

# BMM0413 – Aula 3A: Genética Bacteriana



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<http://www.onehealthbr.com/>

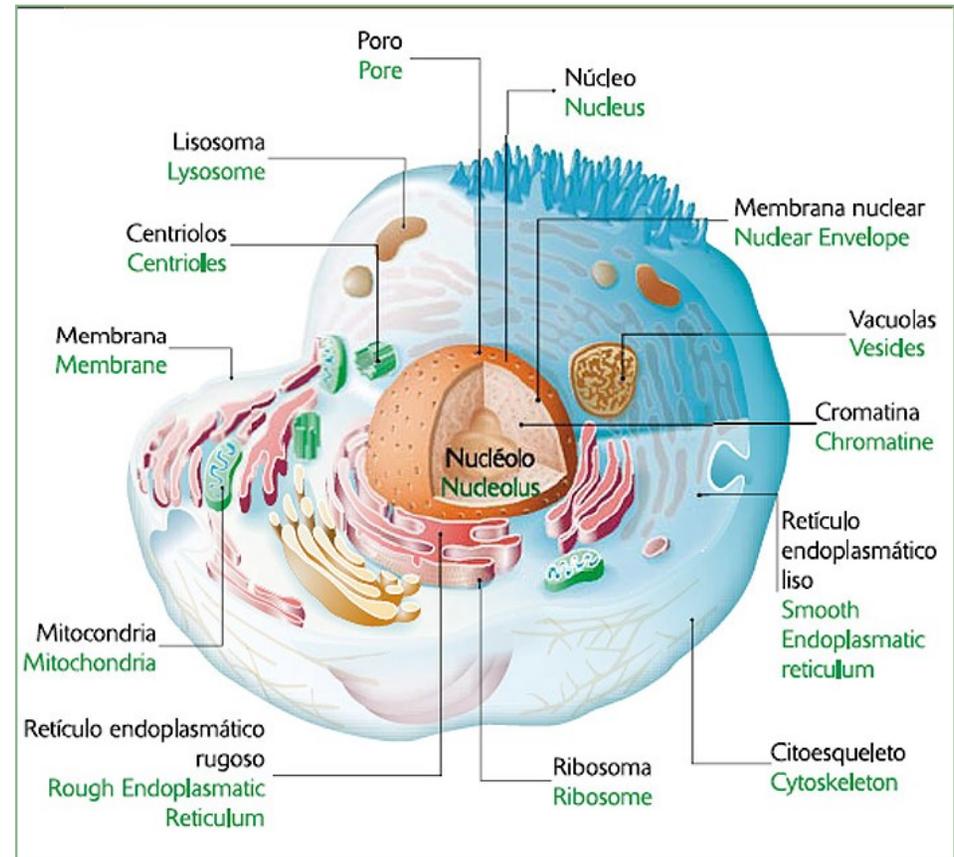
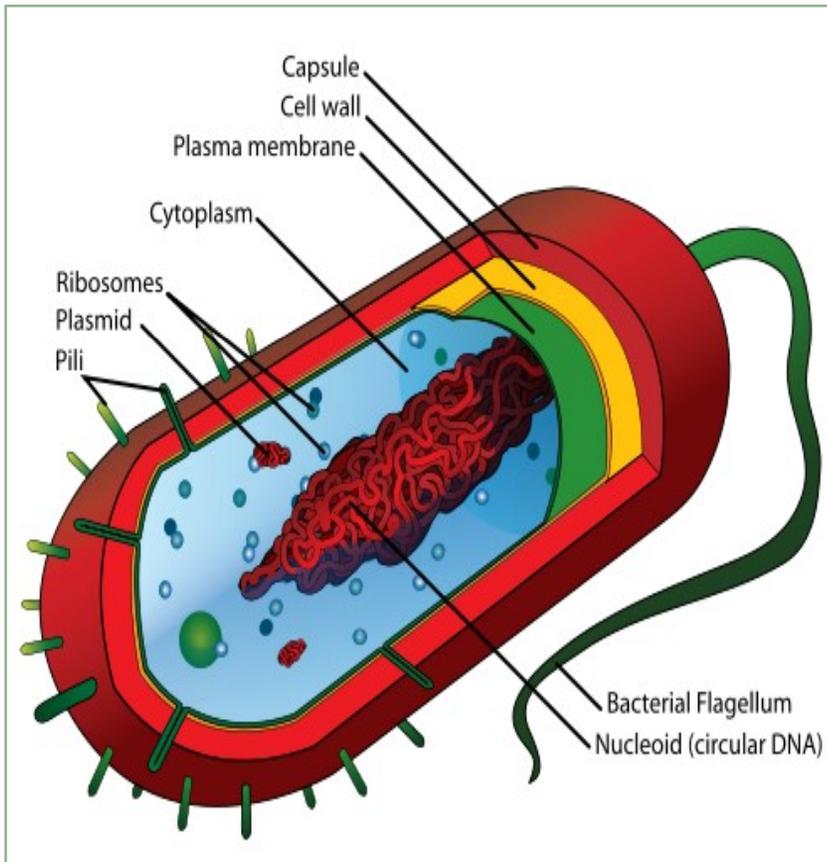


**Departamento de Microbiologia – Instituto de Ciências Biomédicas  
Universidade de São Paulo, Brasil**

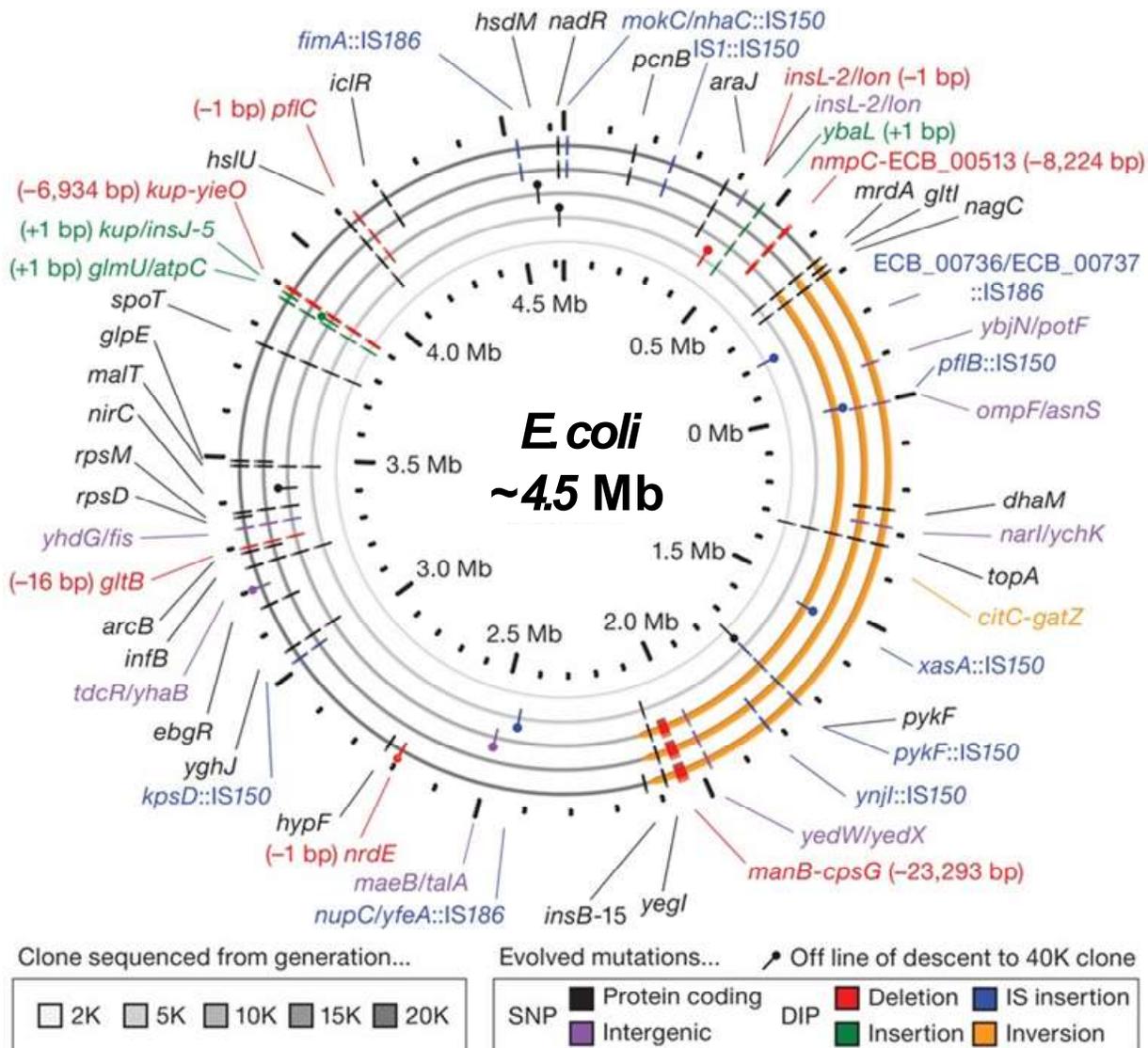
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# Célula Procariota

# Célula Eucariota

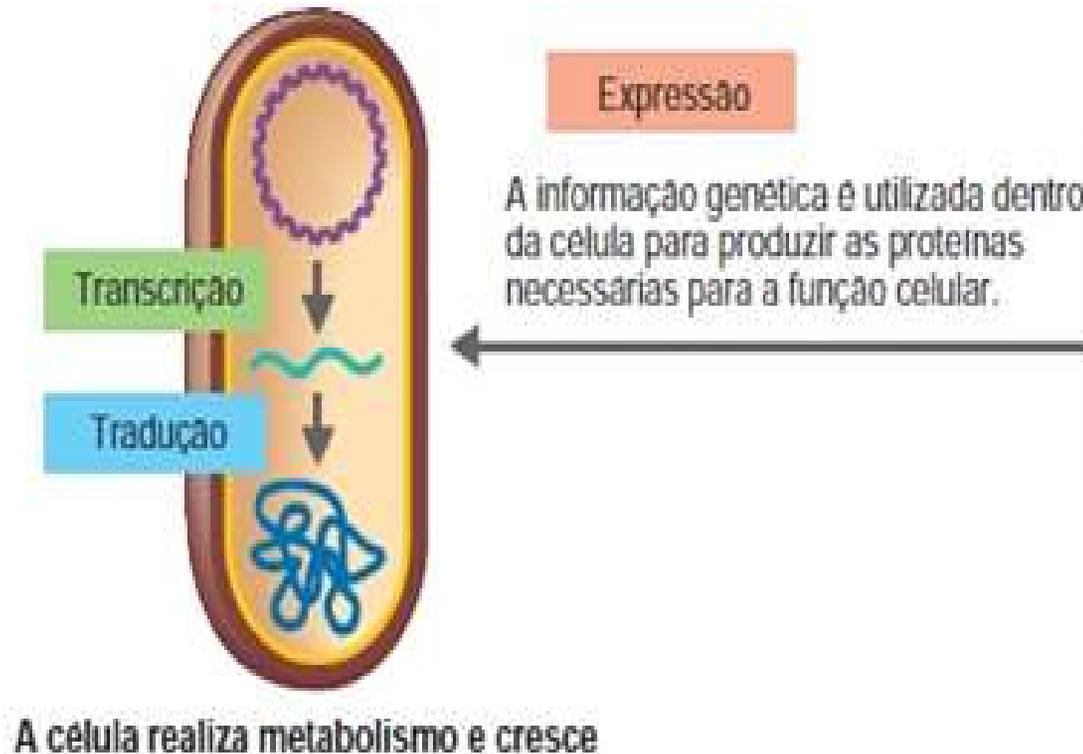


# Cromossomo bacteriano



- ✓ O DNA da *E. coli*, tem cerca de 4,5 milhões de pares de bases e possui cerca de 1 mm de comprimento.
- ✓ Uma célula de *E. coli* mede ~ 1 µm
- ✓ O cromossomo ocupa apenas cerca de 10% do volume celular.
- ✓ DNA é *super enovelado* = *supercoiled*.

