



A.C. Camargo Cancer Center

Centro Integrado de Diagnóstico, Tratamento, Ensino e Pesquisa

Immunomics and cancer

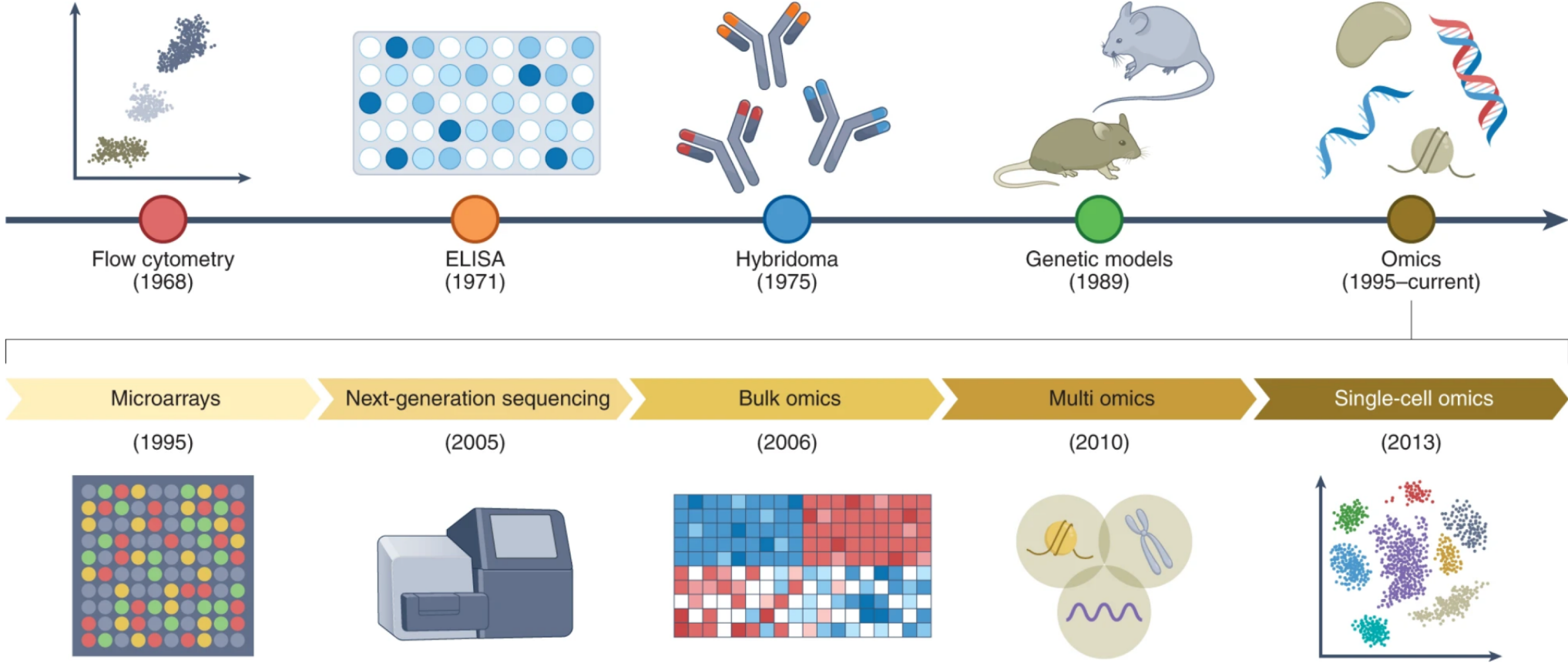
Gabriela Kinker, PhD

gabriela.kinker@accamargo.org.br

Translational Immuno-oncology Lab.

A.C. Camargo Cancer Center

Timeline of methodologies used in immunology research



Summary

Part 1

Introduction to main omics methodologies and data analysis tools.

- Conventional RNA-seq.
 - Infer the abundance of different immune cell types.
 - Evaluate the repertoire of T and B cell receptors.
- RNA-seq alternative: hybridization assays.
- scRNA-seq e scTCR-seq/scBCR-seq.
- Spatial transcriptomics.

Part 2

Examples of how to use these methodologies in your research project.

- Identifying subgroups of tumor-infiltrating T and B cells using scRNA-seq.
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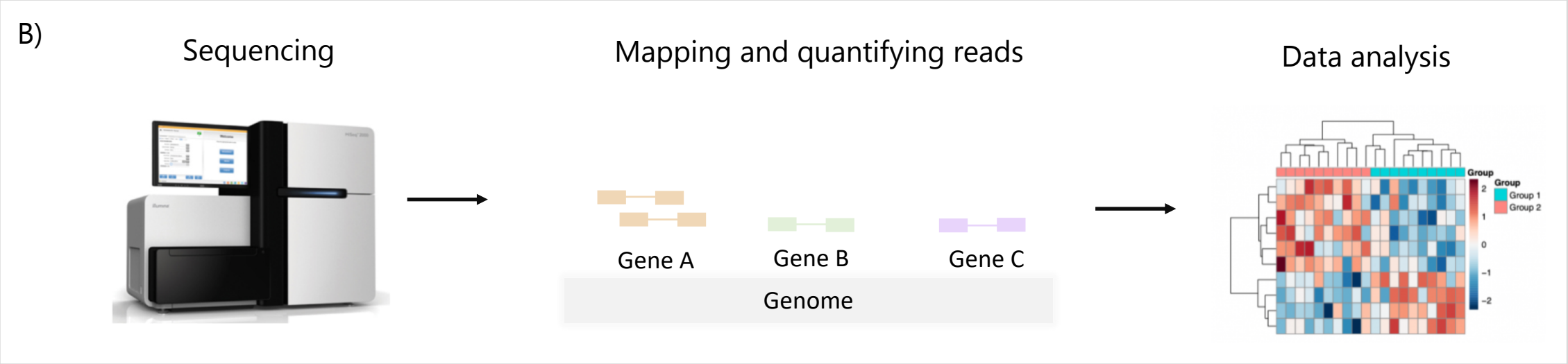
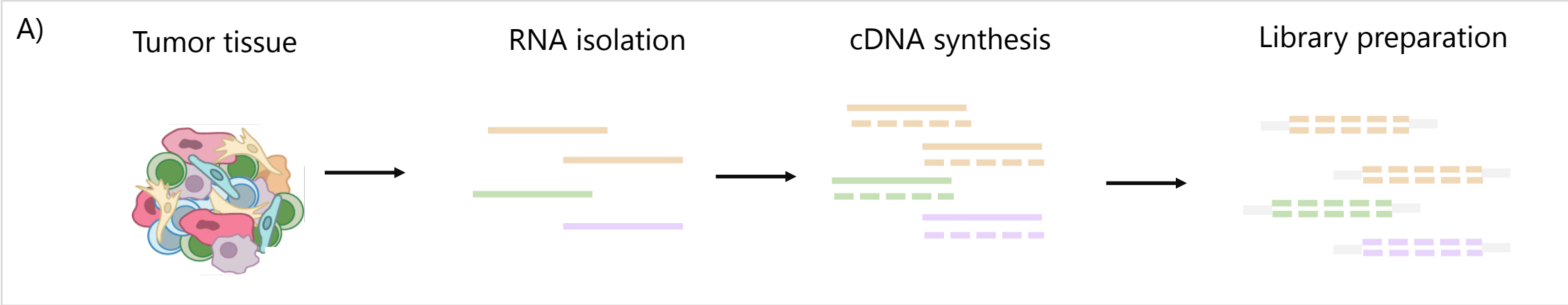
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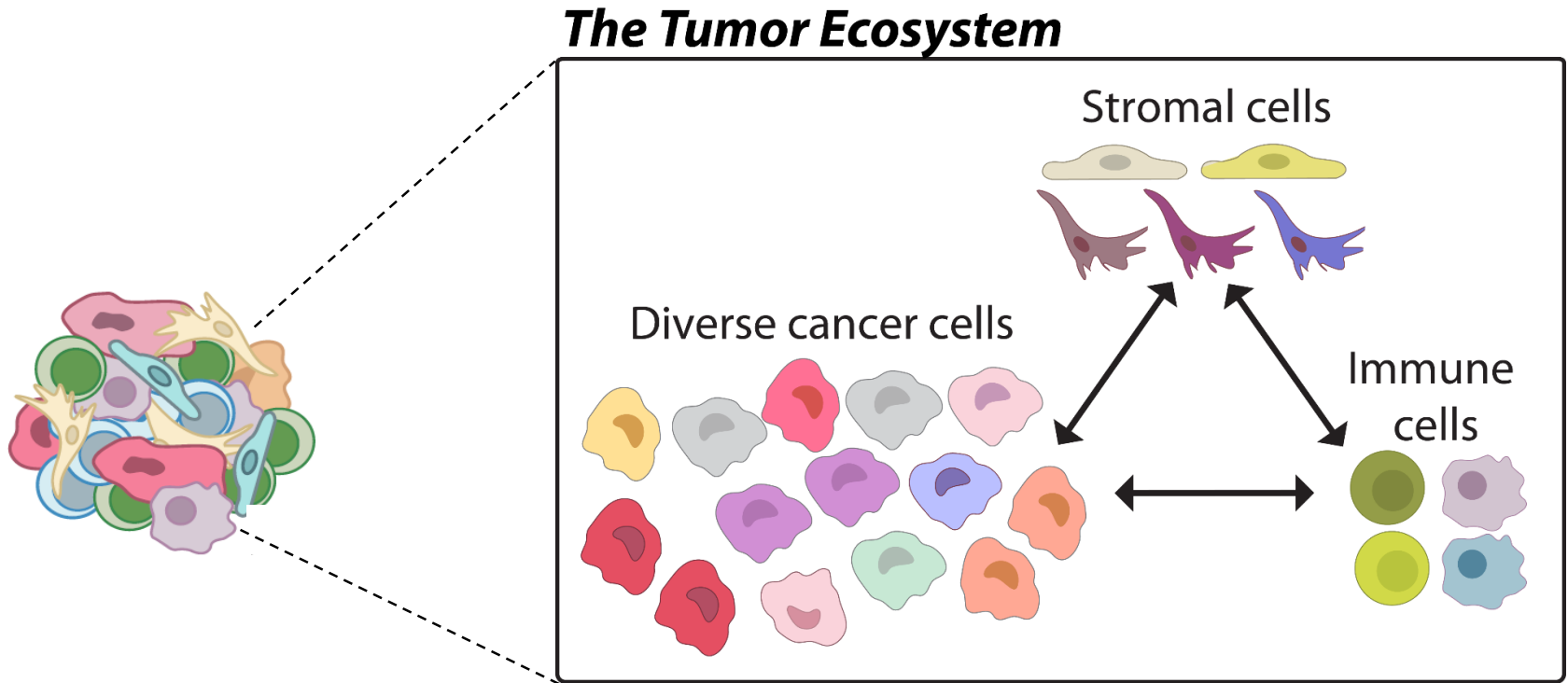
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Conventional RNA-seq ("bulk" RNA-seq)



