Hazardous reagent |
| | HPLC* | High cost. |
| | | |
| Vitamin B6 | Microbiological* | Time; response to different vitamers may not be equal; total values only |
| | HPLC* | High cost |
| | Radiometric-microbiological | High cost |
| Vitamin B12 | Microbiological* | |
| | Radio-isotopic | High cost |
| Folates | Microbiological * | Response to different vitamers may not be equal; total values only |
| | HPLC | High cost. Not all vitamers measured properly |
| | LC-MS | Very high cost, but this method is able to quantify the different isomers of folates |
| Pantothenic acid | Microbiological* | |
| | HPLC | High cost |
| Biotin | Microbiological* | |
| | Isotope dilution | High cost |
| | Radiometric-microbiological | High cost |
| | Radio-immunoassay | High cost |
| | Protein-binding | High cost |
| | HPLC | High cost |

* recommended method

Analysis of other components

| Food component | Available method of analysis | Limitations |
|--|---|---|
| Hemagglutinins/Lectins | RBC agglutination | Not all blood samples of one animal species will react in an identical manner owing to the existence of several blood groups. Agglutination dilution test semi-quantitative |
| | Spectrophotometric methods | |
| | Radioactive labelling of lectin molecules | Requires specialized handling |
| Phytic acid | Anion exchange | Inability to resolve inositol phosphates adequately |
| | HPLC | High cost |
| | GLC | Detects derivatize volatile inositol phosphate forms only after separation by ion-exchange chromatography |
| | Capillary electrophoresis | Not applicable to all foods |
| | NMR-MS | High costs. Specialized application |
| Oxalates | Capillary electrophoresis | Not good for low oxalate content <1.8 mg/100 g. Meant for routine monitoring |
| | Ion chromatography | High running cost |
| | GLC | Some forms of oxalate are difficult to methylate; high instrument cost |
| | Enzymatic | Not applicable to all foods |
| | Colorimetry (AOAC) | Interference from other acids |
| | HPLC | High cost |
| Tannins (grouped into condensed tannins also called proanthocyanidins, hydrolysable and derived tannins) | UV-Spectrometry Vanillin HCL reagent | Parameters like extraction time, temperature, vanillin and HCL concentration need to be strictly controlled |
| | UV-Spectrometry Folin-Denis reagent | Non-specific as they can react with any phenol present in plant tissue |
| | UV-Spectrometry Prussian blue reagent | Non-specific as they can react with any phenol present in plant tissue, qualitative test |

| | | |
|---|--|--|
| | HPLC | Modest success for smaller compounds of derived tannins |
| | Colorimetry | Limited to basic compounds of hydrolysable tannins |
| Saponins | Spectrophotometric method | Not suitable for determination of medicagenic acid for which titrimetric method for the quantitative determination of this aglycone content has to be employed |
| | Bioassays | |
| | HPLC | Identification of individual saponins |
| Trypsin inhibitor | Colorimetric | Does not differentiate between the different protease inhibitors |
| | ELISA method using monoclonal antibodies derived from mice | |
| Flavonoids | HPLC | Sample hydrolysis required for optimum resolution and quantisation of quercetin, kaempferol, myricetin, luteolin and apigenin. Separate extraction without hydrolysis required for analysis of anthocyanidins and flavan-3-ols |
| | LC-MS | Hydrolysis not required as long as masses of individual flavonoid conjugates differ by more than mass resolution of mass spectrometer |
| Isoflavones ¹ and coumestrol | HPLC | Complex conjugates, and their numbers may be difficult to resolve with some reversed-phase columns and simple mobile-phase programs (isocratic) |
| | LC-MS | Hydrolysis not required as long as masses of individual conjugates differ by more than mass resolution of mass spectrometer |
| Lignans | HPLC | Isolariciresinol, pinoresinol, secoisolariciresinol and matairesinol |
| | GLC-MS | Only for matairesinol, secoisolariciresinol and shonanin in foods as trimethylsilyl derivatives |

¹ Isoflavones are a subclass of flavonoids but because they have different and unique biological activities than other subclasses of flavonoids, they are analysed and compiled as a separate group