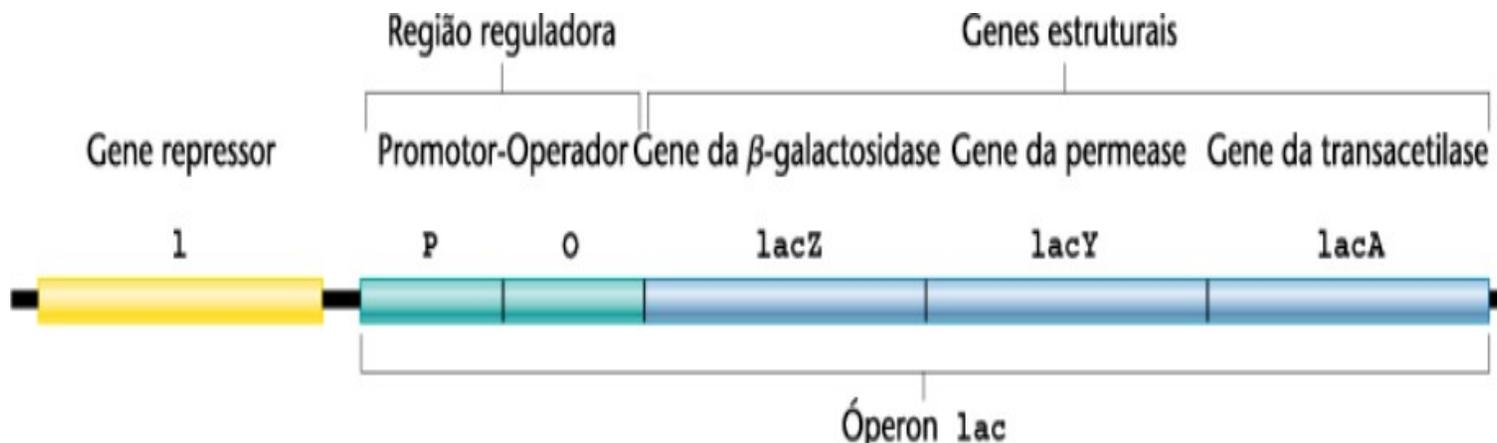
# Cromossomo



- ✓ Existem genes conservados para funções essenciais
- ✓ Genes conservados poder ser utilizados para identificação de espécies
- ✓ Existem genes intrínsecos
- ✓ Existem genes adquiridos

# Elementos genéticos: Operons

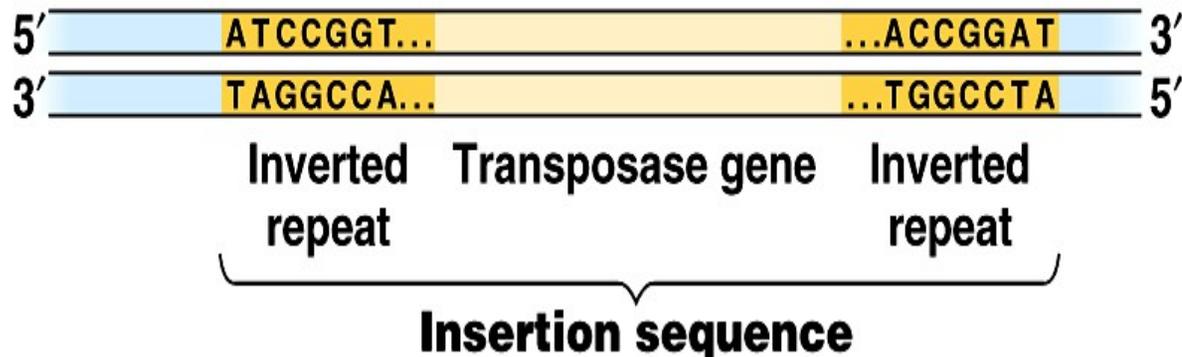
- Agrupamento de genes estruturais localizados no cromossomo bacteriano e que são regulados em conjunto.
- A região reguladora é composta por um promotor.
- Promotores são sequências de nucleotídeos que controlam a expressão (taxa de transcrição) de um gene.



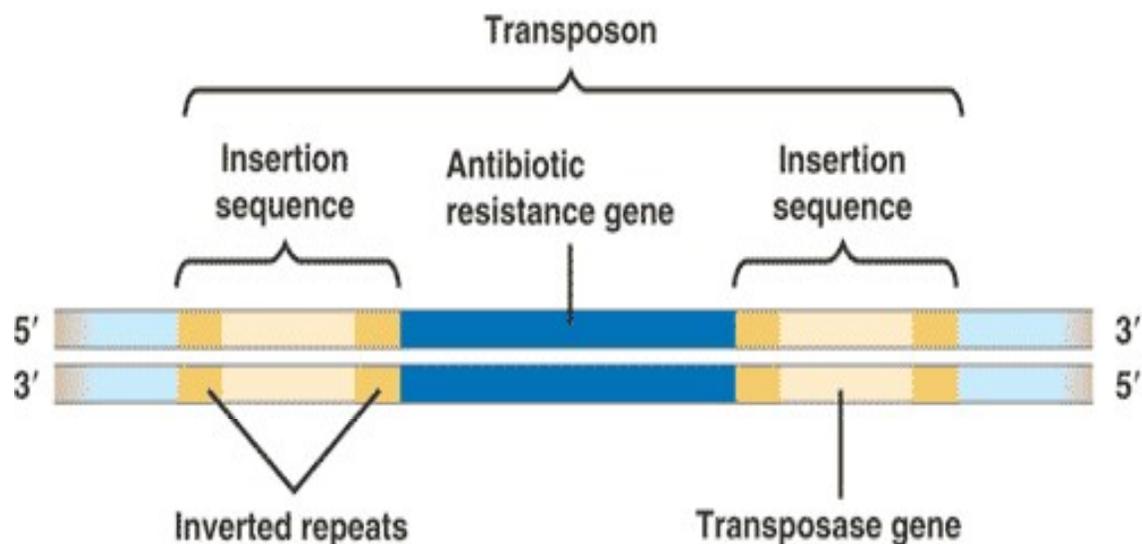
# Elementos genéticos: Sequencias de inserção (IS)

- Elementos simples, com aproximadamente 1000pb;
- Repetições invertidas (10-50pb);
- Único gene: transposase (*tnp*).

DNA



# Elementos genéticos: Transposons

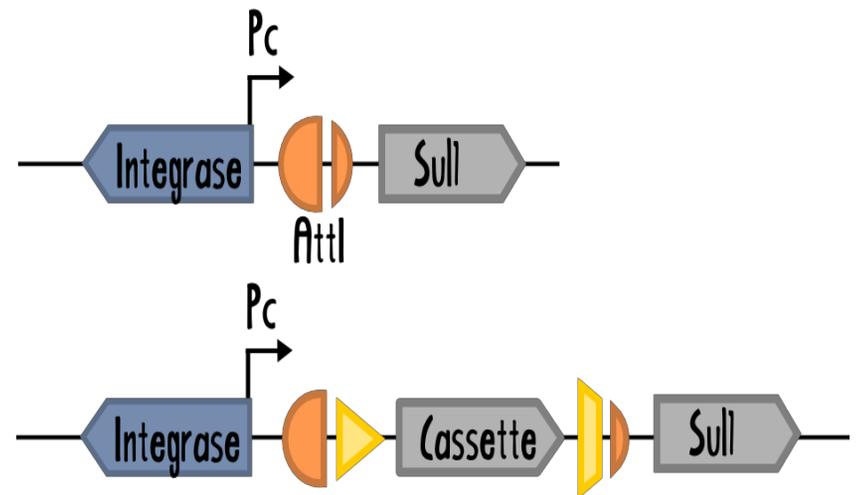


- Estruturas maiores e mais complexas;
- Mesmos componentes essenciais: Repetições invertidas e transposase (*tnp*).
- Codificam genes de resistência, virulência, e enzimas degradativas.

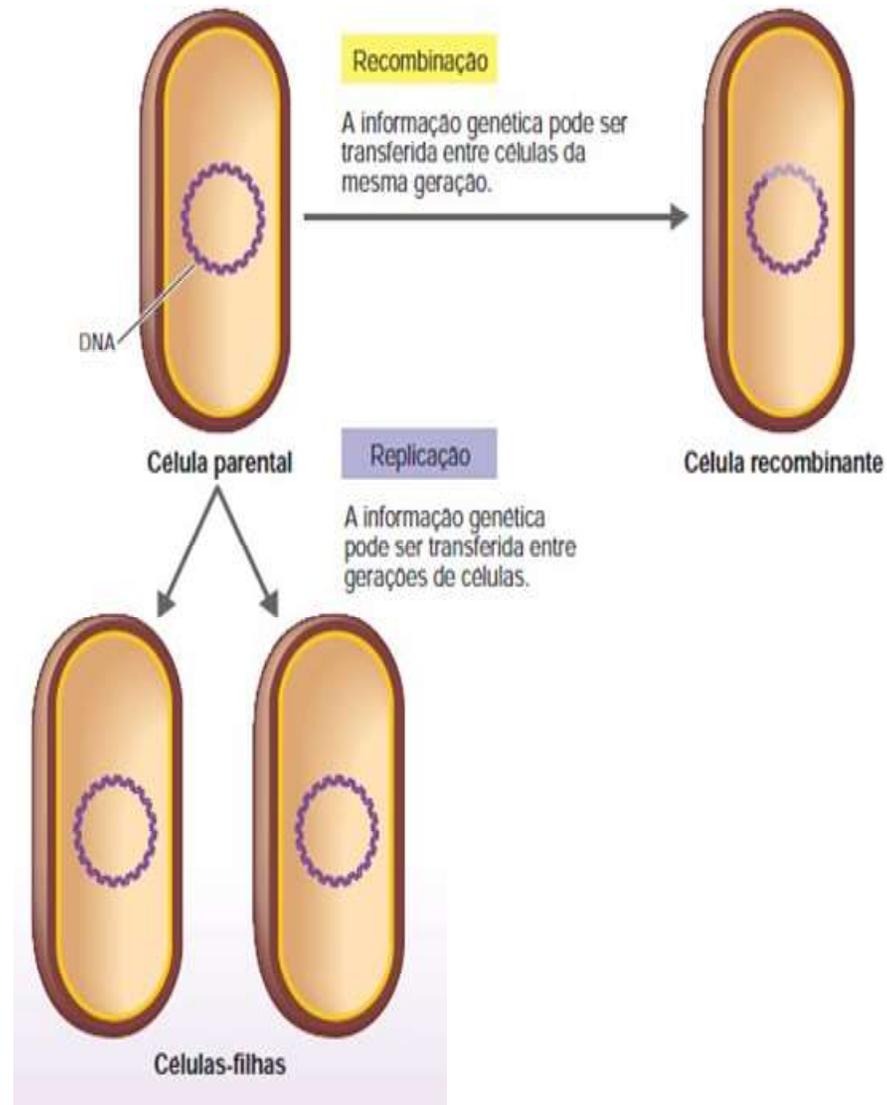
# Elementos genéticos: Integrons

- Sistemas que conseguem capturar genes cassetes.
- Genes cassetes de resistência aos ATM.
- Estrutura:
  - Gene codificador de integrase (intI).
  - Sítio de recombinação (attI).
  - Promotor constitutivo (Pc).

## Class 1 Integrons

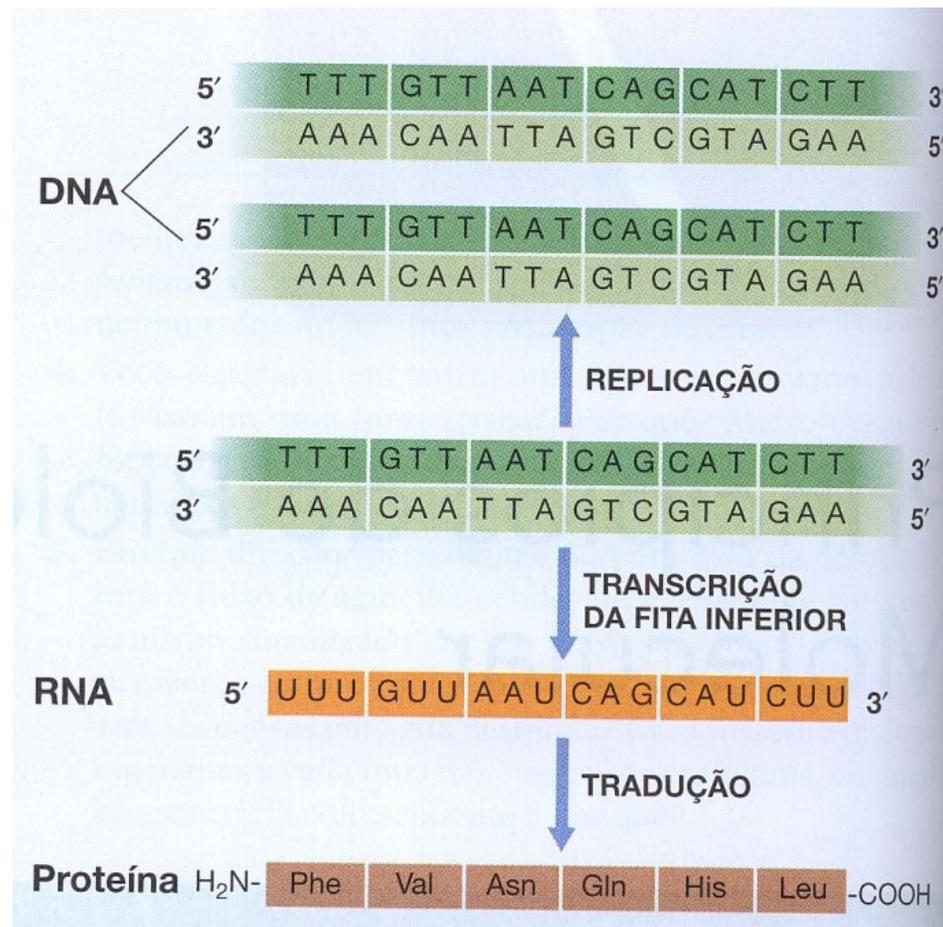


# Fluxo da informação genética

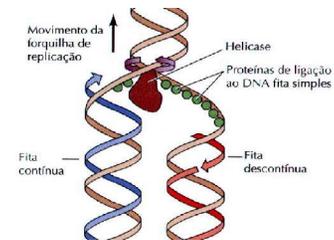


# Síntese dos três tipos de macromoléculas

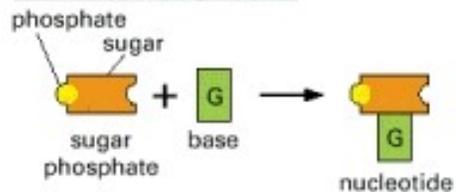
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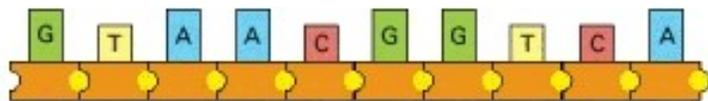
# REPLICAÇÃO



