

## Viewpoint

## Is the human T-cell lymphotropic virus type 2 in the process of endogenization into the human genome?



Jorge Casseb<sup>a,\*</sup>, Luiz Mario Janini<sup>b</sup>, Luis Isamu Barros Kanzaki<sup>c</sup>, Luciano Rodrigo Lopes<sup>d</sup>, Arthur Maia Paiva<sup>a,e</sup>

<sup>a</sup> Institute of Tropical Medicine of Sao Paulo - University of Sao Paulo, Laboratory of Medical Investigation LIM-56 / Faculty of Medicine -USP, Brazil

<sup>b</sup> Discipline of Microbiology, Department of Microbiology, Immunology and Parasitology, Federal University of Sao Paulo - Unifesp, Sao Paulo, SP, Brazil

<sup>c</sup> Laboratory of Bioprospection, Department of Pharmacy, Faculty of Health Sciences, University of Brasilia, DF, Brazil

<sup>d</sup> Bioinformatics and Biomedical Data Science Division, Health Informatics Department, Federal University of Sao Paulo - Unifesp, São Paulo, SP, Brazil

<sup>e</sup> University Hospital Alberto Antunes / Federal University of Alagoas, Brazil

## ARTICLE INFO

## Keywords:

Human retroviruses

HTLV-2

Endogenization

Viral pathogenicity

## ABSTRACT

Human T-cell lymphotropic virus type 2 (HTLV-2) infection has been shown to be endemic among intravenous drug users in parts of North America, Europe and Southeast Asia and in a number of Amerindian populations. Despite a 65% genetic similarity and common host humoral response, the human T-cell lymphotropic viruses type 1 (HTLV-1) and 2 display different mechanisms of host interaction and capacity for disease development. While HTLV-1 pathogenicity is well documented, HTLV-2 etiology in human disease is not clearly established. From an evolutionary point of view, its introduction and integration into the germ cell chromosomes of host species could be considered as the final stage of parasitism and evasion from host immunity. The extraordinary abundance of endogenous viral sequences in all vertebrate species genomes, including the hominid family, provides evidence of this invasion. Some of these gene sequences still retain viral characteristics and the ability to replicate and hence are potentially able to elicit responses from the innate and adaptive host immunity, which could result in beneficial or pathogenic effects. Taken together, this data may indicate that HTLV-2 is more likely to progress towards endogenization as has happened to the human endogenous retroviruses millions of years ago. Thus, this intimate association (HTLV-2/human genome) may provide protection from the immune system with better adaptation and low pathogenicity.

## Introduction

Human T-cell lymphotropic viruses, encompassing the human T-cell lymphotropic virus type 1 and 2 (HTLV-1 and HTLV-2), which make up, with the Simian T-cell lymphotropic virus (STLV), the primate T cell-lymphotropic viruses (PTLV), are members of the delta-retrovirus genus.<sup>1</sup> Primate retrovirus cross-species jumps have occurred for hundreds or thousands of years.<sup>2</sup> Continuous interspecies transmission between a non-human and human primate species with overlapping natural habitats is probably the origin of seven HTLV-1 subtypes (A to G) a few thousand years ago.<sup>3,4</sup> Fig. 1 describes a retrovirus phylogenetic inference extracted from the Gypsy Database (GyDB)<sup>5</sup> which shows evolutionary evidence of several viral transmission events between primates, and even between distant species, that have occurred in the past.

Phylogenetic divergence between HTLV-1 and HTLV-2 has occurred more than one million years ago but they still share about 65% of nucleic acid sequences.<sup>6,7</sup> Therefore, despite a significant similarity they have distinct pathogenic properties. HTLV-1 was the first human retrovirus discovered and has mainly been associated with two illnesses, HTLV-1-associated myelopathy/tropical spastic paraparesis (HAM/TSP) and adult T-cell leukemia/lymphoma (ATL).<sup>8-11</sup> On the contrary, HTLV-2 is described as an asymptomatic or minimally infectious agent<sup>12</sup> with just isolated clinical cases reported.<sup>13</sup>

