LGN 5822 - Biometrical Genetics

L07 – Completely Randomized Design (CRD)

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Introduction

"And so it was ... borne in upon me that very often, when the most elaborate statistical refinements possible could increase the **precision** by only a few percent, yet a different design involving little or no additional experimental labour might increase the **precision** two-fold, or five-fold or even more"

R.A. Fisher (1962)



The Design of Experiments

By

Sir Ronald A. Fisher, Sc.D., F.R.S.

Honorary Research Fellow, Division of Mathematical Statistics, C.S.I.R.O., University of Adelaide: Foreign Associate, United States National Academy of Sciences; and Foreign Honorary Member, American Academy of Arts and Sciences: Foreign Member of the Swedish Royal Academy of Sciences, and the Royal Danish Academy of Sciences and Letters; Member of the Pontifical Academy; Member of the German Academy of Sciences (Leopoldina); formerly Galton Professor, University of London, and Arthur Balfour Professor of Genetics, University of Cambridge.

Fisher, Ronald Aylmer. "The design of experiments." *The design of experiments.* 1st Ed (1935).

Introduction

Suppose you want to make inferences about a given set of treatments

 The levels of the treatment factor can be randomly sampled from a larger population or can be chosen based on specific interest in each particular level: Condition that will be tested! • You carry out an experiment to compare the effects of *t* different treatments

 For doing so, you design a simple experiment with r independent replicates of each treatment level to measure these effects

• There are *tr* experimental units, in total!

Introduction

Completely Randomized Design (CRD)

Main Features

- In the Completely Randomized Design (CRD), randomization is absolutely necessary throughout the treatments
 - Treatments are distributed to experimental units completely at random
 - Every experimental unit has the same probability of receiving any treatment
 - Experimental units are randomized throughout the experiment
 - Consequently, there is no correlation between any two observations

• The simplest form of experimental scheme

• There is no restriction on the number of treatments examined

• Treatments can have different numbers of replicates, but balance is preferable

CRD uses only the basic principles of repetition and randomization

Example of Randomization

Given you have 4 treatments (A, B, C, and D) and 5 replicates, how many experimental units would you have?



Every experimental unit has the same probability of receiving any treatment!

• 4 treatments (A, B, C, and D) and 3 replicates

T2	Т3	Т4	Т2
T3	T2	T1	Т4
T1	Т4	T1	Т3

• 4 treatments (A, B, C, and D) and 3 replicates

• A different layout for the CRD:

Note

 Because there is no local control (error), the CRD is appropriate when experimental units are uniform

Advantages of a CRD

- Very flexible design (i.e. number of treatments and replicates is only limited by the available number of experimental units)
- Statistical analysis is simple compared to other designs
- Loss of information due to missing data is small compared to other designs due to the larger number of degrees of freedom for the error source of variation



Disadvantages of a CRD



- The experimental units must be homogeneous
- High estimate of variance due to error can be obtained because all variations (except treatments) are considered as random variation

Statistical model

 Let y_{ij} represent the observed response of the jth replicate (j = 1, ..., r) of the ith treatment (i = 1, ..., t)

Model

• Data from the CRD can be described with the following model:

$$y_{ij} = \mu + \tau_i + \varepsilon_{ij}$$

where μ is the intercept (an overall mean), τ_i is the treatment effect and ε_{ij} is the associated random error term

Statistical model

We assume that:

•
$$\varepsilon_{ij} \sim N(0, \sigma^2)$$
, for $i = 1, ..., t$ and $j = 1, ..., r$

•
$$cov\left(\varepsilon_{ij},\varepsilon_{i'j'}\right)=0$$

Errors are independent and identically distributed (i.i.d)

Statistical model

Consequently, for the observed values:

$$y_{ij} \sim N(\mu + \tau_i, \sigma^2)$$
, for $i = 1, ..., t$ and $j = 1, ..., r$

Model Fitting

• Unknown parameters μ , τ_i and σ^2 can be estimated via least squares, as **previously discussed** for standard linear models



Model Diagnostics

Important

 After fitting the model and before making inferences, it is important to check whether the model assumptions are met

Main assumptions of errors:

- Errors ε_{ij} are independent
- Errors ε_{ij} are homoscedastic (have the same variance)
- Errors are normally distributed



Model Diagnostics

• Errors ε_{ij} are unknown, so we use the estimated residual errors, or residuals