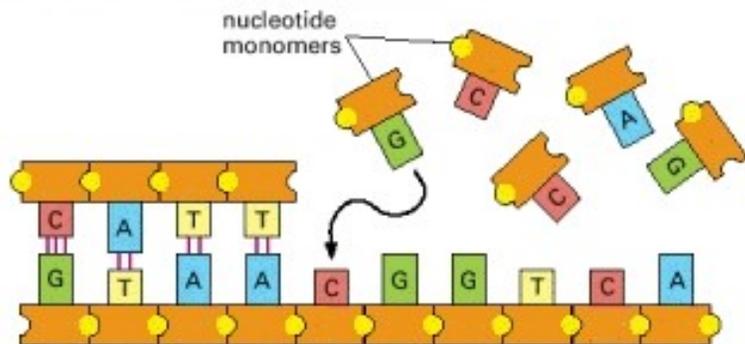
(A) building block of DNA



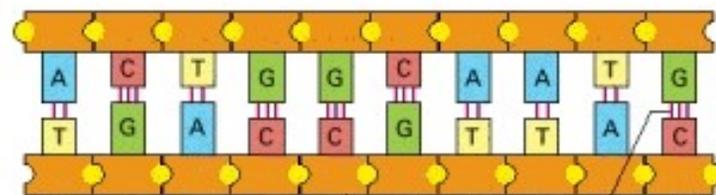
(B) DNA strand



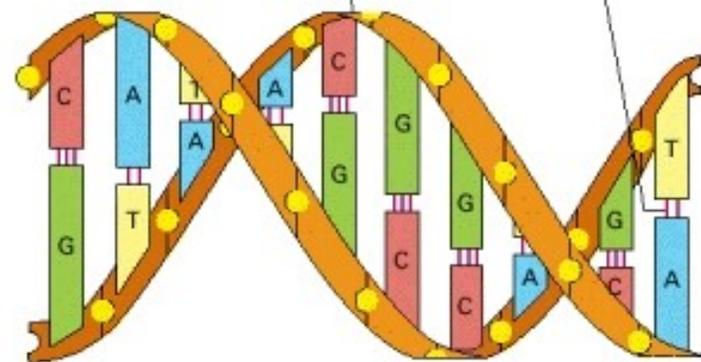
(C) templated polymerization of new strand



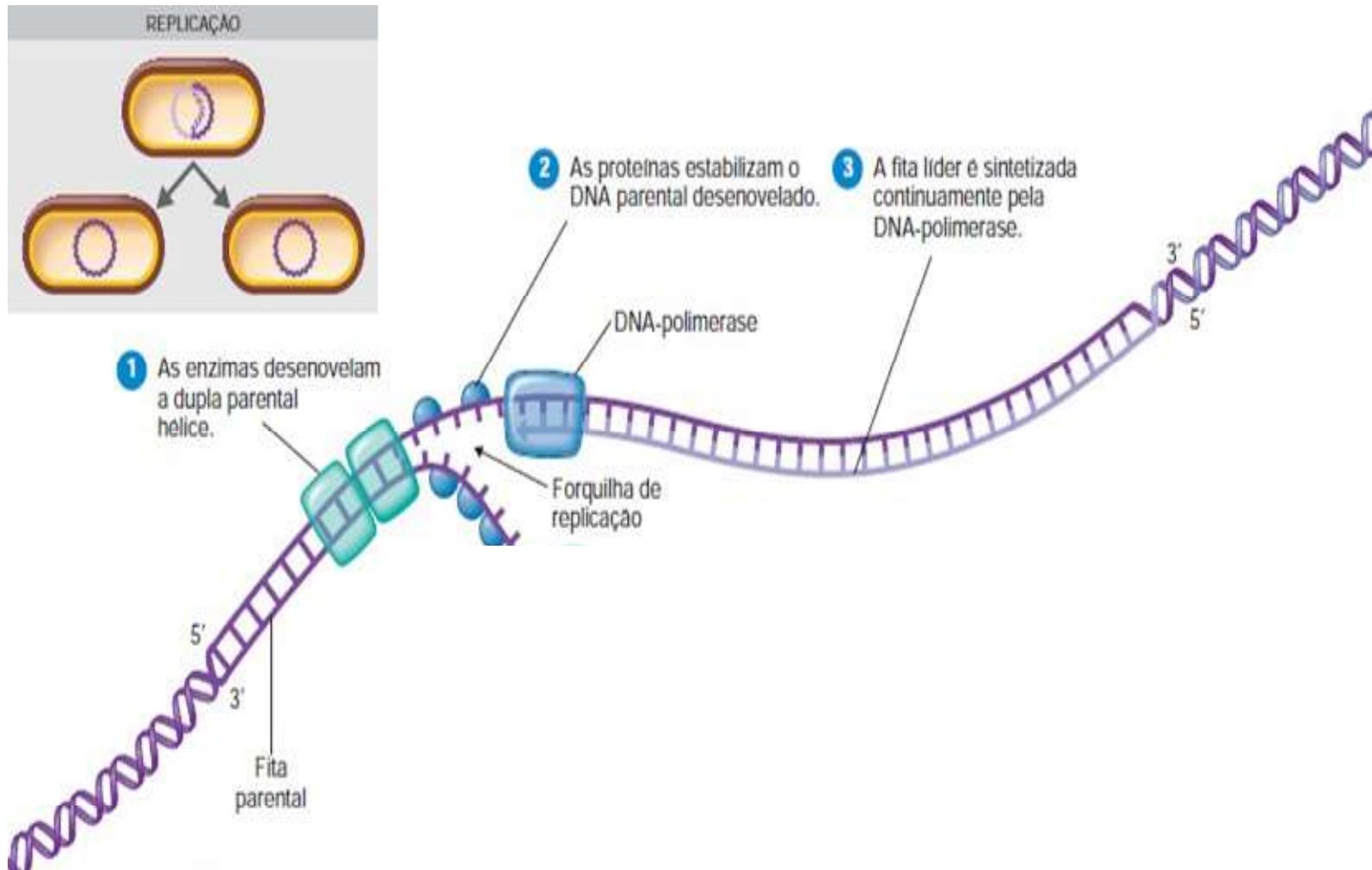
(D) double-stranded DNA



(E) DNA double helix



# REPLICAÇÃO

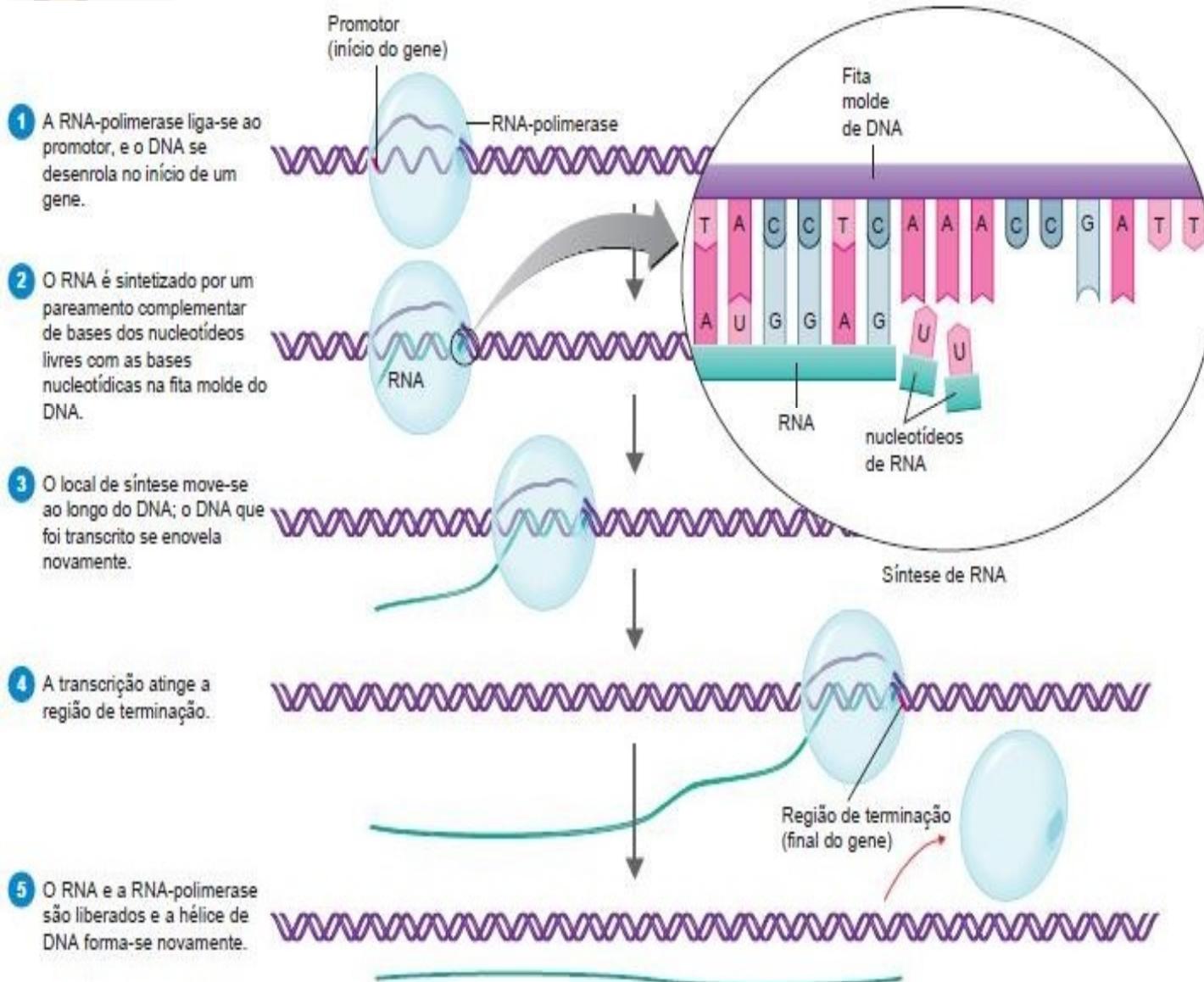


# Elementos da Replicação

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- 2 fitas do DNA genômico
- DNA polimerase
- Primase
- Dinucleotídeos (dNTPs)
- Helicase (Topoisomerase)

# Transcrição



# TRANSCRIÇÃO

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- 1 fita de DNA (fita molde)
- RNA Polimerase
- Helicase
- Primase
- Ribonucleotídeos

Objetivo: Formação de RNA mensageiro (mRNA)

# Tradução

- Conversão para proteínas.
- O mRNA está organizado em forma de **códons**.
  - Grupos de três nucleotídeos, como AUG, GGC ou AAA.
  - A sequência de códons em uma molécula de mRNA determina a sequência de aminoácidos das proteínas.
- Cada códon codifica um aminoácido específico.