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Bulk RNA-seq

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Tools to quantify the abundance of immune cells in a tissue profiled by RNA-seq

Gene signature-based approaches

- For each immune cell type of interest, there is a list of marker genes.
- The abundance of each immune cell type is estimated separately, based on the expression of marker genes.
- Algorithm options for calculating the abundance scores:
 - **MCPcounter**: mean expression of marker genes.
 - **xCell**: single sample GSEA.
- Compare the abundance of each immune cell type between samples.

Deconvolution-based approaches

- First, define a mixture of immune cell types which will be jointly evaluated. For each cell type, there is a list of marker genes.
- Describe the gene expression of a sample as the weighted sum of the expression profiles of the admixed cell types.
- Algorithm options for calculating the abundance scores:
 - **TIMER**: Linear least square regression.
 - **EPIC**: Constrained least square regression.
 - **CIBERSORT**: ν -Support Vector Regression.
- Compare the abundance immune population between samples and also within a sample.

TIMER 2.0

Modules exploring the association between immune infiltrates and genomic changes or clinical outcome in TCGA:

Gene Mutation sCNA Outcome

Gene Expression:

Immune Infiltrates:
T cell CD8+

Submit

Instruction: Gene module allows users to select any gene of interest and visualize the correlation of its expression with immune infiltration level in diverse cancer types. Once your interested gene and immune infiltrates submitted, a heatmap with numbers will show the purity-adjusted spearman's rho across various cancer types. When you click your interested cell on the heatmap, a scatter plot will pop out to present the relationship between infiltrates estimation value and gene expression. Tumor purity is a major confounding factor in this analysis, since most immune cell types are negatively correlated with tumor purity. Therefore, we recommend users to select the "Purity Adjustment" option, which will use the partial Spearman's correlation to perform this association analysis. Specially, for methods like EPIC and quanTIseq, which provide cell fractions referred to total cells, tumor purity and immune infiltration are necessarily negatively correlated, hence there is no need to adjust purity for the association analysis using the estimations from EPIC and quanTIseq.

NATIONAL CANCER INSTITUTE
THE CANCER GENOME ATLAS

TCGA BY THE NUMBERS

TCGA data describes



DIFFERENT
TUMOR TYPES

...including



RARE
CANCERS

...based on paired tumor and normal tissue sets
collected from



PATIENTS

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Evaluate the repertoire of T and B cell clones in a tissue profiled by RNA-seq: TRUST 4.0

Select reads that have significant overlap with V, J, C genes

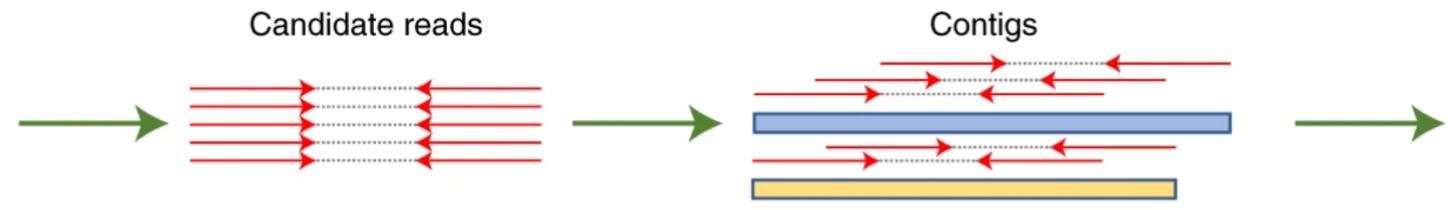
Assemble TCRs and BCRs based from selected reads by creating contigs (series of reads with overlapping sequences)

Align the contig to sequences in the ImmunoGeneTics (IMGT) database for annotating the V, J and C genes.

a



Tumor sample



UTR1	IGHV1-2	CDR3	IGHJ1	IGHG1
UTR2	TRBV2	CDR3	TRBJ2-3	TRBC
UTR3	TRGV4	CDR3	TRGJ2	TRGC

Evaluate the repertoire of T and B cell clones in a tissue profiled by RNA-seq

Outputs

```
graph TD; Outputs[Outputs] --- TCR[TCR e BCR]; Outputs --- BCR[BCR];
```

TCR e BCR

- Receptor diversity
- Rate of clonal expansion
- Shared clonotypes
- Clone reactivity to common viral antigens (VDJdb).