Checking Model Assumptions



17 genotypes and 3 replications

Checking Model Assumptions

- Remember that:
- $\varepsilon \sim N(0, \sigma^2 I)$

• $y \sim N(X\beta, \sigma^2 I)$

Checking Model Assumptions

• We will study the **distribution of the residuals** to verify validity of the assumptions

Residual Diagnostics

 We can use the raw residuals, which are deviations between the observed and fitted values:

$$\hat{\varepsilon}_{ij} = y_{ij} - \hat{y}_{ij}$$
$$\hat{\varepsilon}_{ij} = y_{ij} - \mu - \tau_i$$

Residual Diagnostics

Raw residuals vs fitted values

Residual Diagnostics

Raw residuals vs treatment levels

Residual Diagnostics

Normal quantile-quantile (Q-Q) plots

Normal Q-Q Plot

Data Transformation

 If (some of) the assumptions are not met, we can transform the observed data so that it is normally distributed and the variance is stabilized

Data Transformation : Box Cox

$$y_{ij}(\lambda) = \begin{cases} \frac{y_{ij}^{\lambda} - 1}{\lambda} & \text{if } \lambda \neq 0, \\ \log(y_{ij}) & \text{if } \lambda = 0 \end{cases}$$

y represents the original data $y(\lambda)$ is the transformed data λ is a parameter that determines the type of transformation to be applied

The idea is to find the value of λ that makes the transformed data come closest to a normal distribution

Data Transformation : Box Cox

The value that maximizes the likelihood, i.e., results in the best approximation to a normal distribution, is used to transform the data

Data table

Consider an experiment installed at the CRD with *i* treatments and *j* replicates

		Treatments				
Replicates	1	2		1		
1	Y ₁₁	Y ₂₁		Y _{I1}		
2	Y ₁₂	Y ₂₂		Y_{12}		
J	Y _{1J}	Y_{2J}		Y _{IJ}		
Total	T ₁	T ₂		T _I		

Data table

- Number of experimental units: $N = I \times J$
- Total for treatment *i*: $T_i = \sum_{j=1}^{J} Y_{ij} = Y_{i.}$

		Treatments				
Replicates	1	2		1		
1	Y ₁₁	Y ₂₁		Y _{I1}		
2	Y ₁₂	Y ₂₂		Y_{12}		
 J	 Y	 Yai		 Y		
Total	T ₁	T ₂		т,		

- Mean for treatment *i*: $\widehat{m}_i = \frac{T_i}{J}$
- General mean of the experiment: $\widehat{m} = \frac{G}{II}$

Analysis of Variance

 Allows you to decompose the total variation in the data into different variation components, a (effects of treatments and residuals)

Source of	Degrees of	Sum of	Mean
Variation	Freedom	Squares	Square
Treatments	t - 1	SS _{Trt}	$MS_{Trt} = \frac{SS_{Trt}}{t-1}$
Within treatments	t(r-1)	SS_{Within}	$MS_{Within} = \frac{SS_{Within}}{t(r-1)}$
Total	tr - 1	SS _{Total}	

Remember the error sum of squares (SSE) from past classes

Analysis of Variance

Source of Variation	Degrees of Freedom	Sum of Squares	Mean Square	Expected Mean Square
Treatments	t - 1	SS _{Trt}	MS_{Trt}	$\sigma^2 + r\sigma_t^2$
Within	t(r-1)	SS_{Within}	MS_{Within}	σ^2
Total	tr-1	SS_Total		

Variance Components

If we consider the treatment effects to be random, we can estimate the variance component using the method of moments, by setting the mean squares (MS) equal to the expected mean squares (EMS)

Variance Components

• The residual variance is simply estimated by $\hat{\sigma}^2 = MS_{Within}$

• We can then estimate σ_t^2 by $\hat{\sigma}_t^2 = \frac{MS_{trat} - MS_{within}}{r}$

Hypothesis Testing

• We want to test the null hypothesis of no treatment effects, i.e., $H_0 = \tau_i = 0$ for all *i*

Hypothesis Testing

Source of Variation	Degrees of Freedom	Mean Square	Expected Mean Square	F-Statistic
Treatments	t-1	MSTrt	$\sigma^2 + r \frac{\sum_i \tau_i^2}{\tau_i^2}$	
Within	t(r - 1)	MSwar	σ^2	
	<i>c</i> (<i>r</i> = 1)	Within		
lotal	tr-1			