		Segunda posição				
		U	C	A	G	
Primeira posição	U	UUU } Phe	UCU } Ser	UAU } Tyr	UGU } Cys	U
		UUC } Phe	UCC } Ser	UAC } Tyr	UGC } Cys	C
		UUA } Leu	UCA } Ser	UAA Stop	UGA Stop	A
		UUG } Leu	UCG } Ser	UAG Stop	UGG Trp	G
	C	CUU } Leu	CCU } Pro	CAU } His	CGU } Arg	U
		CUC } Leu	CCC } Pro	CAC } His	CGC } Arg	C
		CUA } Leu	CCA } Pro	CAA } Gln	CGA } Arg	A
		CUG } Leu	CCG } Pro	CAG } Gln	CGG } Arg	G
	A	AUU } Ile	ACU } Thr	AAU } Asn	AGU } Ser	U
		AUC } Ile	ACC } Thr	AAC } Asn	AGC } Ser	C
		AUA } Ile	ACA } Thr	AAA } Lys	AGA } Arg	A
		AUG Met/start	ACG } Thr	AAG } Lys	AGG } Arg	G
	G	GUU } Val	GCU } Ala	GAU } Asp	GGU } Gly	U
		GUC } Val	GCC } Ala	GAC } Asp	GGC } Gly	C
		GUA } Val	GCA } Ala	GAA } Glu	GGA } Gly	A
		GUG } Val	GCG } Ala	GAG } Glu	GGG } Gly	G

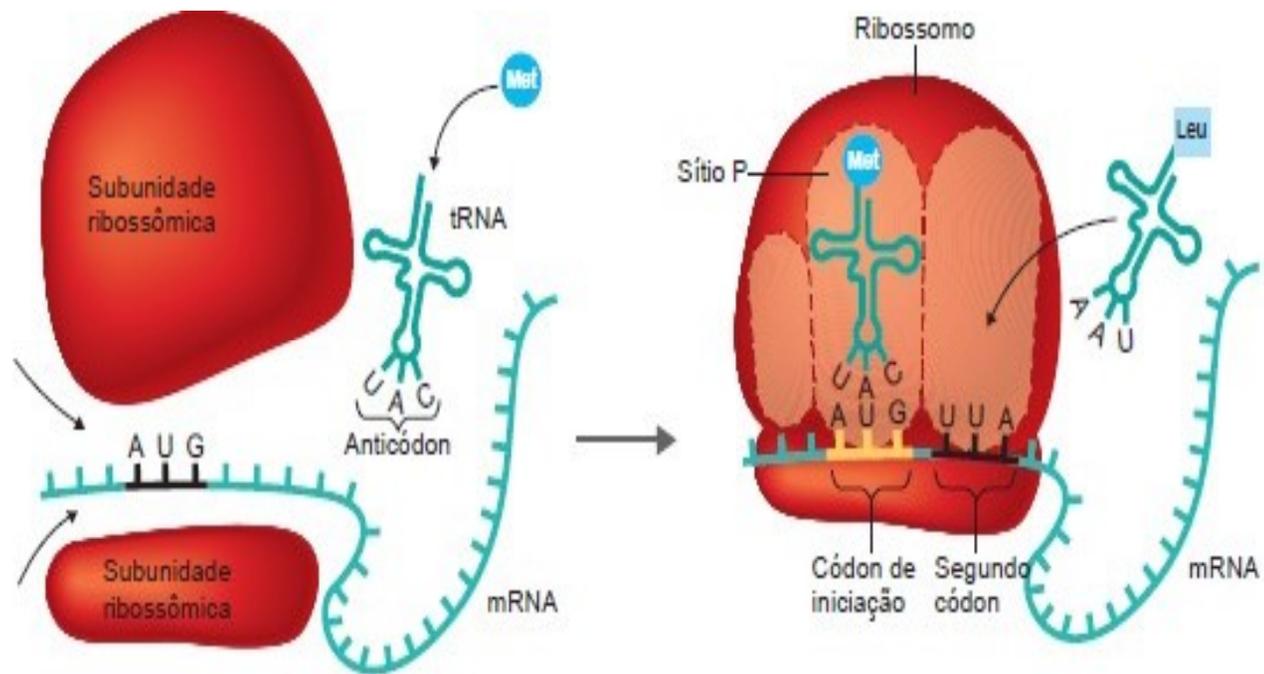
# Código genético

	AGA									UUA					AGC					
	AGG									UUG					AGU					
GCA	CGA						GGA			CUA				CCA	UCA	ACA			GUA	
GCC	CGC						GGC		AUA	CUC				CCC	UCC	ACC			GUC	UAA
GCG	CGG	GAC	AAC	UGC	GAA	CAA	GGG	CAC	AUC	CUG	AAA		UUC	CCG	UCG	ACG		UAC	GUG	UAG
GCU	CGU	GAU	AAU	UGU	GAG	CAG	GGU	CAU	AUU	CUU	AAG	AUG	UUU	CCU	UCU	ACU	UGG	UAU	GUU	UGA
Ala	Arg	Asp	Asn	Cys	Glu	Gln	Gly	His	Ile	Leu	Lys	Met	Phe	Pro	Ser	Thr	Trp	Tyr	Val	stop
A	R	D	N	C	E	Q	G	H	I	L	K	M	F	P	S	T	W	Y	V	



Iniciação

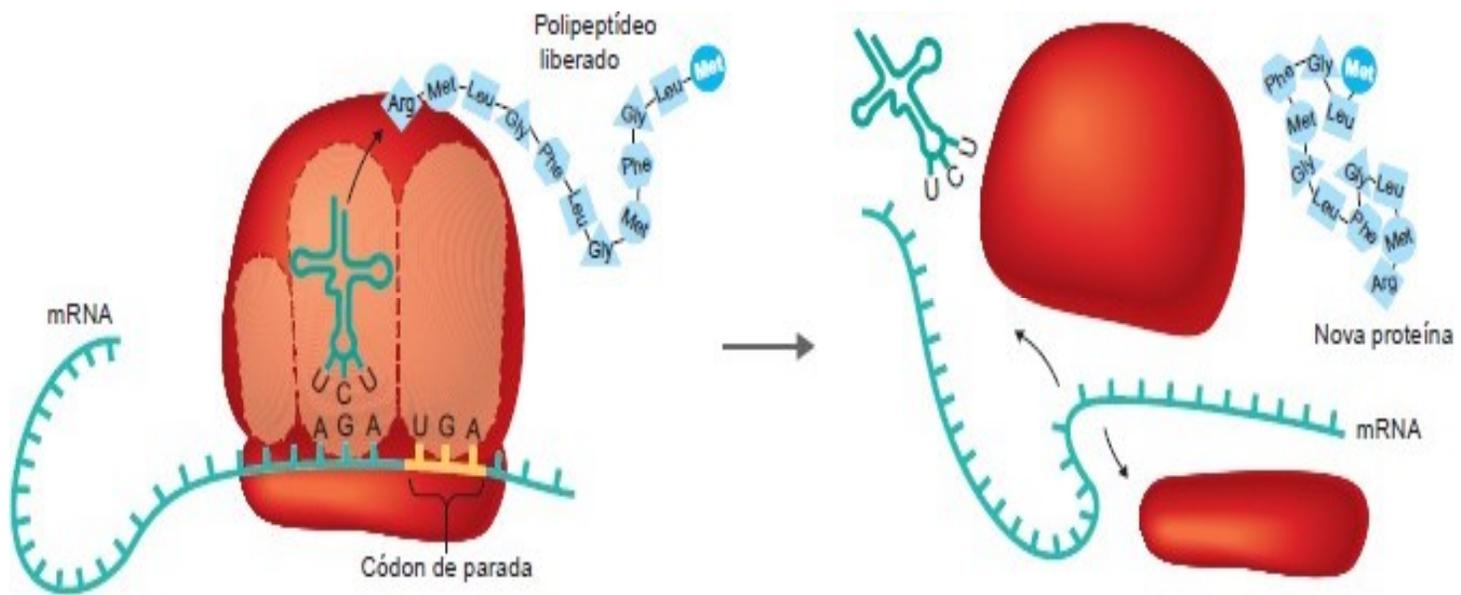
# Tradução



**1** Os componentes necessários para iniciar a tradução são reunidos.

**2** Na montagem do ribossomo, um tRNA carregando o primeiro aminoácido é pareado com o códon de iniciação no mRNA. O lugar onde o primeiro tRNA se estabelece é denominado sítio P. Um tRNA transportando o segundo aminoácido se aproxima.

# Tradução



**7** Quando o ribossomo alcança um códon de parada, o polipeptídeo é liberado.

**8** Finalmente, o último tRNA é liberado e o ribossomo começa a se desligar. O polipeptídeo liberado forma uma nova proteína.

# TRADUÇÃO: SÍNTESE DE PROTEÍNAS

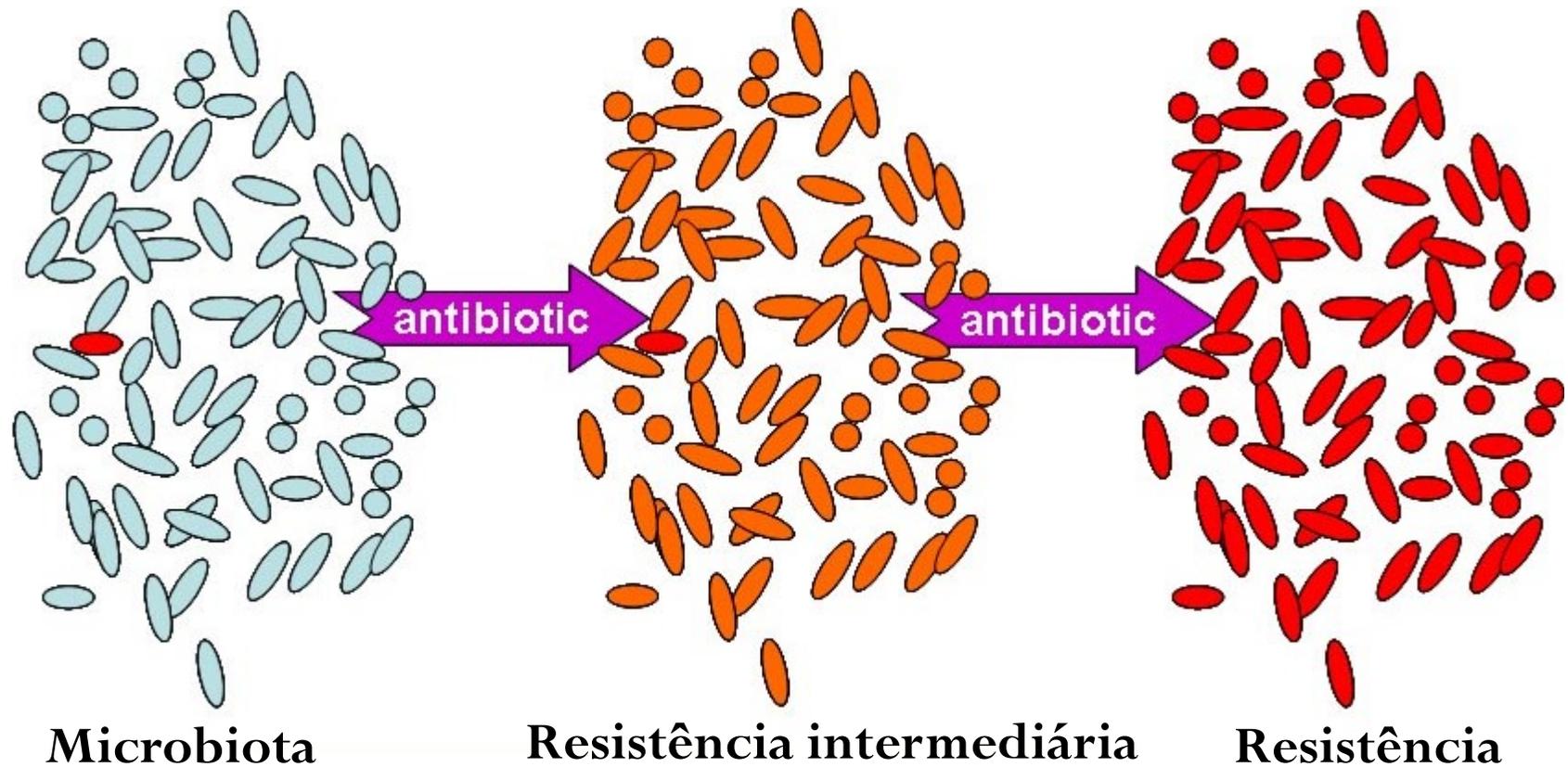
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- A partir do mRNA
- **Leitura dos códons** do mRNA para a tradução em aminoácidos
- Uso de tRNA (anti-códon), aminoacil-tRNA sintetase, Ribossomo.
- Fases: Iniciação, alongação, translocação.
- Formação de **ligação peptídica**.