BCR

- Rate of somatic hypermutation
- Rate of Ig class switch recombination
- Direction of Ig class switch

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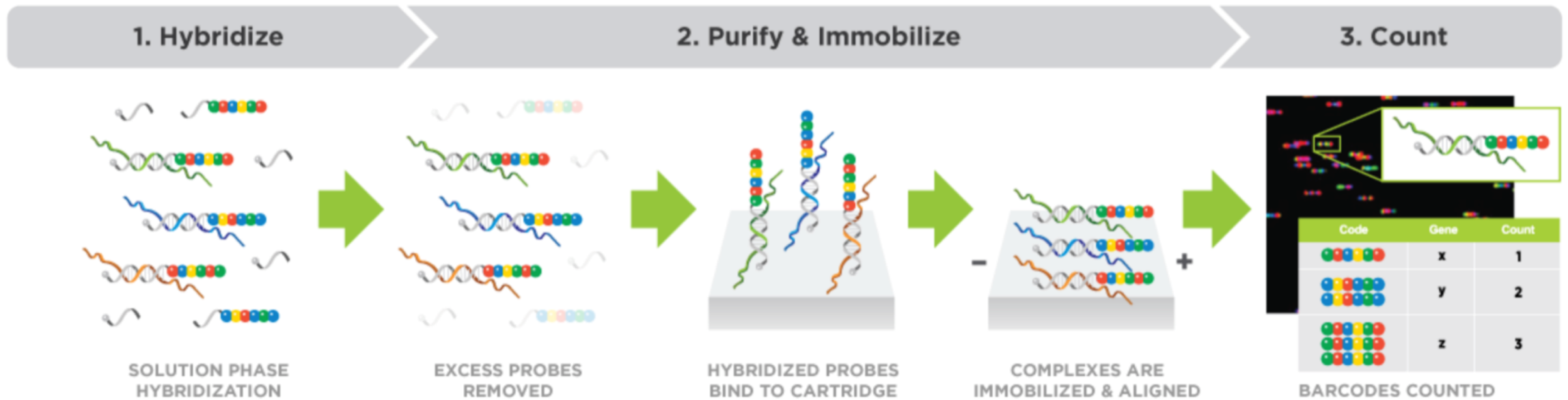
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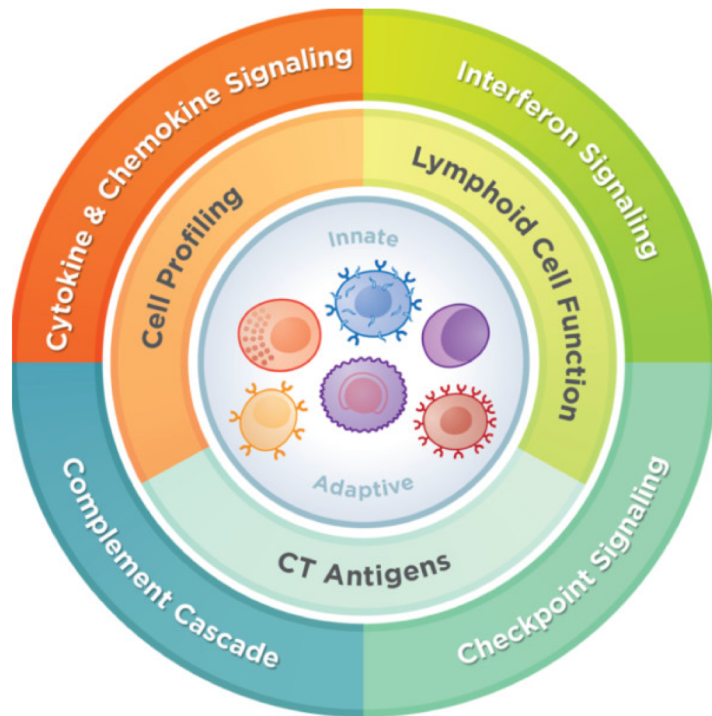
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Hybridization assay: a RNA-seq alternative that works well with FFPE samples

nCounter Workflow for Gene Expression Assays



Hybridization assay: a RNA-seq alternative that works well with FFPE samples



PanCancer Immune Profiling

Human Mouse

770 immune profiling genes for the identification of different immune cell types, key checkpoint inhibitors, cancer antigens, genes for measuring the immune response & up to 40 internal reference controls. [show less](#)

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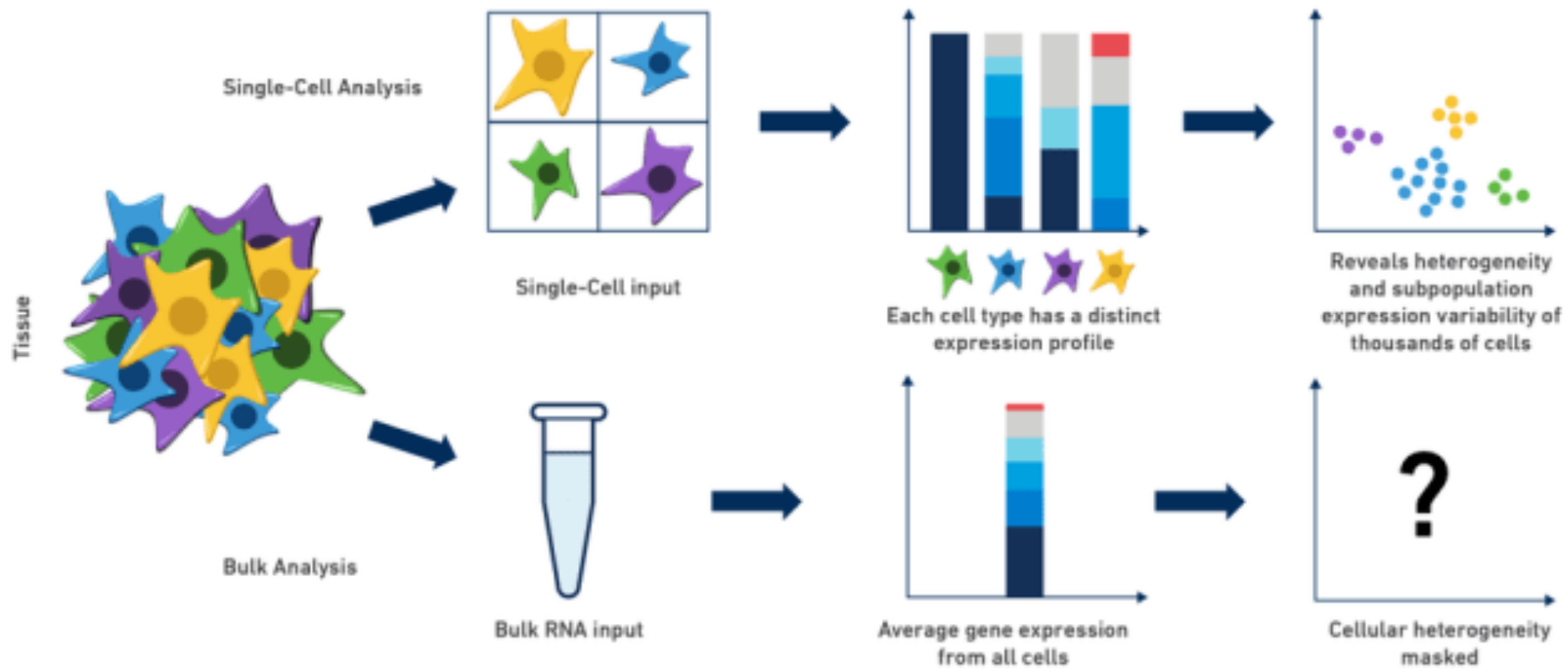
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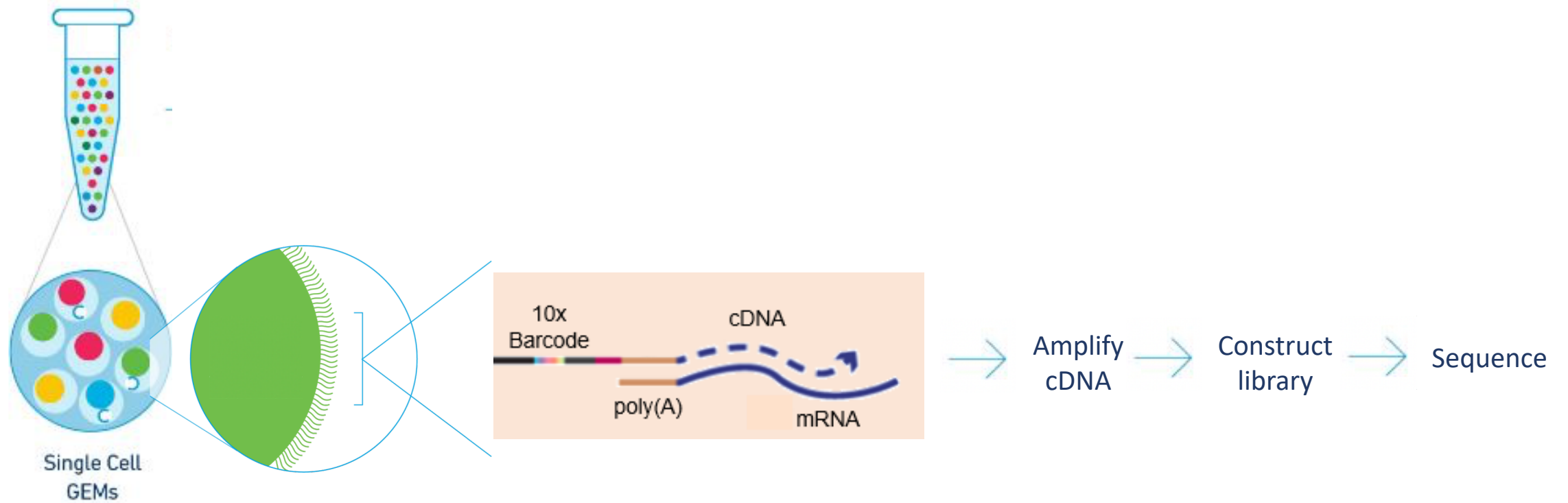
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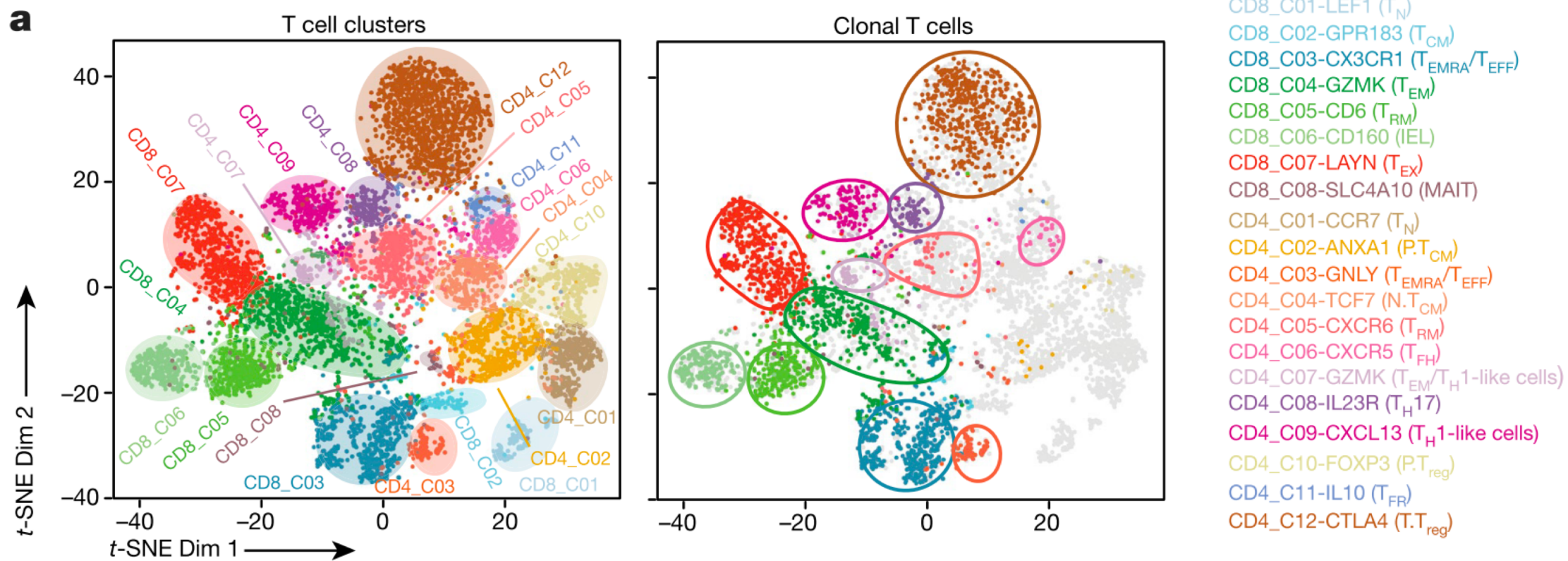
scRNA-seq vs. Bulk RNA-seq