HTLV-1 and HTLV-2 have distinct oncogenic properties.<sup>14,15</sup> They primarily integrate their genomes into T cells, leading them to immortalization.<sup>16</sup> HTLV-1 preferentially infects CD4<sup>+</sup> T cells while HTLV-2 CD8<sup>+</sup> T cells, even though both viruses are detectable in these two populations.<sup>17</sup> Unlike HTLV-1, which is able to induce an aggressive malignant proliferation of activated CD4<sup>+</sup> T cells, such as in ATL,<sup>18</sup>

\* Corresponding author. Dr. Eneas de Carvalho Aguiar, 500; Building II, Third Floor; ZIP code: 05403-907, Brazil.

E-mail addresses: [jcasseb@usp.br](mailto:jcasseb@usp.br), [jcasseb10@gmail.com](mailto:jcasseb10@gmail.com) (J. Casseb).

<https://doi.org/10.1016/j.jve.2020.100009>

Received 15 November 2019; Received in revised form 27 August 2020; Accepted 27 August 2020

Available online 1 September 2020

2055-6640/© 2020 The Authors. Published by Elsevier Ltd. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

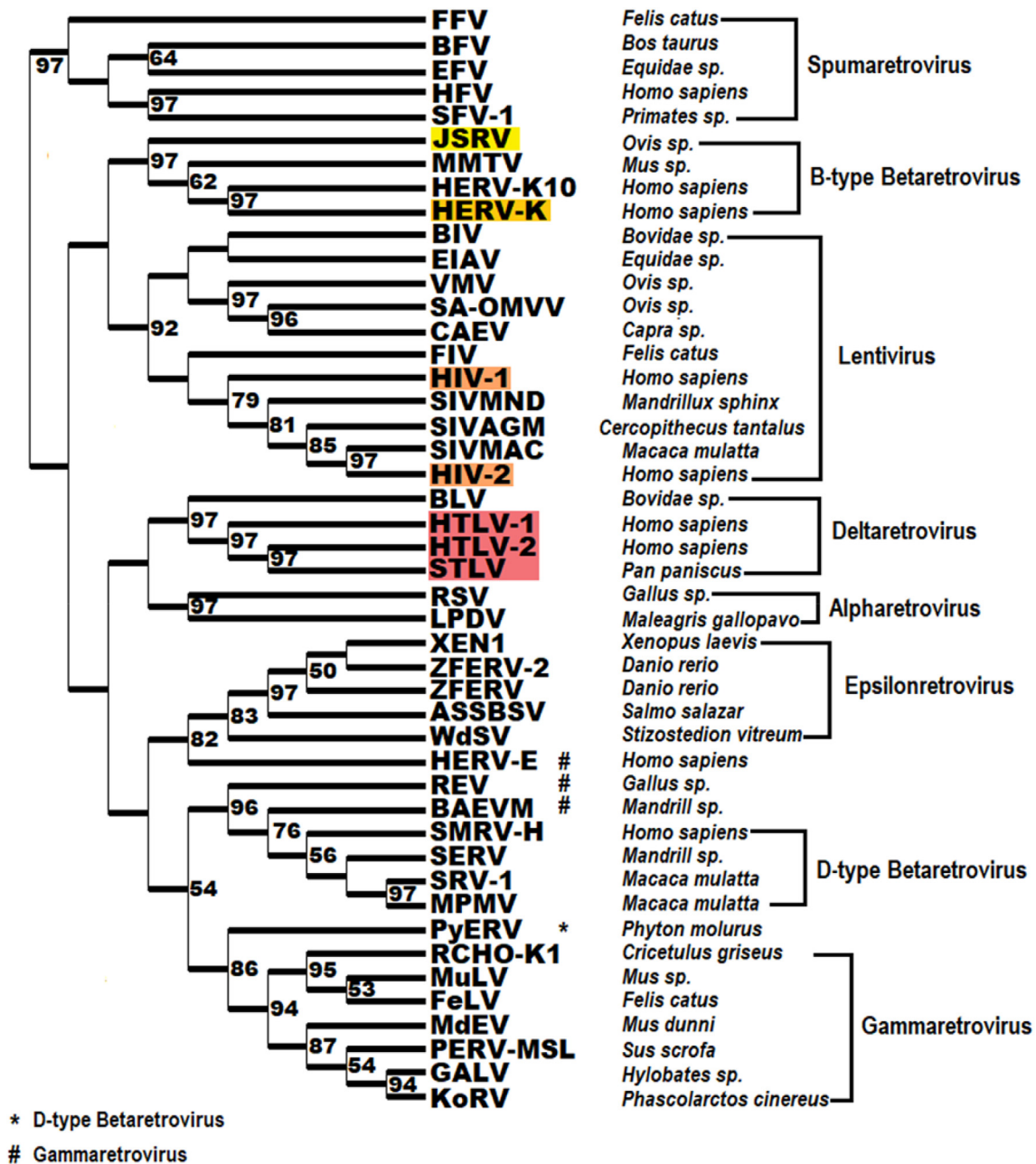


Fig. 1. Evolutionary evidence of transmission events between primates and distant species in the past. Note: Phylogenetic inference extracted from Gypsy Database (GyDB).

HTLV-2 promotes an oligoclonal proliferation of non-malignant CD8<sup>+</sup> T cells.<sup>14</sup>

Estimates of the global number of HTLV-1 and HTLV-2 infected individuals range from 10 to 20 million.<sup>19</sup> Some countries in Africa, the Caribbean basin, South America and Japan are considered to be areas of higher endemicity.<sup>19</sup> HTLV-2 infection has been shown to be endemic among intravenous drug users in parts of North America, Europe and Southeast Asia<sup>20,21</sup> and in a number of Amerindian populations.<sup>22–25</sup> Some indigenous communities, such as the Kayapo, who inhabit the Amazonian basin, have a 47% prevalence of infection.