# Mutações

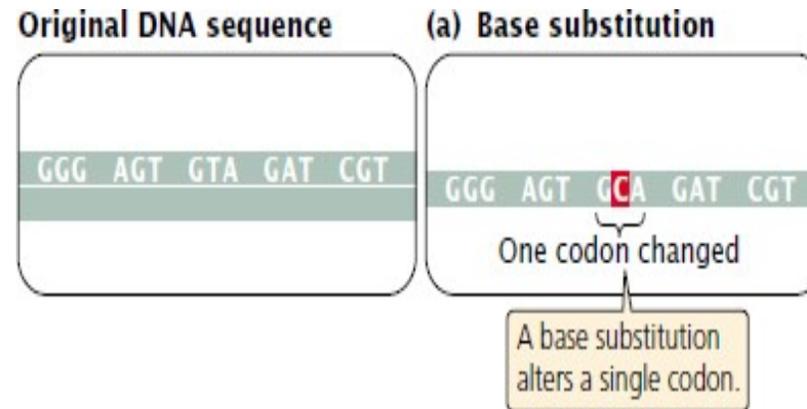
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## Pressão seletiva *in vitro*

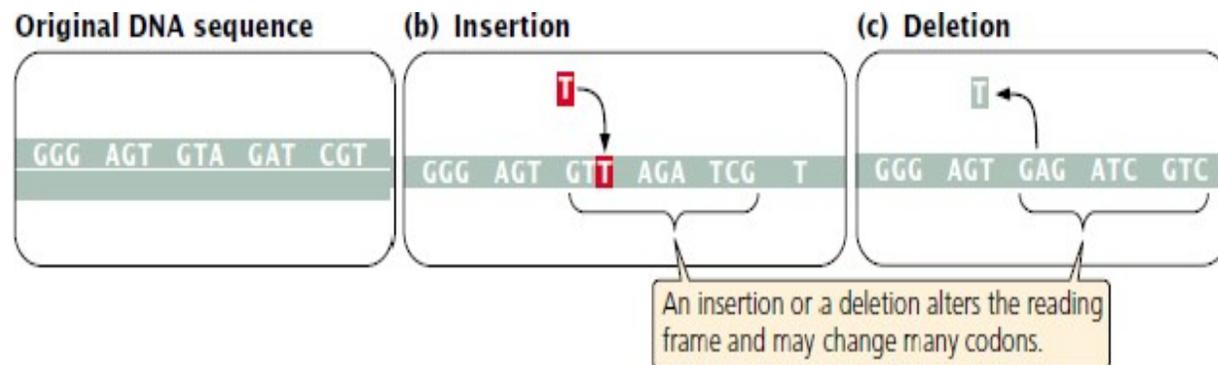


# Mutações

- Substituição:

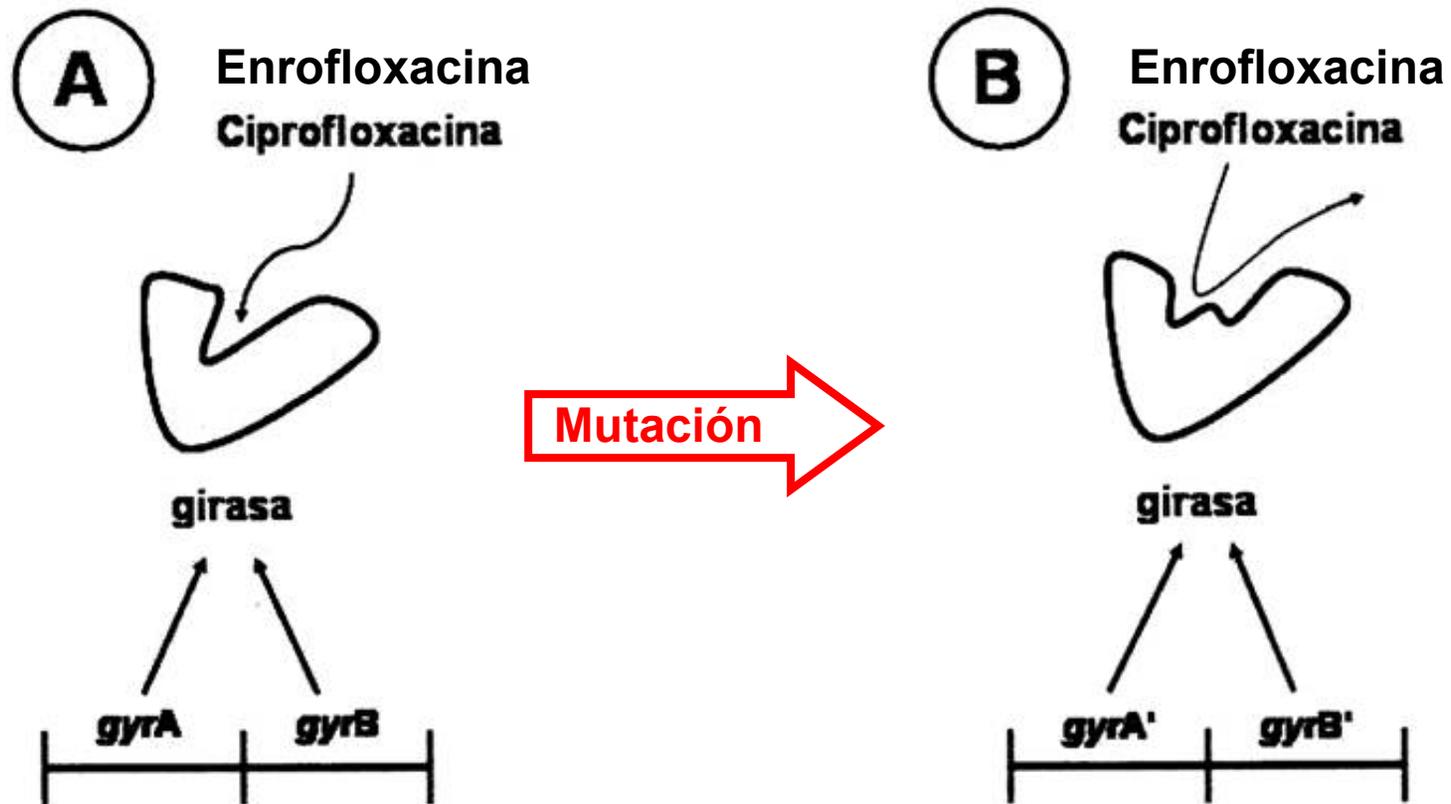


- Inserções e deleções :



# Resistência às fluoroquinolonas

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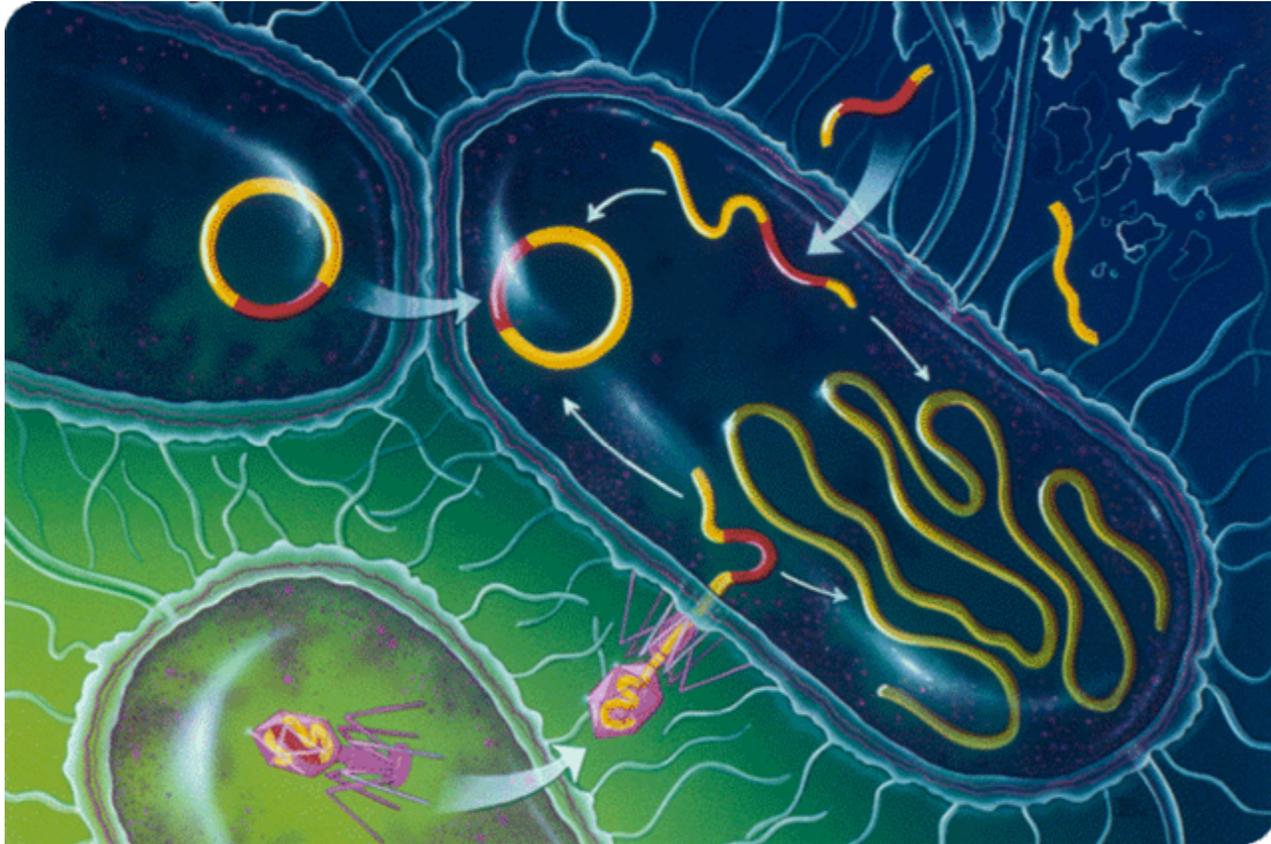


**Bactéria Suscetível**

**Bactéria Resistente**

# Mobilização genética

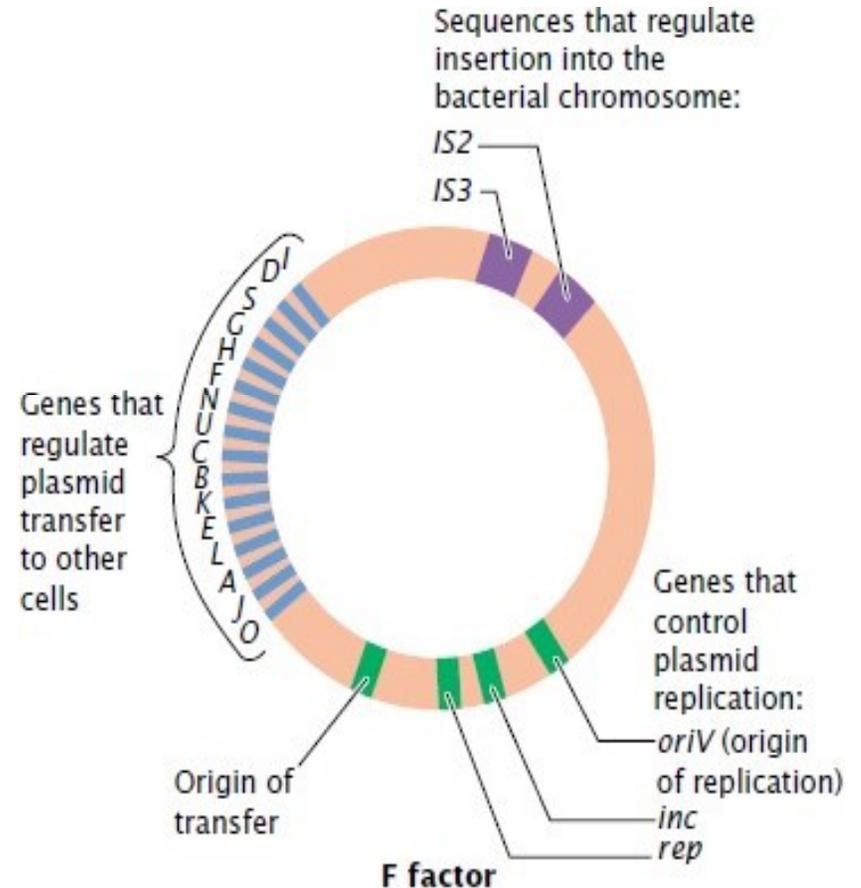
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1. Conjugação
2. Transformação
3. Transdução

# Conjugação

- Fator F
  - Plasmídeos conjugativos.
  - Origem de replicação.
  - Genes para conjugação.
    - Pili sexual: finas extensões na membrana celular pela qual ocorre a transferência do material genético.