scRNA-seq workflow



scRNA-seq coupled to scTCR-seq/scBCR-seq



scRNA-seq coupled to scTCR-seq/scBCR-seq

Examples of analyses using tumor samples

- Characterize the expression profile of tumor reactive clones (i.e. clonally expanded).
- Infer trajectories of cellular differentiation using algorithms such as pseudotime.
- Determine whether expanded clonotypes in the tumor tissue could also be detected in the blood.
- Evaluated expanded clones in the tumor microenvironment before and after treatment.
- Detection of bystander tumor-infiltrating lymphocytes that recognize common viral antigens.
- Evaluate the antibody affinity maturation of B cells (i.e. somatic hypermutation and Ig class switch recombination)

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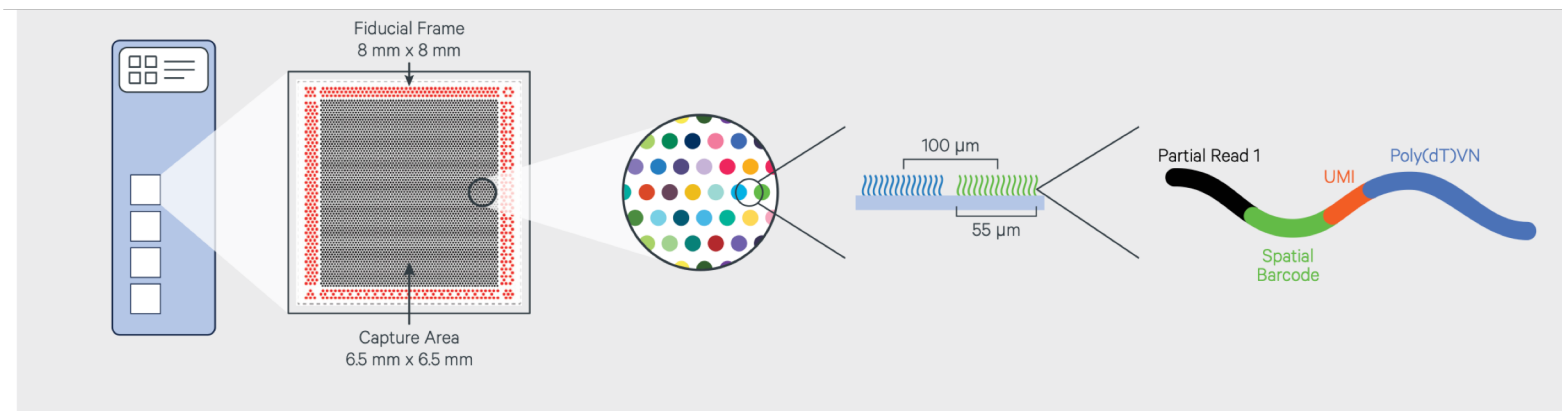
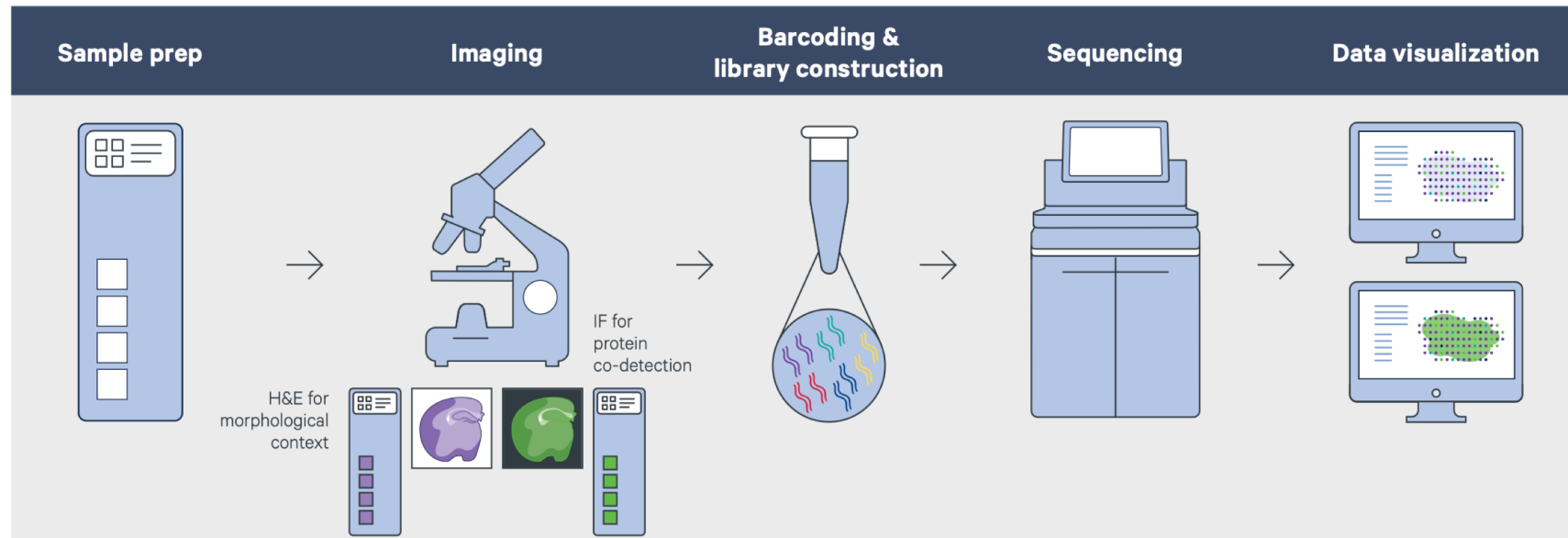
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Spatial transcriptomics workflow