Hypothesis Testing

Source of	Degrees of	Mean	Expected	F-Statistic
Variation	Freedom	Square	Mean Square	
Treatments	t-1	MS _{Trt}	$\sigma^2 + r \frac{\sum_i \tau_i^2}{t-1}$	$F = \frac{MS_{Trt}}{MS_{Within}}$
Within	t(r-1)	MS_{Within}	σ^2	
Total	tr-1		-	

F-Statistic

The F-Statistic follows an F distribution with t -1 and t(r - 1) degrees of freedom

Multiple Comparions

- The null hypothesis ($H_0 = \tau_i = 0$ for all *i*) is tested against the alternative that $\tau_i \neq 0$ for at least one of the treatments
 - If H_0 is rejected, we need to determine which of the treatments differ
 - This can be accomplished via pairwise comparisons between all possible combinations of treatments

Multiple Comparions

• We thus perform multiple comparisons between the treatment means

Tukey, Bonferroni, Scheffé, Dunnett's Test

 We will work with yield data of 17 rice genotypes, measured in three replications each

- This data set can be found in the R package *agridat* and is a subset of the data from Gomez & Gomez (1984) Statistical Procedures for Agricultural Research. Wiley-Interscience
- Let y_{ij} denote the yield of the *i*th rice hybrid, measured from replicate *j*

- Use the R function read.csv to import the data
- Fit the model with fixed effects #Use Im() function and summary()
- Check if model assumptions are met #Check raw residuals vs fitted values

- Build the ANOVA table and test the null hypothesis of no difference between the rice genotypes
- Use multiple pairwise comparisons to assess which genotypes differ
- Fit the model with random effects

Graphic library(lattice) xyplot(yield ~ reorder(gen, yield), data = dados, type = c("p", "a"))

Fit the model with fixed effects
 #Use Im() function and summary()

```
# Fit the model with fixed effects
fm <- lm(yield ~ gen, data = dados) # fixed effects
fm
summary(fm)</pre>
```

Fit the model with fixed effects
 #Use Im() function and summary()

> summary(fm)

Call: lm(formula = yield ~ gen, data = dados)

Residuals:

Min 1Q Median 3Q Max -0.87467 -0.28683 -0.01633 0.27500 1.09433

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)	
(Intercept)	5.8483	0.2867	20.402	< 2e-16	***
genG17	2.0473	0.4054	5.050	1.48e-05	***
genG18	2.0913	0.4054	5.159	1.07e-05	***
genG19	1.3300	0.4054	3.281	0.002397	**
genG20	0.7083	0.4054	1.747	0.089615	
genG21	1.1917	0.4054	2.940	0.005872	××
genG22	1.5513	0.4054	3.827	0.000530	***
genG23	1.9607	0.4054	4.837	2.80e-05	***
genG24	1.6793	0.4054	4.143	0.000215	***
genG25	1.8563	0.4054	4.579	5.99e-05	***
genG26	1.0453	0.4054	2.579	0.014423	*
genG27	0.9090	0.4054	2.242	0.031574	*
genG28	0.7557	0.4054	1.864	0.070968	
genG29	1.6380	0.4054	4.041	0.000288	***
genG30	1.5007	0.4054	3.702	0.000754	***
genG31	1.6943	0.4054	4.180	0.000193	***
genG32	1.5370	0.4054	3.791	0.000586	***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 0.4965 on 34 degrees of freedom Multiple R-squared: 0.6395, Adjusted R-squared: 0.4699 F-statistic: 3.77 on 16 and 34 DF, p-value: 0.0005615

 Check if model assumptions are met #Check raw residuals vs fitted values

```
# Raw residuals vs fitted values:
plot(residuals(fm) ~ fitted(fm))
```


fitted(fm)

 Check if model assumptions are met #Check raw residuals vs fitted values

Q-Q plot: Normal Q-Q Plot qqnorm(stdres(fm)) 0 0 0 \sim Sample Quantiles 0 $\overline{\gamma}$ P 0 -2 -1 0 2 1

Theoretical Quantiles

```
Anova
```

```
#Anova
anova(fm)
```

Calculate the estimated means for the different levels of the variable

```
# Obtaining estimated marginal means:
library(emmeans)
(fm_means <- emmeans(fm, "gen"))
plot(fm_means)
G32-
```


Pairwise comparisons with Tukey adjustment

#Pairwise comparisons
Pairwise comparisons with Tukey adjustment:
pairs(fm_means)