# Food Animals and Antimicrobials: Impacts on Human Health

Bonnie M. Marshall<sup>1,2</sup> and Stuart B. Levy<sup>1,2,3\*</sup>

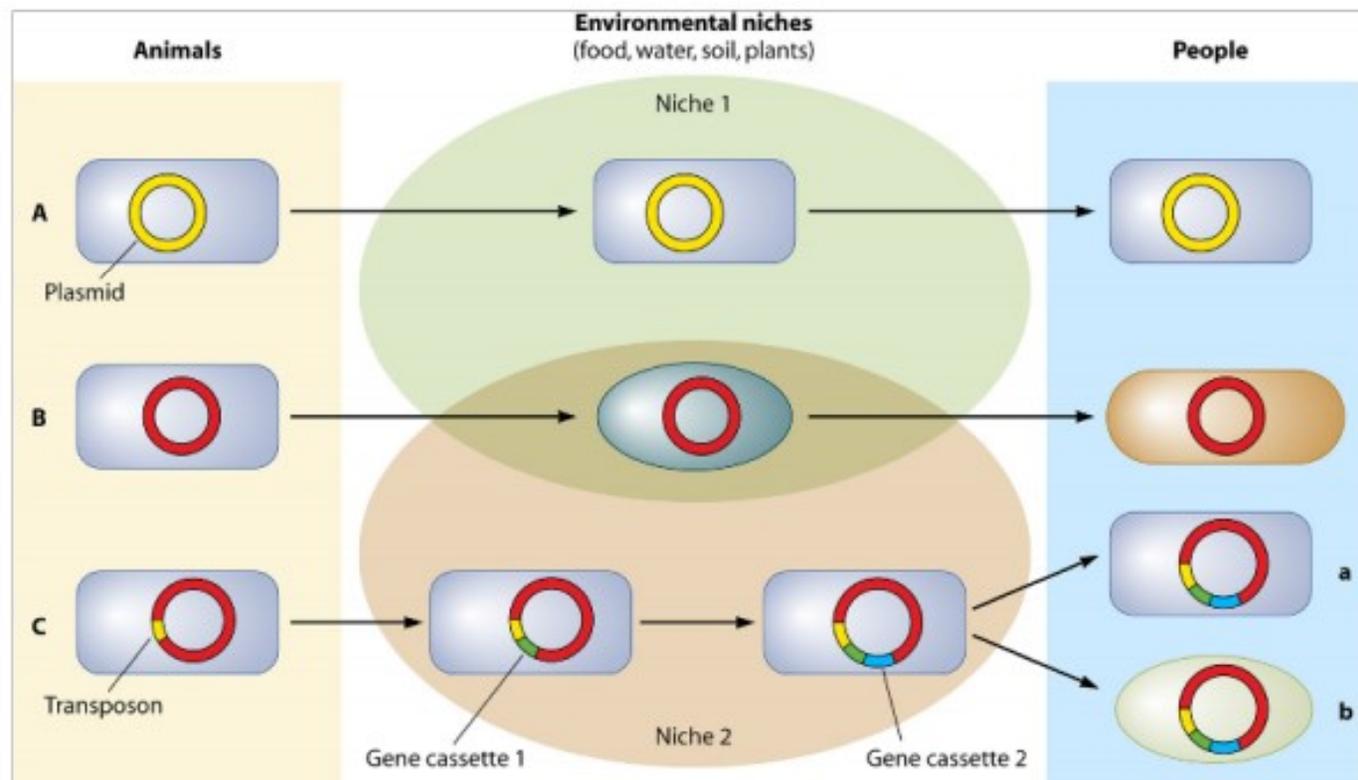
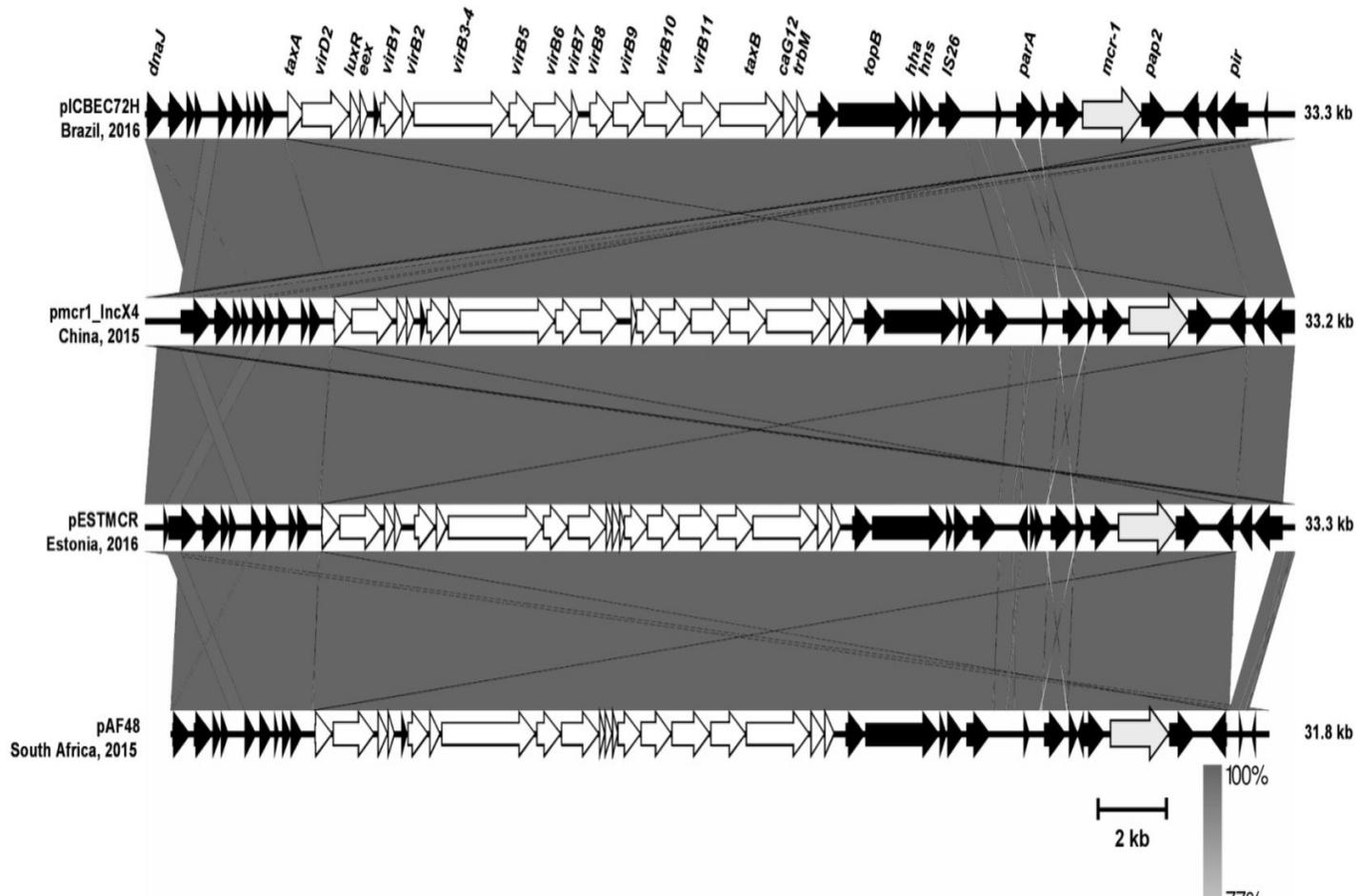


FIG. 1. Several scenarios may present themselves in the genetic transport that occurs as bacteria migrate from animal to human environments. (A) The same host and its indigenous genes in animals are transported unchanged to humans, with a resulting 100% match of the bacterial strain. (B) The genetic structure passes through one or more different hosts, ending in a new host (humans), with a resulting 100% match of DNA. (C) The host and its plasmid-borne genes pass through the environment, picking up gene cassettes en route, with a resulting 100% match for the host only (a) or a low-% match for DNA only (b). In both examples, the plasmid core remains the same.

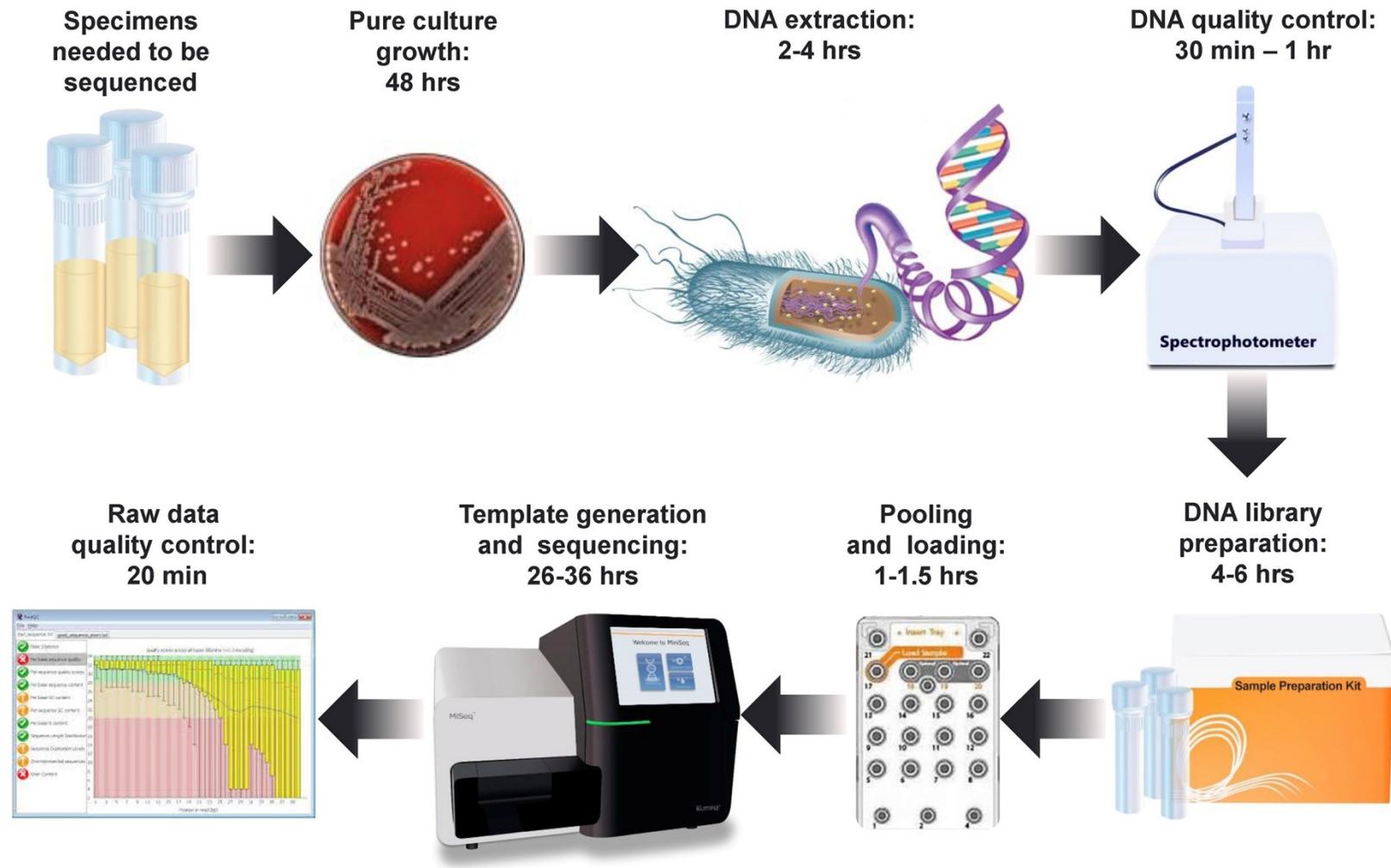
# Plásmidos pandémicos



**IncX4 (*mcr-1*)**  
**IncI1 (*bla*<sub>CTX-M</sub>)**  
**IncA/C (MDR)**  
**IncN (*bla*<sub>KPC</sub>)**

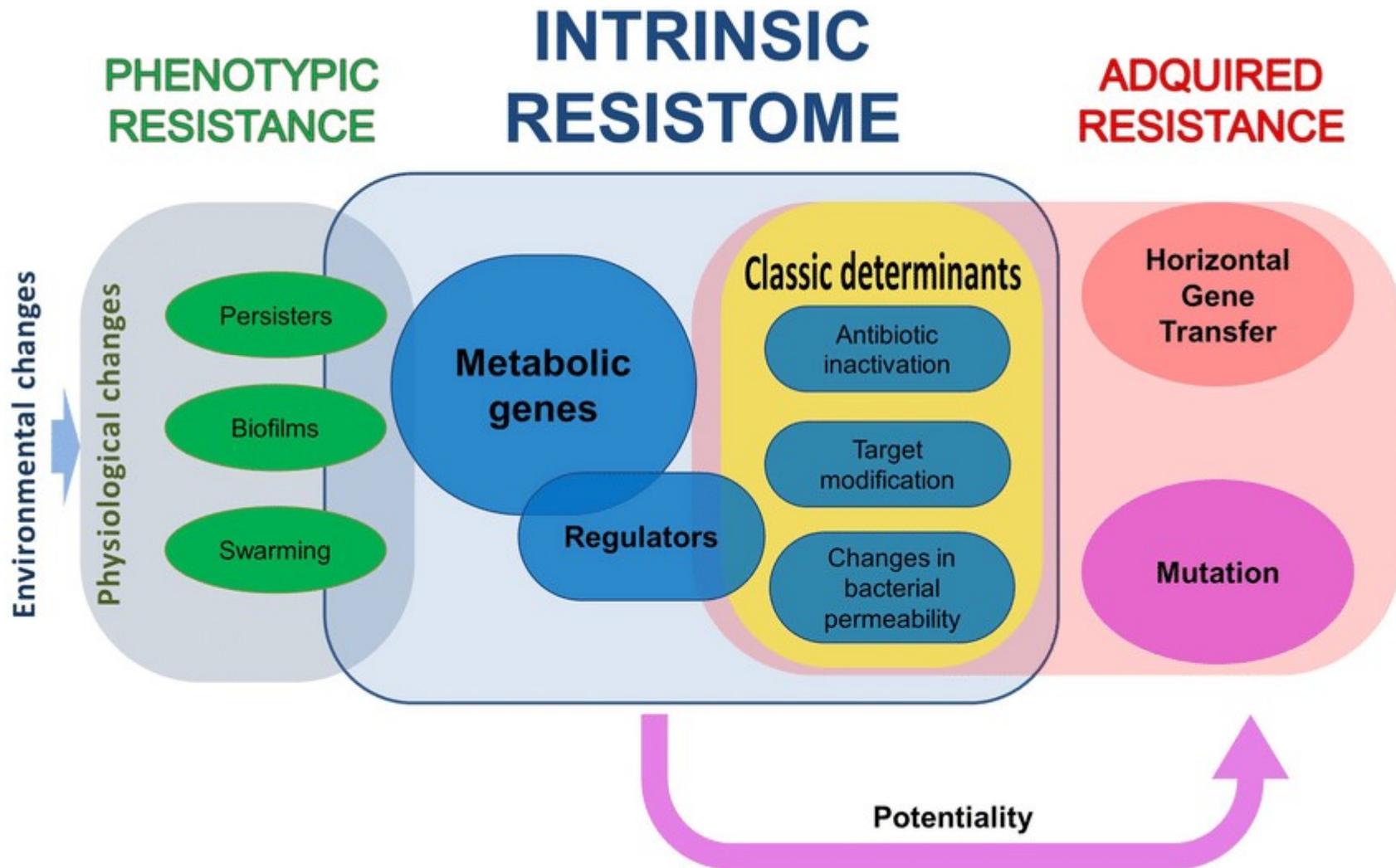
***E. coli***  
***Salmonella* spp.**  
***K. pneumoniae***

# Vigilancia genómica/epidemiológica



Besser J., et al. *Clinical Microbiology and Infection* 2018, 24:335-341.