<sup>24</sup> Their ancestors most probably came to the Americas through the land bridge, the Bering strait, connecting Asia to the Americas, up to some twelve thousand years ago.<sup>26</sup> Given the high prevalence of infection in Japan, it is interesting to speculate whether the virus had come to the Americas from Asia with those ancient immigrants. Despite the high HTLV-2 incidence, clinically symptomatic patients are not in large numbers.

Benign or low pathogenic HTLV-2 infection may be related to viral latency provided by accessory proteins.<sup>14</sup> While, HTLV-1 accessory proteins are an important driving force for infectivity, cell proliferation and transformation, the pathogenic impact of HTLV-2 accessory proteins is attenuated.<sup>14</sup> Despite Tax-1 and Tax-2 homology, differences in activity may be responsible for the outcome of the infection.<sup>27</sup> Tax-1, unlike Tax-2, causes DNA damage, activates the non-canonical NF-κB pathway and deregulates autophagy.<sup>14,27–29</sup> HTLV-1 HBZ and HTLV-2 APH-2 viral proteins play a similar role, but with subtle differences resulting in low HTLV-2 pathogenicity. Unlike APH-2, HBZ is a more stable protein and can repress IRF-1, a component of innate immunity, and enhance TGF-beta signaling and subsequent Foxp3 expression, thereby inducing a CD4<sup>+</sup> regulatory T cell phenotype.<sup>30</sup>

Additionally, we note that HTLV-2 infection induces a decrease in beta-chemokine production<sup>31</sup> and seems to be more adapted to the human host which may provide an enhanced type of immune responses

during HIV-1 co-infection as compared to HIV-1 infection alone with delayed progression to AIDS.<sup>32,33</sup> Furthermore, HTLV-2 as mediated by Tax2 expression can down-regulate CCR5 expression on lymphocytes, thereby modulating HIV-1 infection and replication.<sup>34</sup>