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Mature tertiary lymphoid structures are key niches of tumour-specific immune responses in pancreatic ductal adenocarcinomas

St

Gabriela Sarti Kinker¹, Glauco Akelington Freire Vitiello¹, Ariane Barros Diniz², Mariela Pires Cabral-Piccin¹,  Pedro Henrique Barbosa Pereira¹, Maria Letícia Rodrigues Carvalho¹, Wallax Augusto Silva Ferreira^{1, 3}, Alexandre Silva Chaves¹, Amanda Rondinelli¹, Arianne Fagotti Gusmão¹, Alexandre Defelicibus¹, Gabriel Oliveira dos Santos⁴, Warley Abreu Nunes⁴, Laura Carolina López Claro⁵, Talita Magalhães Bernardo⁵, Ricardo Tadashi Nishio⁵, Adhemar Monteiro Pacheco⁵, Ana Carolina Laus⁶, Lidia Maria Rebolho Batista Arantes⁶, Julia Lima Fleck⁷, Victor Hugo Fonseca de Jesus⁸, André de Moricz⁵, Ricardo Weinlich², Felipe José Fernandez Coimbra⁹, Vladmir Cláudio Cordeiro de Lima⁸,  Tiago da Silva Medina^{1, 10}

Correspondence to Dr Tiago da Silva Medina, International Research Center, A.C.Camargo Cancer Center, São Paulo, 01508-010, Brazil; tiago.medina@accamargo.org.br

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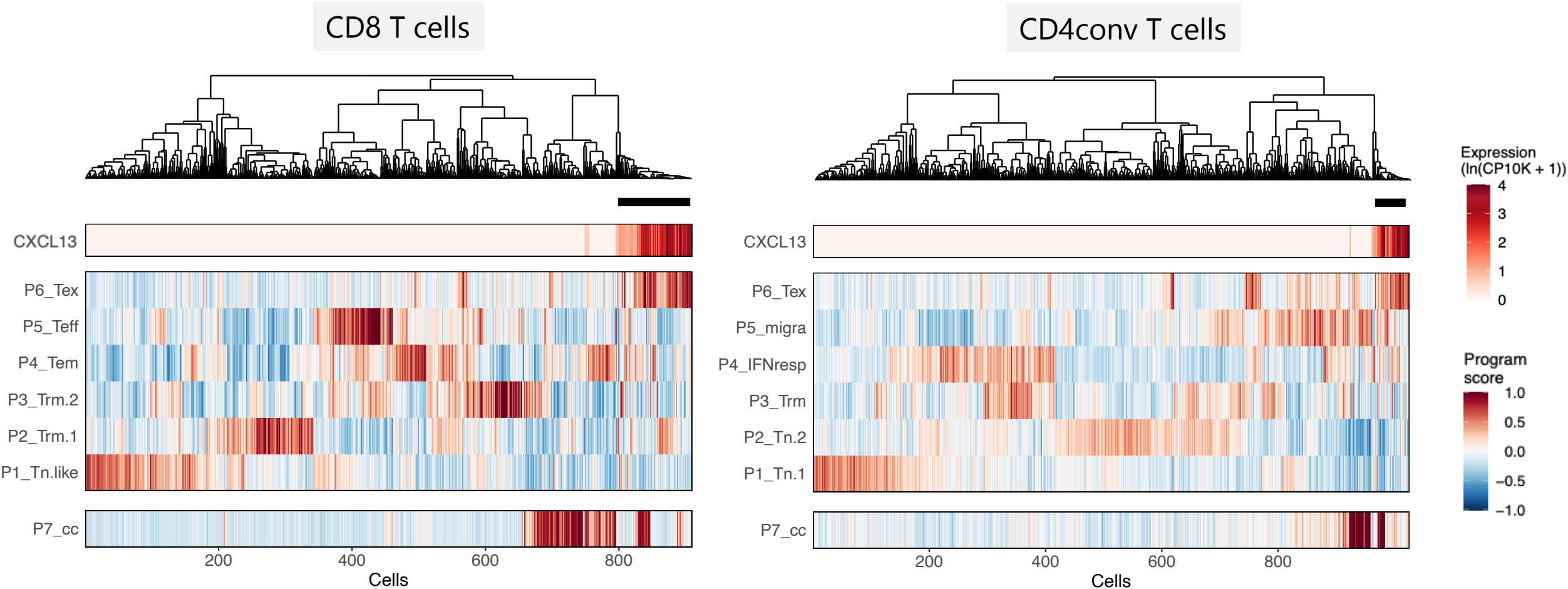
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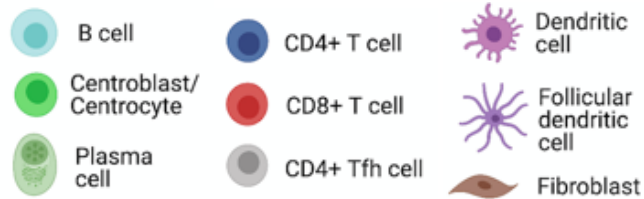
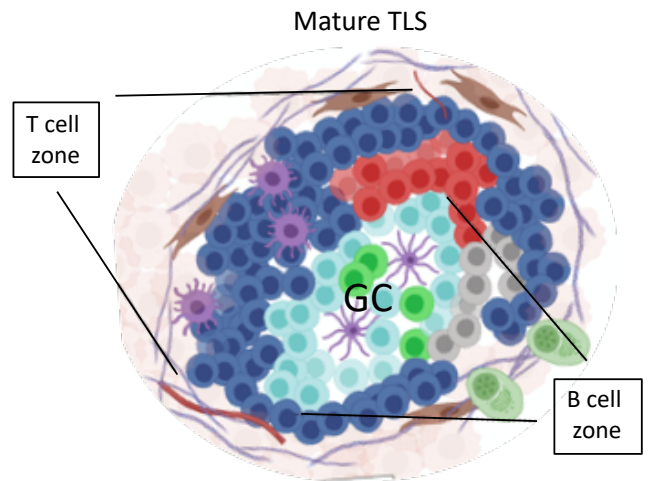
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Chronically activated tumor-infiltrating T cells produce de B cell chemoattractant CXCL13



scRNA-seq

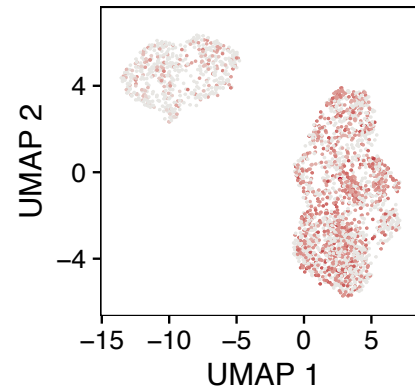
CXCL13 orchestrate the formation of tertiary lymphoid structures (TLSs) in the tumor tissue