# A Origem da RAM



# Center for Genomic Epidemiology

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Organization

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Services

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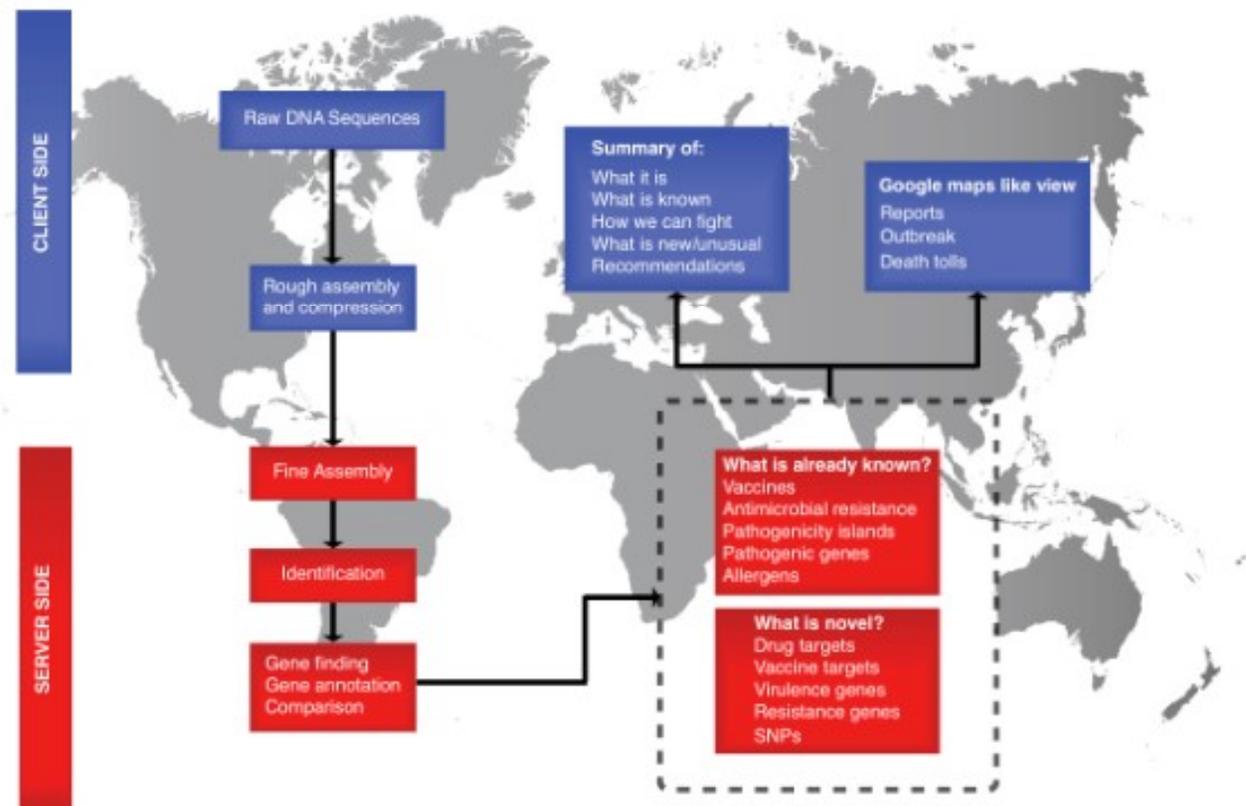
## Services

### Pipeline:

- Identification of acquired antibiotic resistance genes. [Bacterial Analysis Pipeline \(BAP\)](#).

### Phenotyping:

- Identification of acquired antibiotic resistance genes. [ResFinder](#)
- Identification of functional metagenomic antibiotic resistance determinants. [ResFinderFG](#)
- Identification of acquired antibiotic resistance genes using Kmers. [KmerResistance](#)
- Prediction of a bacteria's pathogenicity towards human hosts. [PathogenFinder](#)
- Identification of acquired virulence genes. [VirulenceFinder](#)



<http://www.genomicepidemiology.org/>

# ResFinder 4.1

ResFinder identifies acquired genes and/or finds chromosomal mutations mediating antimicrobial resistance in total or partial DNA sequence of bacteria.

## Updates

ResFinder and PointFinder software: ([2021-05-27](#))

ResFinder database: ([2021-08-04](#))

PointFinder database: ([2021-02-01](#))

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Chromosomal point mutations

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Acquired antimicrobial resistance genes

### Select Antimicrobial configuration

Select multiple items, with Ctrl-Click (or Cmd-Click on Mac) - as default all databases is selected

Aminoglycoside	▲
Beta-lactam	
Colistin	
Disinfectant	
Fluoroquinolone	
Fosfomycin	▼

### Select threshold for %ID

90 %	▼
------	---

### Select minimum length

60 %	▼
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# Multilocus Sequence Typing *E. coli*

*Escherichia coli* MLST Database.

<i>adk</i>	<i>fumC</i>	<i>gyrB</i>	<i>icd</i>	<i>mdh</i>	<i>purA</i>	<i>recA</i>
<input type="text"/>						

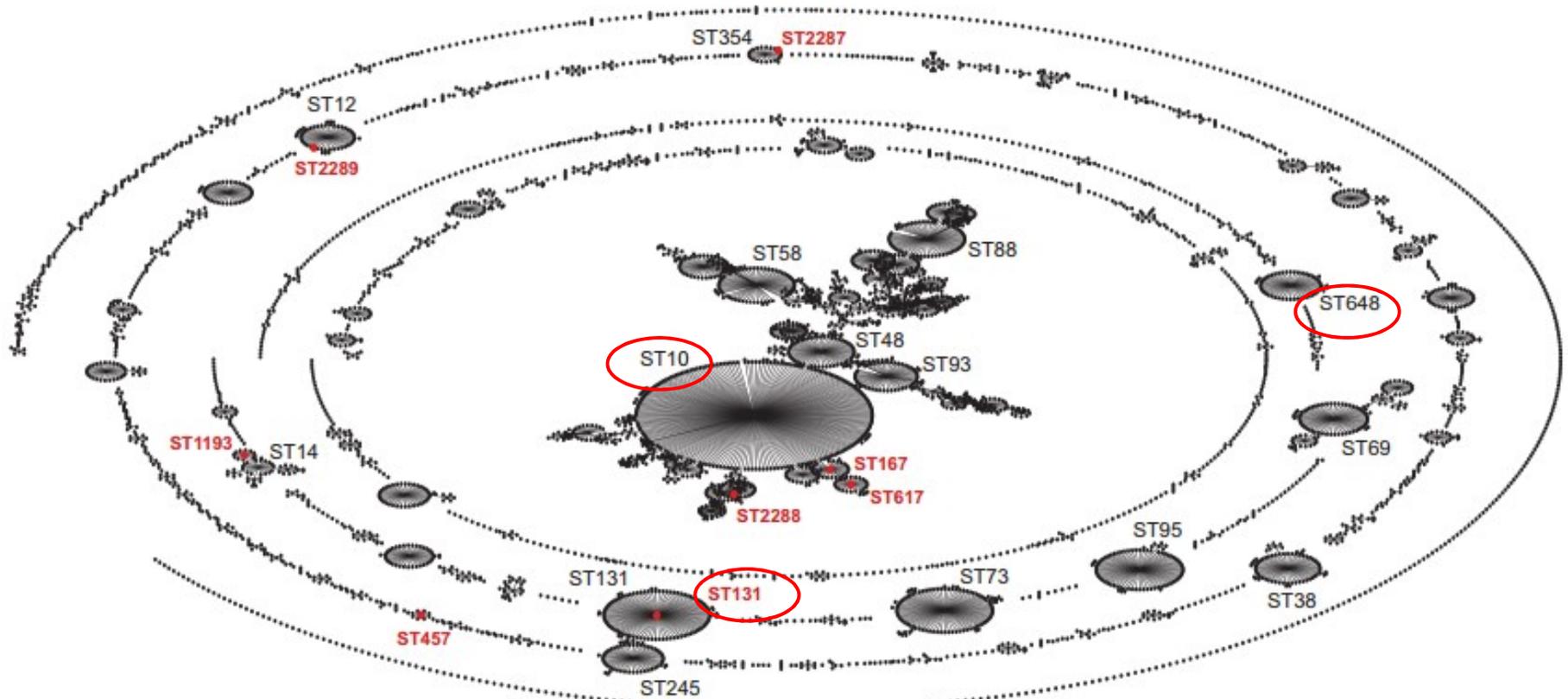
Sequence:

*adk* (adenylate kinase)  
*fumC* (fumarate hydratase)  
*gyrB* (DNA gyrase)  
*icd* (isocitrate/isopropylmalate  
dehydrogenase)  
*mdh* (malate dehydrogenase)  
*purA* (adenylosuccinate dehydrogenase)  
*recA* (ATP/GTP binding motif)

**MLST permite  
identificar  
clones  
designados  
como  
secuencias  
tipo (ST)**

# Multilocus Sequence Typing (MLST)

## *E. coli* Complexos Clonales (CC)



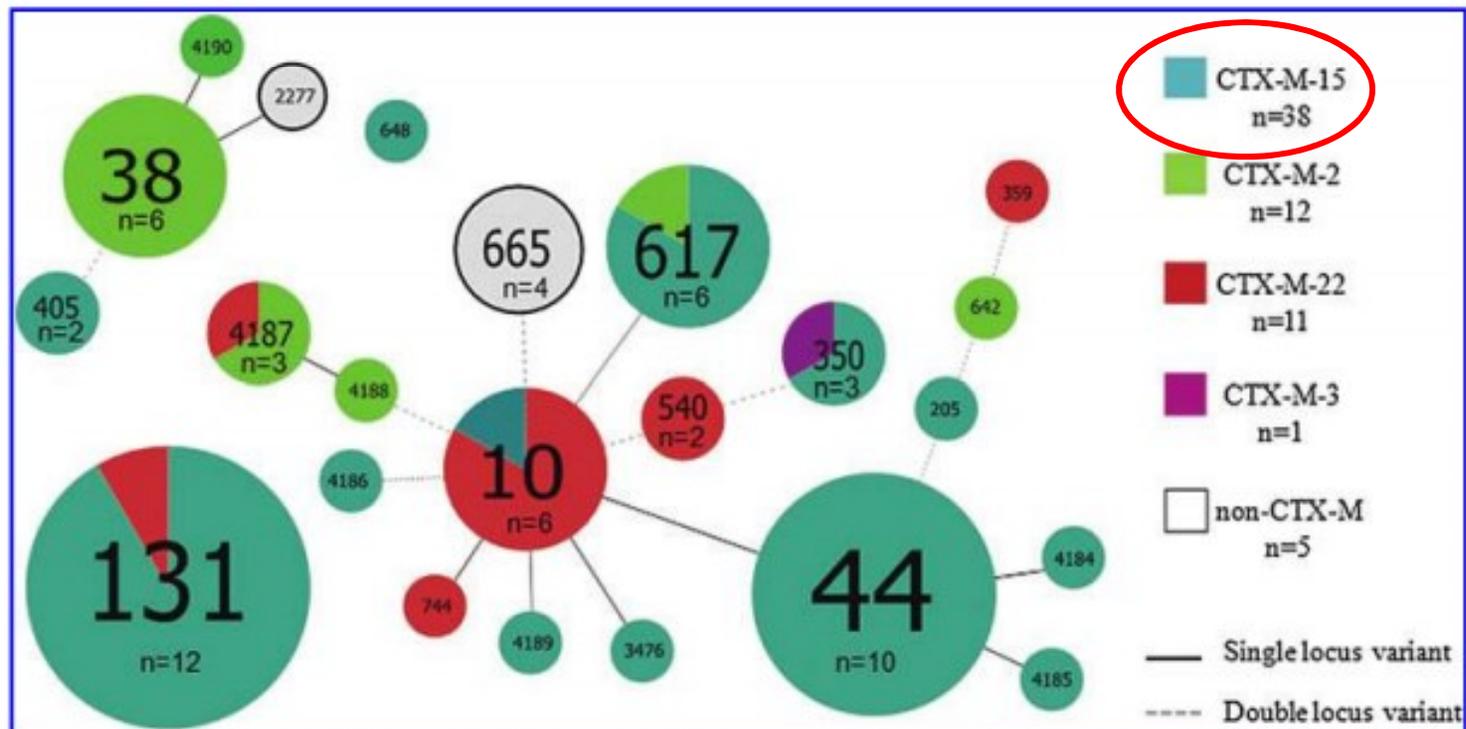
**Pandémicos: ST131, ST10, ST648, ST224, ST410, ST90, ST155**