As well as the HIV-1/HTLV-2 co-infection relationship, some endogenous retroviruses (ERVs) play a critical role in protecting the host against infection from related pathogenic and exogenous retroviruses.<sup>35</sup> For example, an endogenous version of the Jaagsiekte sheep retrovirus (enJSRV) interferes with the replication of exogenous JSRV, acting like a restriction host mechanism.<sup>36</sup> Furthermore, enJSRV-26, a specific provirus which was integrated in the host genome recently, exerts an antagonistic effect on exogenous beta-retroviruses in sheep.<sup>36</sup> Surprisingly, enJSRV-26 endogenization occurred around 200 years ago after sheep domestication.<sup>35,36</sup>

In the human host, HIV-1 infectivity and core assembly are altered due to the interference of Gag formation by HERV-K Gag particles.<sup>37</sup> HIV elite controllers present HERV-K Gag specific cellular and humoral responses that promote an immunoprotective effect.<sup>38,39</sup> Biological processes that can be favorable to the host are an important requirement for retroviral endogenization. Thus, some ERV proteins play a role in host defenses against retroviral infection. In summary, the protective ability of endogenous retroviruses against infection by related pathogenic retroviruses seems to be an important driving force that positively selected and fixed endogenous retroviruses.<sup>35</sup>

When considering that a “candidate” for viral endogenization should have some degree of adaptation such as a low pathogenic course in exogenous retroviruses, HTLV-2 could be in this process. HTLV-2 has been infecting human beings since the Prehistoric Period and a minority of individuals have presented with an illness. There is obviously an advantage for a virus to have the ability to escape immune selection. However, the HTLV-2 envelope protein becomes non-functional with only a limited number of mutations, which is in contrast to the HIV-1 envelope.<sup>40</sup> The mutation rate is lower in HTLV-2 than in HIV-1, conferred by accurate reverse transcription.<sup>41</sup> Moreover, HTLV-2 uses the host DNA polymerase during clonal expansion of infected cells as a replication strategy, which decreases mutation rates and contributes to viral genomic stability.<sup>42</sup> Compared to endogenous retroviruses, exogenous retroviruses such as HIV-1 and HTLV HTLV-2 have a higher replication rate, embedded into the host genome. A decreasing trend in retroviral replication rate is observed in more recently emerging retroviruses when compared to older ones<sup>43</sup> (see Table 1). Mutation rates are also much lower among ERVs than exogenous retroviruses, since the

former are subject to the lower evolutionary rates of the host genome,<sup>3</sup> in contrast to exogenous retroviruses where the reverse transcription step is not subject to editing or error correction.<sup>44</sup>

Viral endogenization has resulted in a massive insertion of endogenous viral elements from diverse origins and ages which are distributed in the human genome and other vertebrates.<sup>48,49</sup> Retrovirus endogenization seems to be occurring after continuous retroviral invasion and integration in the germ cell genome.<sup>35,50</sup> The HTLV-2 integration locus in T cells is variable, with no specific chromosomal integration site or pattern identified within transformants.<sup>51</sup>

The integration process in the host germ cell genome is also required for the endogenization process. Despite the fact that the HTLV-2 genome has not been found integrated into the germinal cells,<sup>45,46</sup> we can speculate about the possibility of its integration into the germ cell chromosomes. A good example among human retroviruses is HIV-1 which appears to integrate in the male cell genome. HIV-1 DNA was detected in sperm after chromatin decondensation, suggesting a viral presence in the sperm nucleus or integration into its genome.<sup>45</sup> In addition, HIV-1 particles in sperm cells from AIDS patients can be transferred to normal oocytes.<sup>46</sup> Despite the absence of a CD4<sup>+</sup> T cell membrane receptor in sperm cells, HIV-1 can use a galactosylceramide-like compound as an alternative receptor.<sup>46,47</sup> Therefore, due to the molecular plasticity of retroviruses, it is not surprising that HTLV-2 was able to find a receptor to infect germ cells.