> # Pairwis	se compart	isons v	vitł	ı Tukev a	adiustment:
> pairs(fm_	_means)				
contrast	estimate	SE	df	t.ratio	p.value
G16 - G17	-2.0473	0.405	34	-5.050	0.0015
G16 - G18	-2.0913	0.405	34	-5.159	0.0011
G16 - G19	-1.3300	0.405	34	-3.281	0.1372
G16 - G20	-0.7083	0.405	34	-1.747	0.9312
G16 - G21	-1.1917	0.405	34	-2.940	0.2657
G16 - G22	-1.5513	0.405	34	-3.827	0.0395
G16 - G23	-1.9607	0.405	34	-4.837	0.0027
G16 - G24	-1.6793	0.405	34	-4.143	0.0178
G16 - G25	-1.8563	0.405	34	-4.579	0.0056
G16 - G26	-1.0453	0.405	34	-2.579	0.4693
G16 - G27	-0.9090	0.405	34	-2.242	0.6904
G16 - G28	-0.7557	0.405	34	-1.864	0.8913
G16 - G29	-1.6380	0.405	34	-4.041	0.0232
G16 - G30	-1.5007	0.405	34	-3.702	0.0535
G16 - G31	-1.6943	0.405	34	-4.180	0.0162
G16 - G32	-1.5370	0.405	34	-3.791	0.0431
G17 - G18	-0.0440	0.405	34	-0.109	1.0000
G17 - G19	0.7173	0.405	34	1.769	0.9245
G17 - G20	1.3390	0.405	34	3.303	0.1310
G17 - G21	0.8557	0.405	34	2.111	0.7706
G17 - G22	0.4960	0.405	34	1.224	0.9974
G17 - G23	0.0867	0.405	34	0.214	1.0000
G17 - G24	0.3680	0.405	34	0.908	0.9999
G17 - G25	0.1910	0.405	34	0.471	1.0000

Pairwise comparisons with Tukey adjustment

Pairwise comparisons with Bonferroni correction: pairs(fm_means, adjust = "bonferroni")

> pairs(fm_means, adjust = "bonferroni")							
contrast	estimate	SE	df	t.ratio	p.value		
G16 - G17	-2.0473	0.405	34	-5.050	0.0020		
G16 - G18	-2.0913	0.405	34	-5.159	0.0015		
G16 - G19	-1.3300	0.405	34	-3.281	0.3259		
G16 - G20	-0.7083	0.405	34	-1.747	1.0000		
G16 - G21	-1.1917	0.405	34	-2.940	0.7986		
G16 - G22	-1.5513	0.405	34	-3.827	0.0721		
G16 - G23	-1.9607	0.405	34	-4.837	0.0038		
G16 - G24	-1.6793	0.405	34	-4.143	0.0292		
G16 - G25	-1.8563	0.405	34	-4.579	0.0082		
G16 - G26	-1.0453	0.405	34	-2.579	1.0000		
G16 - G27	-0.9090	0.405	34	-2.242	1.0000		
G16 - G28	-0.7557	0.405	34	-1.864	1.0000		
G16 - G29	-1.6380	0.405	34	-4.041	0.0392		
G16 - G30	-1.5007	0.405	34	-3.702	0.1026		
G16 - G31	-1.6943	0.405	34	-4.180	0.0262		
G16 - G32	-1.5370	0.405	34	-3.791	0.0797		
G17 - G18	-0.0440	0.405	34	-0.109	1.0000		
G17 - G19	0.7173	0.405	34	1.769	1.0000		
G17 - G20	1.3390	0.405	34	3.303	0.3071		
G17 - G21	0.8557	0.405	34	2.111	1.0000		
G17 - G22	0.4960	0.405	34	1,224	1.0000		

Fit the model with random effects

```
library(nlme)
fme <- lme(yield \sim 1, random = list(gen = \sim 1), data = dados)
fme
                          Linear mixed-effects model fit by REML
                            Data: dados
                            Log-restricted-likelihood: -48.51987
                            Fixed: yield ~ 1
                          (Intercept)
                             7.230471
                          Random effects:
                           Formula: ~1 | gen
                                  (Intercept) Residual
                          StdDev: 0.4770553 0.4964977
                          Number of Observations: 51
                          Number of Groups: 17
```

References

- Chapter 7 Analysis of Variance I: The One-Way Classification¹ (for a more classical view)
- Chapter 2 Completely Randomized Designs²

1. Steel, R. G. & Torrie, J. H. Principles and Procedures of Statistics: A Biometrical Approach. 2nd Edition. (1980).

2. Casella, G. Statistical Design. (2008).