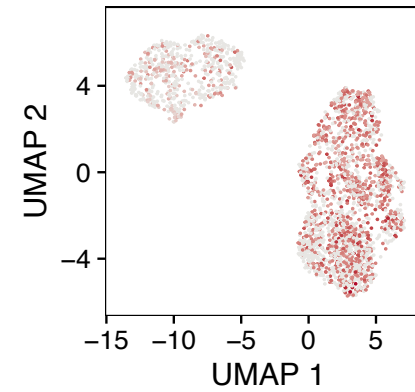
scRNA-seq

B cells

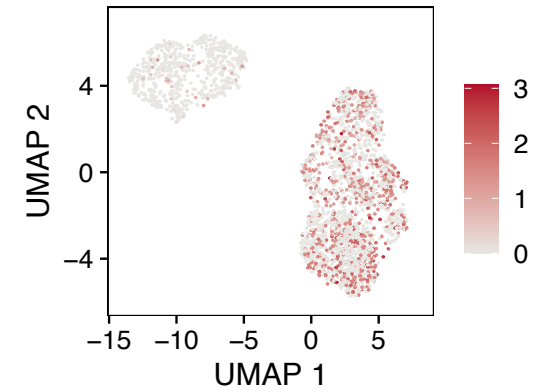
SELL



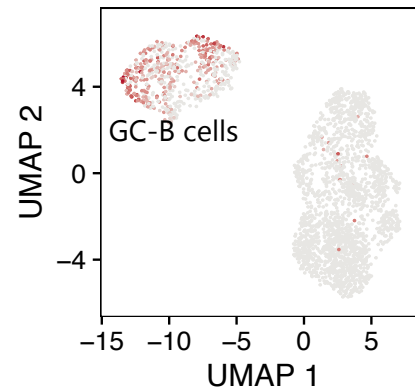
CD44



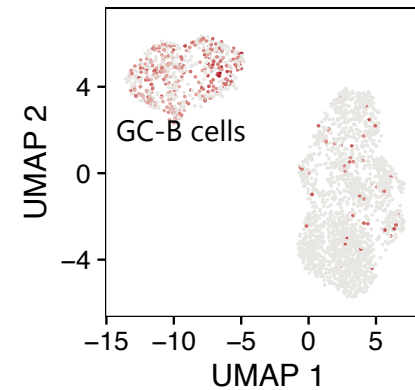
CCR7



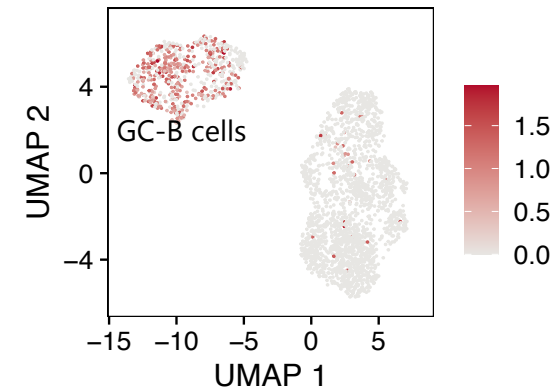
AICDA



BCL6



EZH2



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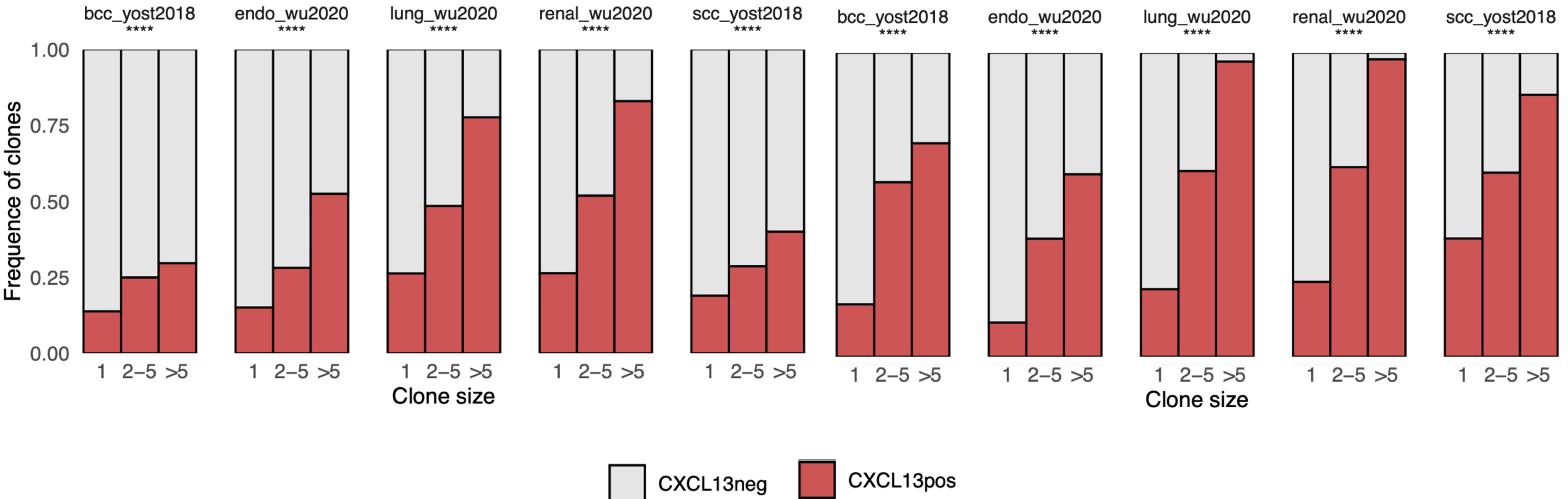
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CXCL13-producing T cells are tumor-reactive