From an evolutionary point of view, the exogenous genome integration into the germ cell chromosome of host species could be considered as the final stage of parasitism and evasion from host immunity. The extraordinary abundance of endogenous viral sequences in the genome (more than 8%) of all vertebrate species, including the hominid family, is evidence for this invasion.<sup>52</sup> As expected, evolutionary competition between endogenous and exogenous retroviruses is a continuous balancing process, the potentially pathogenic effects of endogenous viral elements to the host being compensated by its beneficial effects.<sup>52</sup>

Taken together, this data may indicate that HTLV-2, regardless of subtypes, is on its way towards potential endogenization, as shown by the example of an exogenous Koala retrovirus, known for its role in the etiology of neoplasia, that has endogenize in some koala populations.<sup>53</sup> This process may have happened to HERVs millions of years ago.<sup>44</sup> However, pathogenicity is related to cell specificity and not cytotoxicity. A good example is the rabies virus which is hardly cytotoxic but leads to cell death. In contrast, enteroviruses are highly cytotoxic with a rapid turnover but allow patients to recover. One may suggest that if a virus infects the germline, its original pathogenicity may be relevant providing infected people have enough time to reproduce, which would be the case for HTLV-2.<sup>54</sup> Thus, this intimate association (HTLV-2/human genome) provides potential protection from the immune system and some adaptive properties from retroviruses such as HTLV-2. This hypothesis suggests that HTLV-2 represents a possible example of ongoing in vivo endogenization.

Fapesp 2014/22827-7; Ministério da Saúde do Brasil; Fundação Faculdade de Medicina and CNPq Grant to JC: 301275/2019-0.

**Table 1**  
Characteristics of human retroviruses and human diploid cells.

Variable	HIV-1	HTLV-1	HTLV-2	HERV	Human diploid cells
Mutation rate	10 <sup>-4</sup>	10 <sup>-5</sup>	10 <sup>-6</sup>	10 <sup>-6</sup>	10 <sup>-7</sup>
Viral load (no treatment)	High	Low	Very low	Absence	Absence
CD4 <sup>+</sup> T cells loss	Yes	No	No	No	—
CD8 <sup>+</sup> T cells increase	Yes	Yes?	Yes?	No data	—
IL-2 production	decreased	Increased	Very high?	No data	—
Estimated time in human genome (years)	~ 10 <sup>2</sup>	10 <sup>6</sup>	10 <sup>7</sup>	10 <sup>8</sup>	10 <sup>9</sup>
Apoptosis rate	High	Low	Low	Very low	Very low
Morbidity (no treatment)	High	Intermediate	None	None	0

HERV: human endogenous retroviruses; HIV-1 human immunodeficiency virus type 1.

HTLV-1: Human T-cell lymphotropic virus type 1; HTLV-2: Human T-lymphotropic virus type II; IL-2: interleukin-2.

Note: HTLV-3 and HTLV-4 were not included for lack of data.

## References

- Mata EC, Bezerra RM, Proietti Júnior AA, et al. HTLV-1/2 prevalence in two Amazonian communities. *J. Virus Erad.* 4(3):174–178.
- Sharp PM, Hahn BH. The evolution of HIV-1 and the origin of AIDS. *Philos. Trans. R Soc. B Biol. Sci.* 2010 Aug 27;365(1552):2487–2494.
- Greenwood AD, Ishida Y, O'Brien SP, Roca AL, Eiden MV. Transmission, evolution, and endogenization: lessons learned from recent retroviral invasions. *Microbiol. Mol. Biol. Rev. MMBR*; 2017 Dec 13 [cited 2019 Oct 29];82(1). Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC5813887/>.
- Van Dooren S, Salemi M, Vandamme A-M. Dating the origin of the African human T-cell lymphotropic virus type-I (HTLV-I) subtypes. *Mol Biol Evol.* 2001 Apr 1;18(4):661–671.
- Llorens C, Futami R, Covelli L, et al. The Gypsy Database (GyDB) of mobile genetic elements: release 2.0. *Nucleic Acids Res.* 2011 Jan 1;39(suppl\_1):D70–D74.
- Salemi M, Desmyter J, Vandamme AM. Tempo and mode of human and simian T-lymphotropic virus (HTLV/STLV) evolution revealed by analyses of full-genome sequences. *Mol Biol Evol.* 2000 Mar;17(3):374–386.