## Molecular Characterization and Genetic Diversity of ESBL-Producing *Escherichia coli* Colonizing the Migratory Franklin's Gulls (*Leucophaeus pipixcan*) in Antofagasta, North of Chile

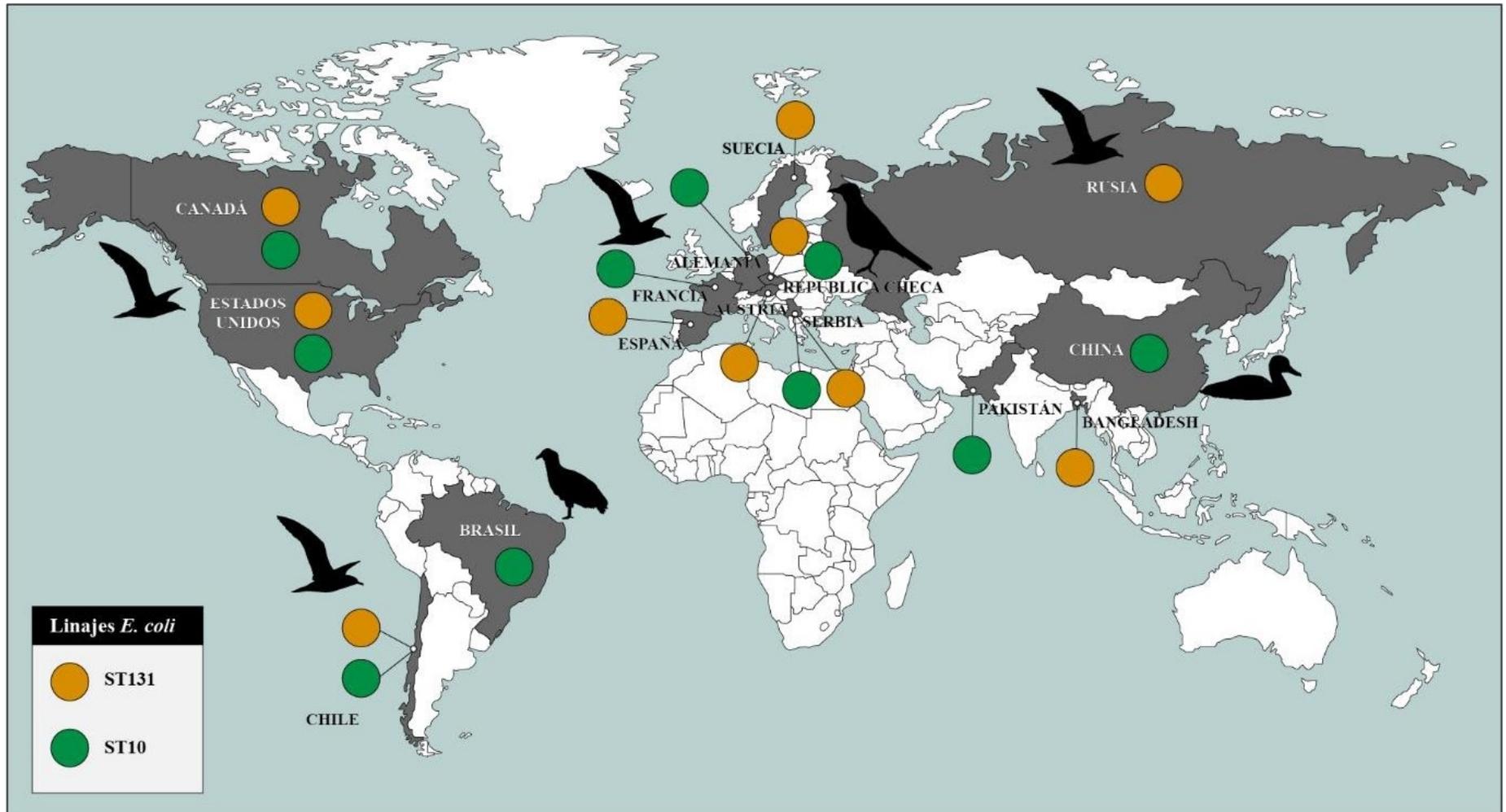
### US/Canada para Chile?

John Báez,<sup>1,\*</sup> Marta Hernández-García,<sup>2,\*</sup> Constanza Guamparito,<sup>1</sup> Sofía Díaz,<sup>1</sup> Abdon Olave,<sup>1</sup> Katherine Guerrero,<sup>1</sup> Rafael Cantón,<sup>2,3</sup> Fernando Baquero,<sup>3,4</sup> Joselyne Gahona,<sup>1</sup> Nicomedes Valenzuela,<sup>1</sup> Rosa del Campo,<sup>2,3</sup> and Juan Silva<sup>1</sup>



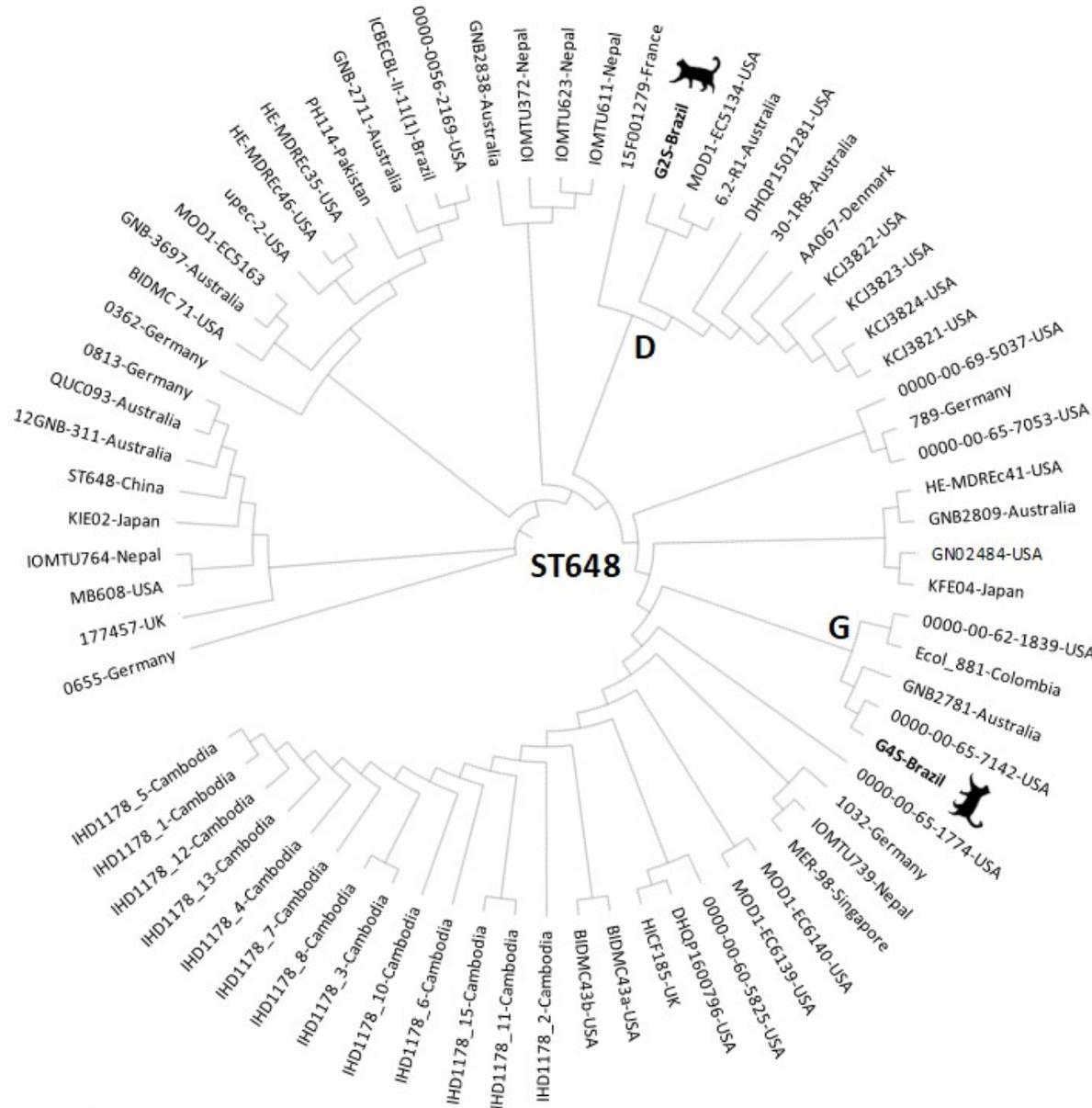
# Clones internacionales em aves migratórias

## *E. coli* ST131 y ST10





# Infection, Genetics and Evolution



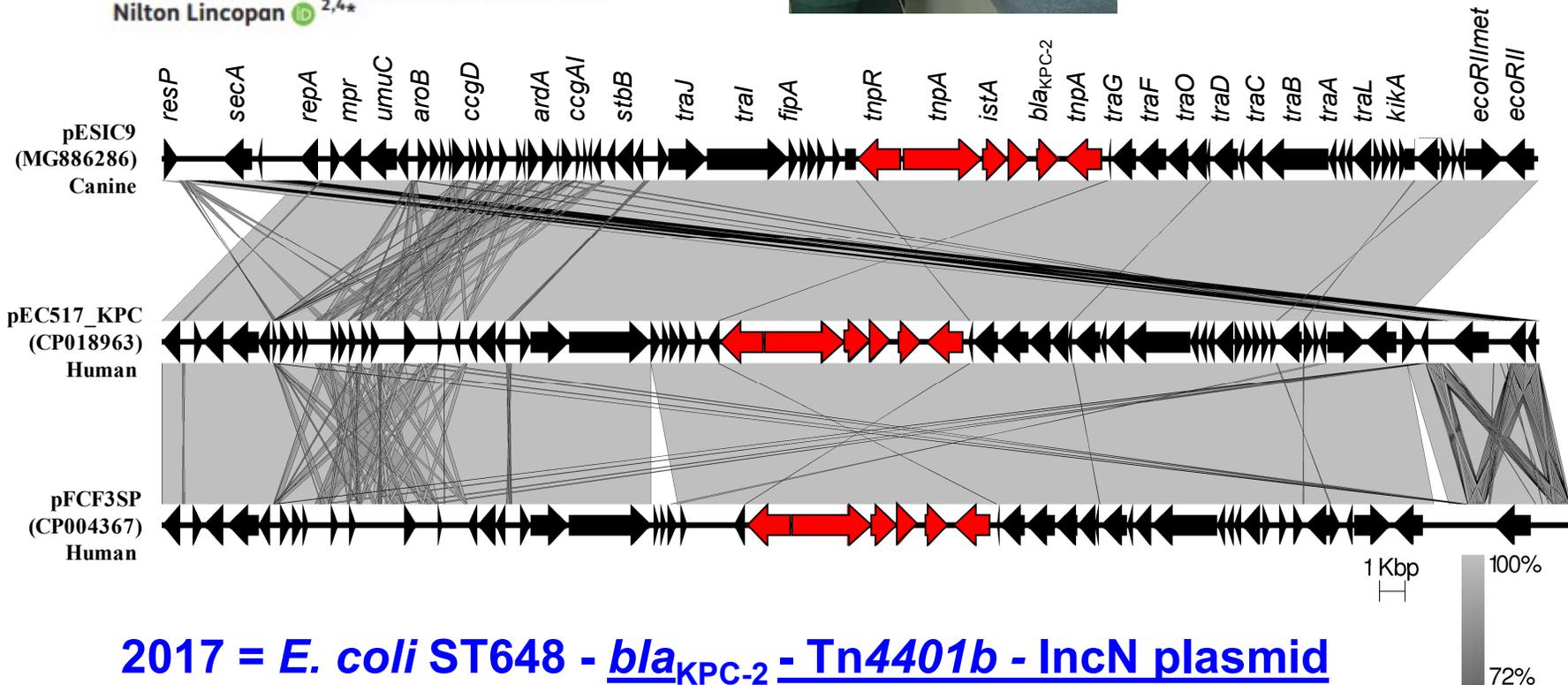
lton



J Antimicrob Chemother  
doi:10.1093/jac/dky173

**Identification of KPC-2-producing *Escherichia coli* in a companion animal: a new challenge for veterinary clinicians**

Fábio P. Sellera<sup>1†</sup>, Miriam R. Fernandes<sup>2†</sup>, Regina Ruiz<sup>3</sup>, Ana C. M. Falleiros<sup>3</sup>, Fernanda P. Rodrigues<sup>3</sup>, Louise Cerdeira<sup>2</sup> and Nilton Lincopan<sup>2,4\*</sup>



2017 = *E. coli* ST648 - bla<sub>KPC-2</sub> - Tn4401b - IncN plasmid