CD8 T cells

CD4conv T cells



scRNA-seq + scTCR-seq

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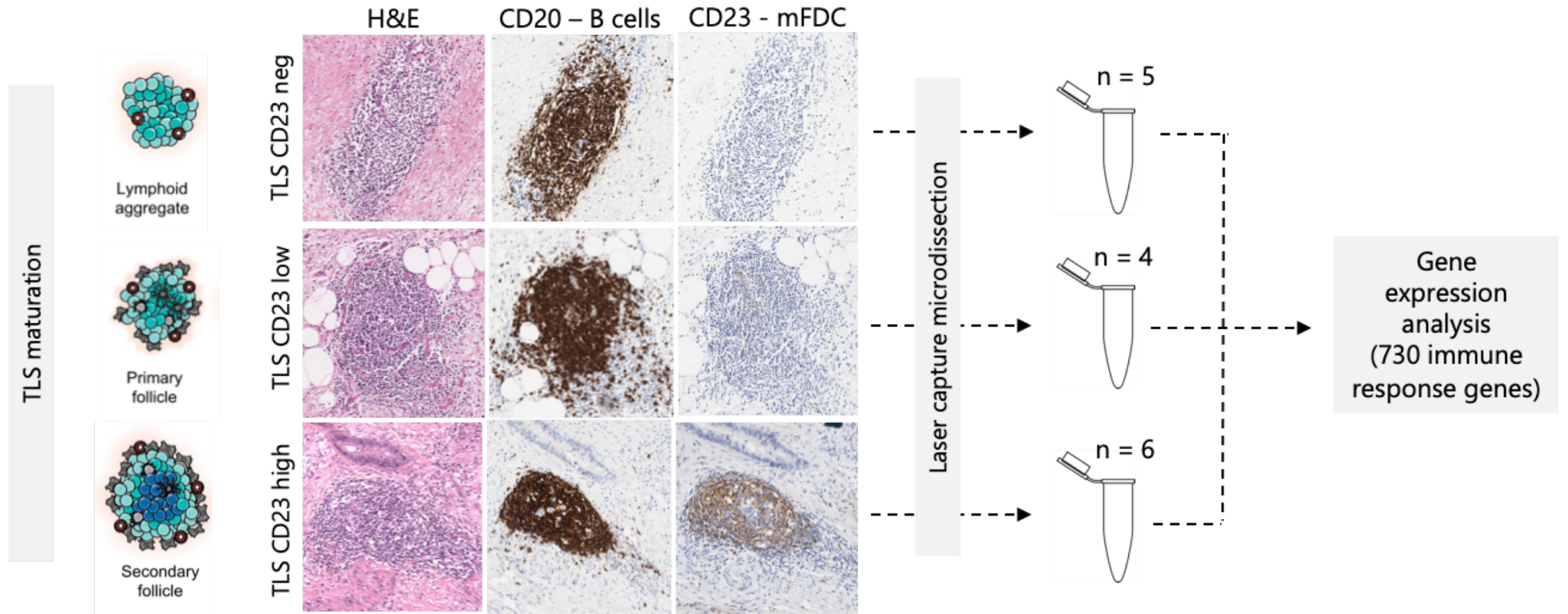
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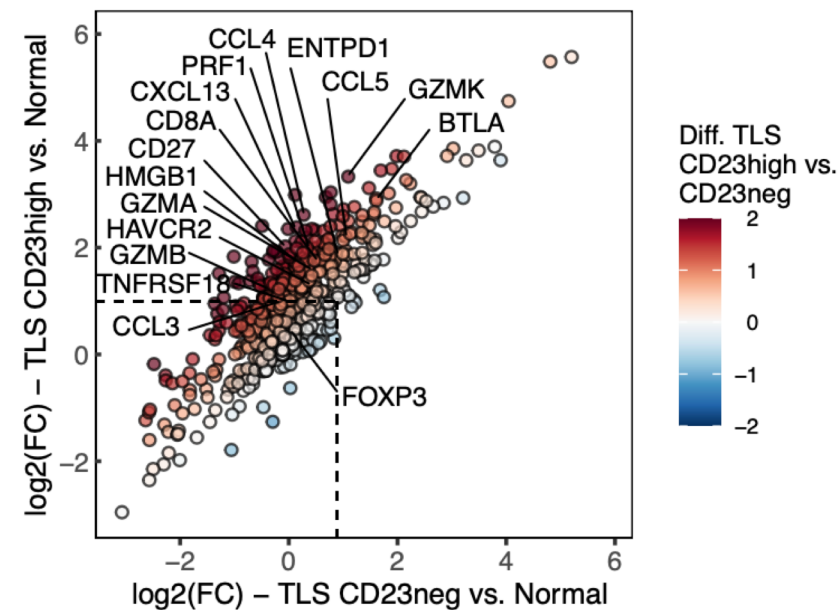
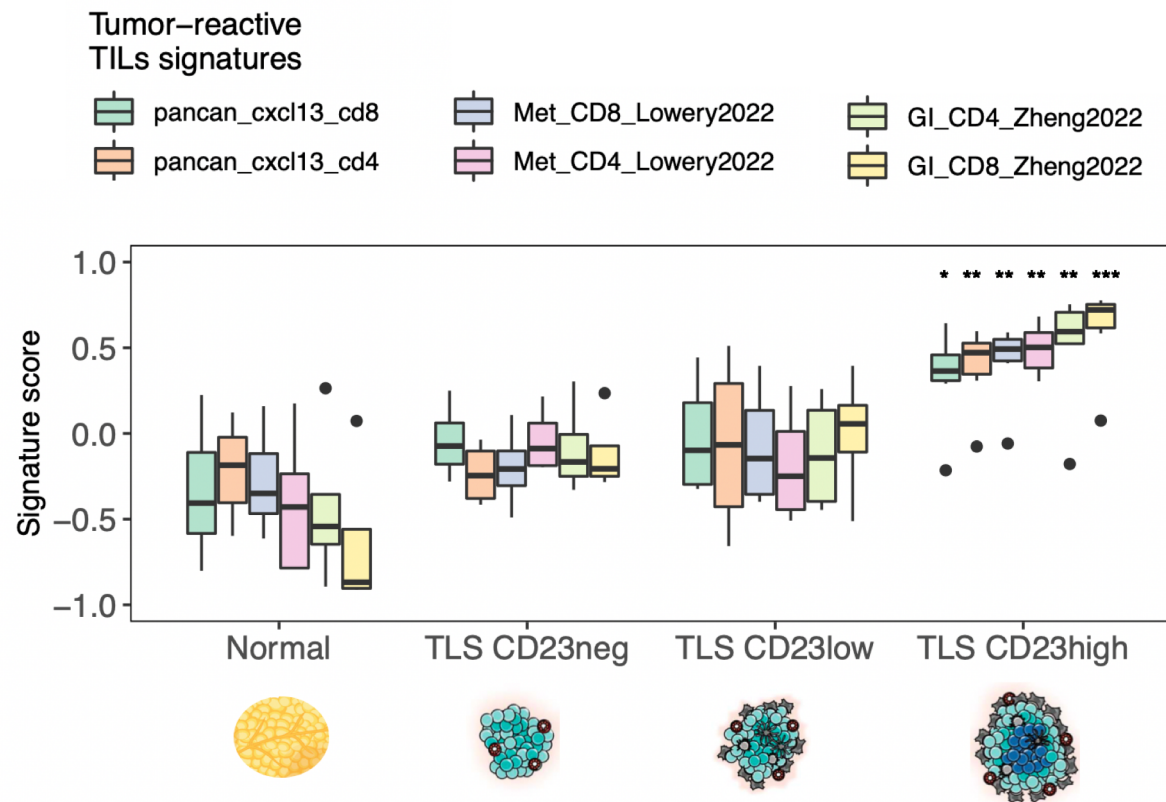
Expression profile of TLSs in different maturation stages



N = 41 treatment-naïve PDACs

Hybridization assay

Mature TLSs are enriched with tumor-reactive T cells



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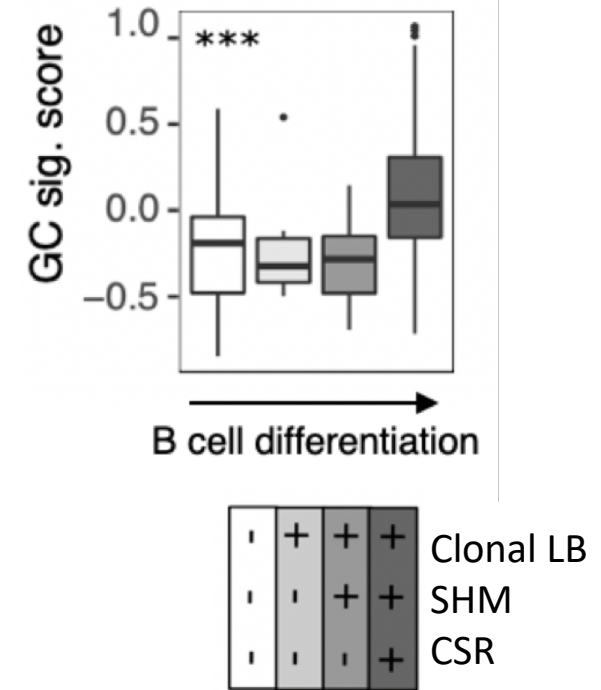
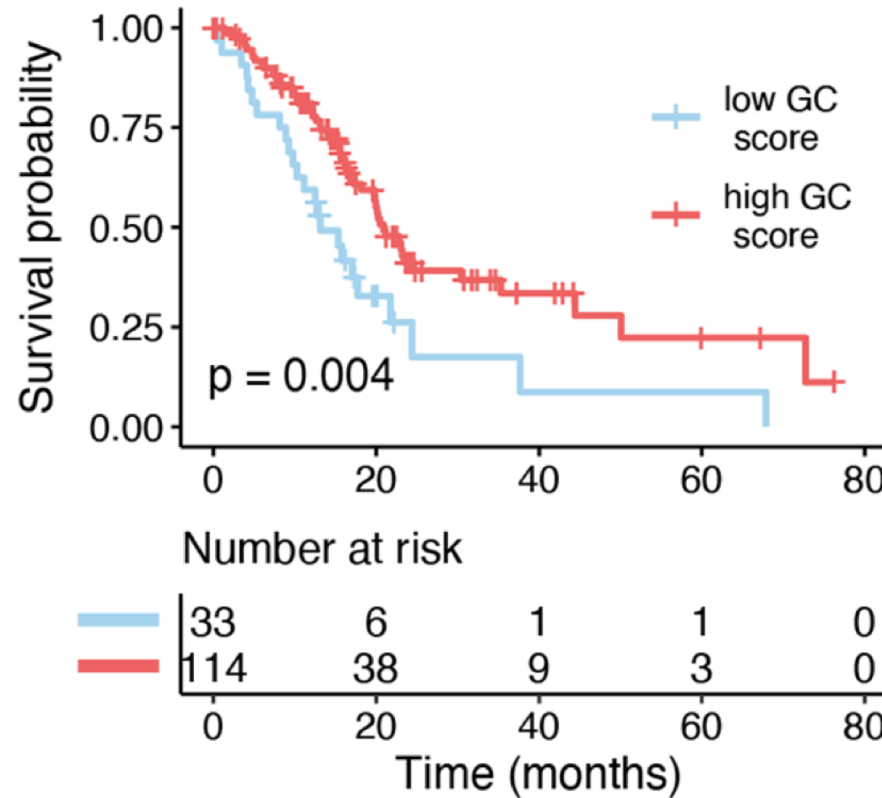
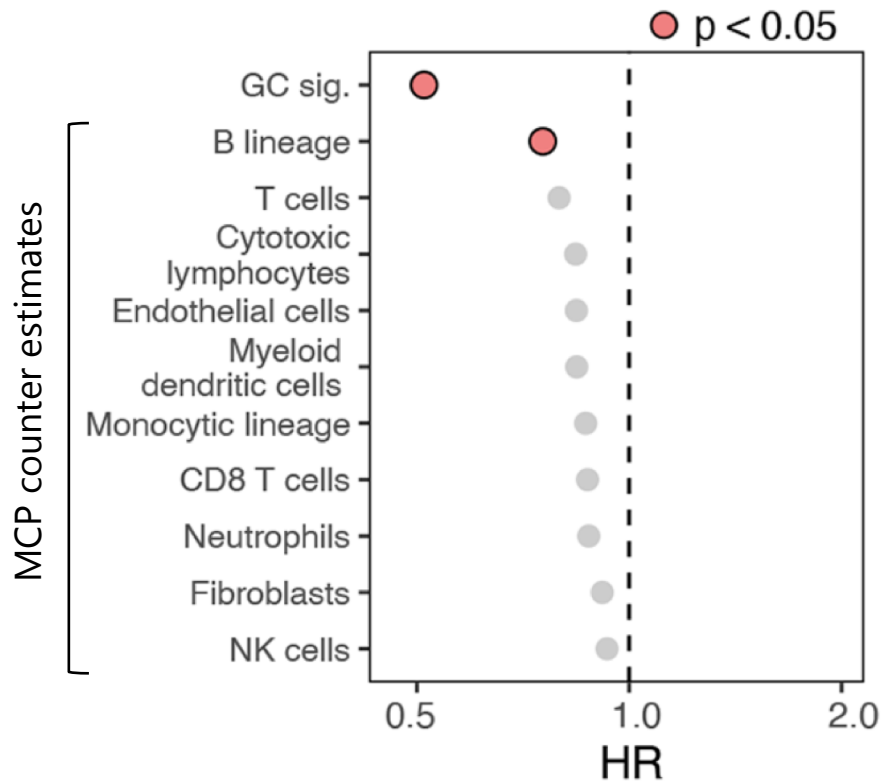
Part 2