7. Van Brussel M, Salemi M, Liu HF, et al. The simian T-lymphotropic virus STLV-PP1664 from *Pan paniscus* is distinctly related to HTLV-2 but differs in genomic organization. *Virology*. 1998 Apr 10;243(2):366–379.
8. Gallo RC. History of the discoveries of the first human retroviruses: HTLV-1 and HTLV-2. *Oncogene*. 2005 Sep 5;24(39):5926–5930.
9. Proietti FA, Carneiro-Proietti ABF, Catalan-Soares BC, Murphy EL. Global epidemiology of HTLV-I infection and associated diseases. *Oncogene*. 2005 Sep 5;24(39):6058–6068.
10. Poiesz BJ, Ruscetti FW, Reitz MS, Kalyanaram VS, Gallo RC. Isolation of a new type C retrovirus (HTLV) in primary uncultured cells of a patient with Sézary T-cell leukaemia. *Nature*. 1981 Nov 19;294(5838):268–271.
11. Edlich RF, Arnette JA, Williams FM. Global epidemic of human T-cell lymphotropic virus type-I (HTLV-I). *J Emerg Med*. 2000 Jan;18(1):109–119.
12. Anupam R, Doueiri R, Green PL. The need to accessorize: molecular roles of HTLV-1 p30 and HTLV-2 p28 accessory proteins in the viral life cycle. *Front Microbiol*; 2013 [cited 2018 Oct 22];4. Available from: <https://www.frontiersin.org/articles/10.3389/fmicb.2013.00275/full>.
13. Posada-Vergara MP, Montanheiro P, Fukumori LMI, et al. Clinical and epidemiological aspects of HTLV-II infection in São Paulo, Brazil: presence of Tropical Spastic Paraparesis/HTLV-Associated Myelopathy (TSP/HAM) simile diagnosis in HIV-1-co-infected subjects. *Rev Inst Med Trop São Paulo*. 2006 Aug;48(4):207–210.
14. Ciminale V, Rende F, Bertazzoni U, Romanelli MG. HTLV-1 and HTLV-2: highly similar viruses with distinct oncogenic properties. *Front Microbiol*; 2014 Jul 29 [cited 2018 Oct 22];5. Available from: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4114287/>.
15. Rende F, Cavallari I, Romanelli MG, Diani E, Bertazzoni U, Ciminale V. Comparison of the genetic organization, expression strategies and oncogenic potential of HTLV-1 and HTLV-2. *Leuk Res. Treat*; 2012 [cited 2019 Oct 24];2012. Available from: <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3504254/>.
16. Feuer G, Green PL. Comparative biology of human T-cell lymphotropic virus type 1 (HTLV-1) and HTLV-2. *Oncogene*. 2005 Sep 5;24(39):5996–6004.
17. Kannian P, Yin H, Doueiri R, Lairmore MD, Fernandez S, Green PL. Distinct transformation tropism exhibited by human T lymphotropic virus type 1 (HTLV-1) and HTLV-2 is the result of postinfection T cell clonal expansion. *J Virol*. 2012 Apr;86(7):3757–3766.
18. Ahmadi Ghezeldasht S, Shirdel A, Assarehzadegan MA, et al. Human T lymphotropic virus type I (HTLV-I) oncogenesis: molecular aspects of virus and host interactions in pathogenesis of adult T cell leukemia/lymphoma (ATL). *Iran J. Basic Med. Sci*. 2013 Mar;16(3):179–195.
19. Gessain A, Cassar O. Epidemiological aspects and world distribution of HTLV-1 infection. *Front Microbiol*. 2012;3:388.
20. Zella D, Mori L, Sala M, et al. HTLV-II infection in Italian drug abusers. *Lancet Lond. Engl*. 1990 Sep 1;336(8714):575–576.
21. Fukushima Y, Takahashi H, Hall WW, et al. Extraordinary high rate of HTLV type II seropositivity in intravenous drug abusers in south Vietnam. *AIDS Res. Hum. Retrovir*. 1995 May;11(5):637–645.
22. Biglione M, Vidan O, Mahieux R, et al. Seroepidemiological and molecular studies of human T cell lymphotropic virus type II, subtype b, in isolated groups of Mataco and Toba Indians of northern Argentina. *AIDS Res. Hum. Retrovir*. 1999 Mar 20;15(5):407–417.
23. Duenas-Barajas E, Bernal JE, Vaught DR, et al. Human retroviruses in Amerindians of Colombia: high prevalence of human T cell lymphotropic virus type II infection among the Tunebo Indians. *Am J Trop Med Hyg*. 1993 Dec;49(6):657–663.
24. Black FL, Biggar RJ, Neel JV, Maloney EM, Waters DJ. Endemic transmission of HTLV type II among Kayapo Indians of Brazil. *AIDS Res. Hum. Retrovir*. 1994 Sep;10(9):1165–1171.
25. Ferrer JF, Del Pino N, Esteban E, et al. High rate of infection with the human T-cell leukemia retrovirus type II in four Indian populations of Argentina. *Virology*. 1993 Dec;197(2):576–584.
26. Paiva A, Casseb J, Paiva A, Casseb J. Origin and prevalence of Human T-lymphotropic virus type 1 (HTLV-1) and type 2 (HTLV-2) among indigenous populations in the Americas. *Rev Inst Med Trop São Paulo*. 2015 Feb;57(1), 01–14.
27. Harrod R. Silencers of HTLV-1 and HTLV-2: the pX-encoded latency-maintenance factors. *Retrovirology*. 2019 Sep 6;16(1):25.
28. Semmes OJ, Majone F, Cantemir C, Turchetto L, Hjelle B, Jeang KT. HTLV-I and HTLV-II Tax: differences in induction of micronuclei in cells and transcriptional activation of viral LTRs. *Virology*. 1996 Mar 1;217(1):373–379.
29. Ren T, Takahashi Y, Liu X, et al. HTLV-1 Tax deregulates autophagy by recruiting autophagic molecules into lipid raft microdomains. *Oncogene*. 2015 Jan 15;34(3):334–345.
30. Panfil AR, Dissinger NJ, Howard CM, et al. Functional comparison of HBZ and the related APH-2 protein provides insight into human T-cell leukemia virus type 1 pathogenesis. *J Virol*. 2016 Jan 27;90(7):3760–3772.
31. Olah I, Fukumori LMI, Montanheiro P, et al. Patterns of in vitro lymphoproliferative responses among HTLV-1-infected subjects: upregulation by HTLV-1 during HIV-1 co-infection. *Scand J Immunol*. 2007 Jun;65(6):577–580.
32. Turci M, Pilotti E, Ronzi P, et al. Coinfection with HIV-1 and human T-Cell lymphotropic virus type II in intravenous drug users is associated with delayed progression to AIDS. *J. Acquir. Immune Defic. Syndr*. 1999. 2006 Jan 1;41(1):100–106.
33. Beilke MA, Theall KP, O'Brien M, et al. Clinical outcomes and disease progression among patients coinfecting with HIV and human T lymphotropic virus types 1 and 2. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc Am*. 2004 Jul 15;39(2):256–263.
34. Barrios CS, Abuerreish M, Lairmore MD, Castillo L, Giam C-Z, Beilke MA. Recombinant human T-cell leukemia virus types 1 and 2 Tax proteins induce high levels of CC-chemokines and downregulate CCR5 in human peripheral blood mononuclear cells. *Viral Immunol*. 2011 Dec;24(6):429–439.
35. Varela M, Spencer TE, Palmarini M, Arnaud F. Friendly viruses: the special relationship between endogenous retroviruses and their host. *Ann N Y Acad Sci*. 2009 Oct;1178:157–172.
36. Arnaud F, Caporale M, Varela M, et al. A paradigm for virus–host coevolution: sequential counter-Adaptations between endogenous and exogenous retroviruses. *PLoS Pathog*. 2007 Nov 9;3(11):e170.