Examples of how to use these methodologies in your research project.

- Identifying subgroups of tumor-infiltrating T and B cells using scRNA-seq.
- Evaluate the dynamics of clonal expansion of tumor-reactive T cells using scTCR-seq.
- Investigate the expression profile of microdissected TLSs using hybridization assays.
- Infer the abundance of different tumor-infiltrating lymphocytes using RNA-seq.
- Evaluate distribution of TLS-derived plasma cells throughout the tumor tissue using spatial transcriptomics.

The GC signature is associated with improved prognosis

TCGA bulk RNA-seq



Summary

Part 1

Introduction to main omics methodologies and data analysis tools.

- Conventional RNA-seq.
 - Infer the abundance of different immune cell types.
 - Evaluate the repertoire of T and B cell receptors.
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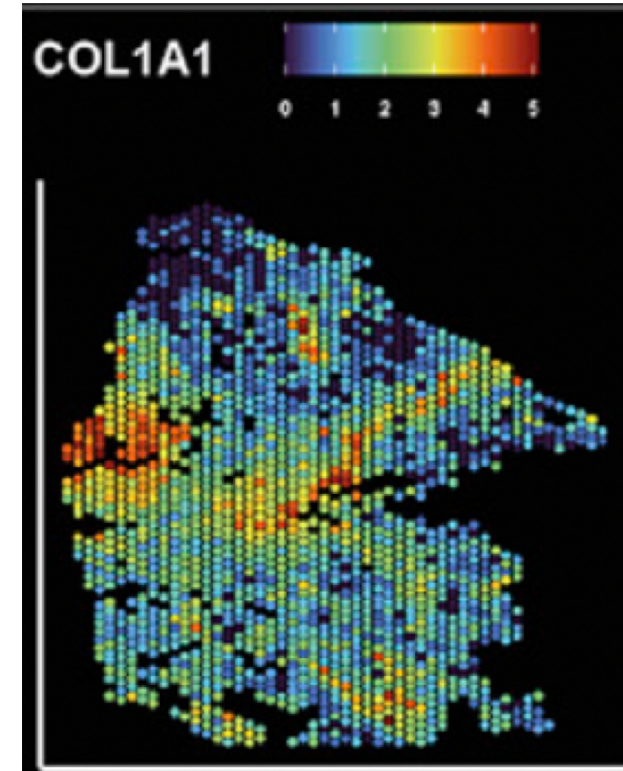
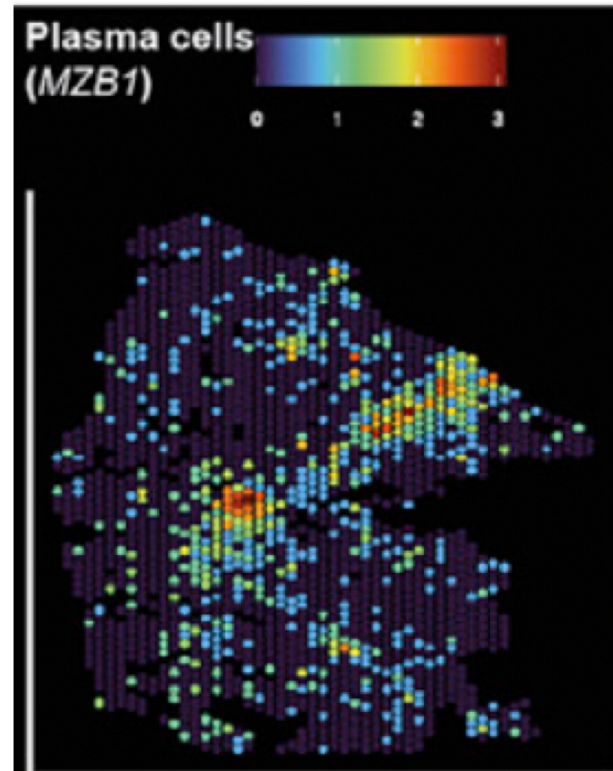
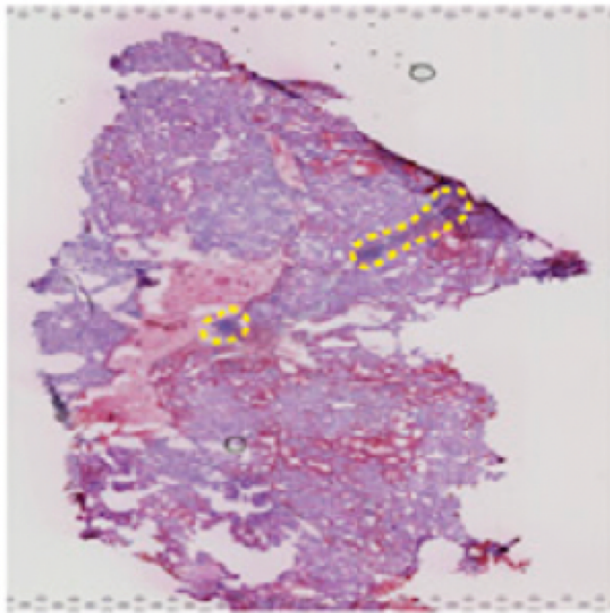
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Coming soon 😊

Renal clear cell carcinoma



! Reuse and Recycle !

There is a lot o public omics data out there.

Explore

Be creative

Have fun