37. Monde K, Terasawa H, Nakano Y, et al. Molecular mechanisms by which HERV-K Gag interferes with HIV-1 Gag assembly and particle infectivity. *Retrovirology*; 2017 Apr 26 [cited 2019 Oct 27];14. Available from <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5406883/>.
38. de Mulder M, SenGupta D, Deeks SG, et al. Anti-HERV-K (HML-2) capsid antibody responses in HIV elite controllers. *Retrovirology*; 2017 Aug 22 [cited 2019 Oct 27];14. Available from: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5568399/>.
39. Michaud H-A, SenGupta D, de Mulder M, et al. Cutting edge: an antibody recognizing ancestral endogenous virus glycoproteins mediates antibody-dependent cellular cytotoxicity on HIV-1-infected cells. *J. Immunol. Baltim. Md*. 1950. 2014 Aug 15;193(4):1544–1548.
40. Pique C, Tursz T, Dokhelar MC. Mutations introduced along the HTLV-I envelope gene result in a non-functional protein: a basis for envelope conservation? *EMBO J*. 1990 Dec;9(13):4243–4248.
41. Martin JL, Maldonado JO, Mueller JD, Zhang W, Mansky LM. Molecular studies of HTLV-1 replication: an update. *Viruses*. 2016 Jan 27;8(2).
42. Salemi M, Vandamme AM, Desmyter J, Casoli C, Bertazzoni U. The origin and evolution of human T-cell lymphotropic virus type II (HTLV-II) and the relationship with its replication strategy. *Gene*. 1999 Jun 24;234(1):11–21.
43. Aiweesakun P, Katzourakis A. Endogenous viruses: connecting recent and ancient viral evolution. *Virology*. 2015 May 1;479–480:26–37.
44. Weiss RA. Human endogenous retroviruses: friend or foe? *APMIS Acta Pathol. Microbiol. Immunol. Scand*. 2016 Feb;124(1–2):4–10.
45. Muciaccia B, Corallini S, Vicini E, et al. HIV-1 viral DNA is present in ejaculated abnormal spermatozoa of seropositive subjects. *Hum Reprod*. 2007 Nov 1;22(11):2868–2878.
46. Baccetti B, Benedetto A, Burrini AG, et al. HIV-particles in spermatozoa of patients with AIDS and their transfer into the oocyte. *J Cell Biol*. 1994 Nov;127(4):903–914.
47. Wang D, Li L-B, Hou Z-W, et al. The integrated HIV-1 provirus in patient sperm chromosome and its transfer into the early embryo by fertilization. *PLoS One*; 2011 Dec 14 [cited 2019 Oct 30];6(12). Available from: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3237474/>.
48. Feschotte C, Gilbert C. Endogenous viruses: insights into viral evolution and impact on host biology. *Nat Rev Genet*. 2012 Apr;13(4):283–296.
49. Johnson WE. Endogenous retroviruses in the genomics era. *Annu. Rev. Virol*. 2015;2(1):135–159.
50. Shimode S, Nakagawa S, Miyazawa T. Multiple invasions of an infectious retrovirus in cat genomes. *Sci Rep*. 2015 Feb 2;5:8164.
51. Tarsis SL, Yu M-T, Parks ES, Persaud D, Muñoz JL, Parks WP. Human T-lymphocyte transformation with human T-cell lymphotropic virus type 2. *J Virol*. 1998 Jan 1;72(1):841–846.
52. Kassiotis G, Stoye JP. Immune responses to endogenous retroelements: taking the bad with the good. *Nat Rev Immunol*. 2016 Apr;16(4):207–219.
53. Tarlinton RE, Meers J, Young PR. Retroviral invasion of the koala genome. *Nature*. 2006;442(7098):79–81.
54. Dietzschold B, Li J, Faber M, Schnell M. Concepts in the pathogenesis of rabies. *Future Virol*. 2008;3(5):481–490. <https://doi.org/10.2217/17460794.3.5.481>.