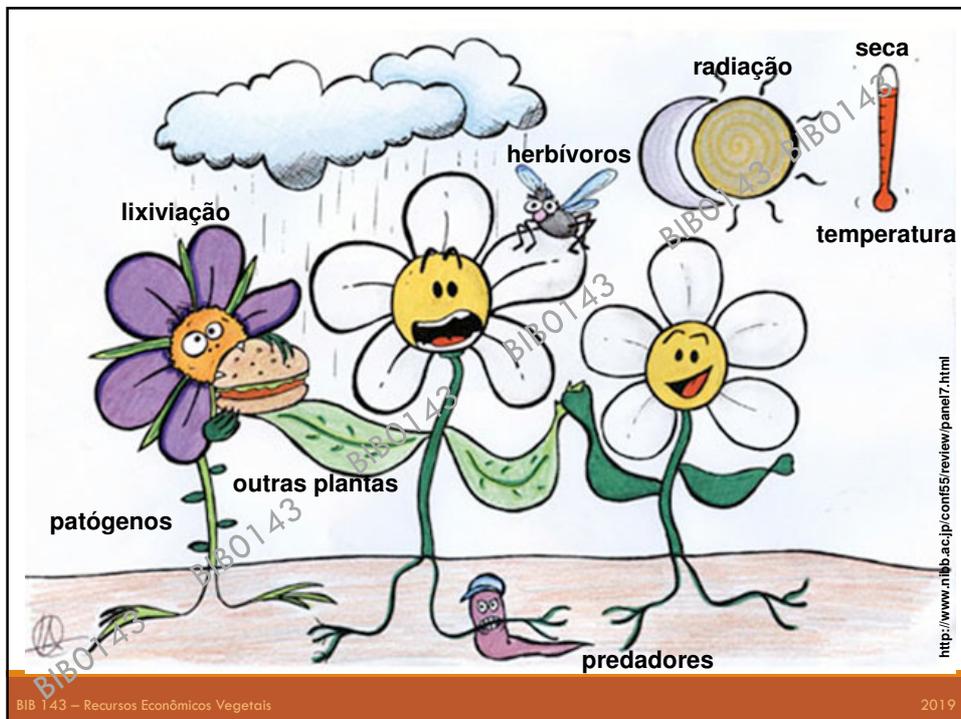
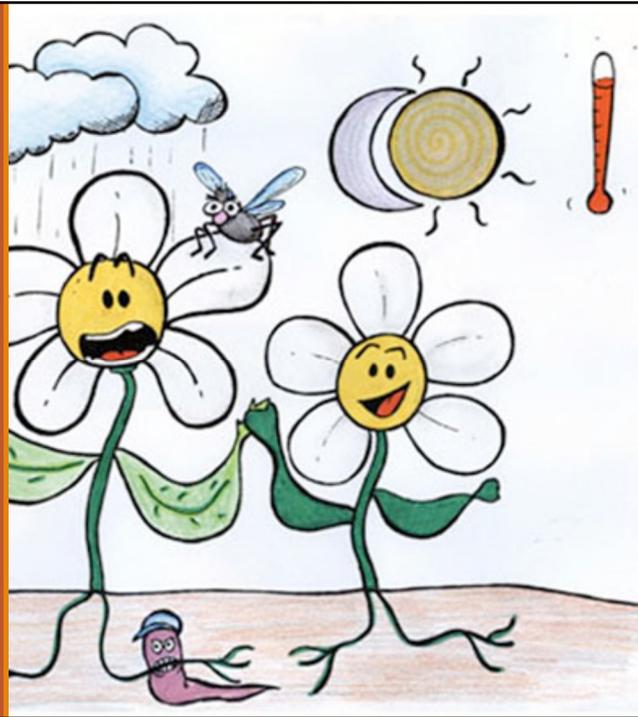
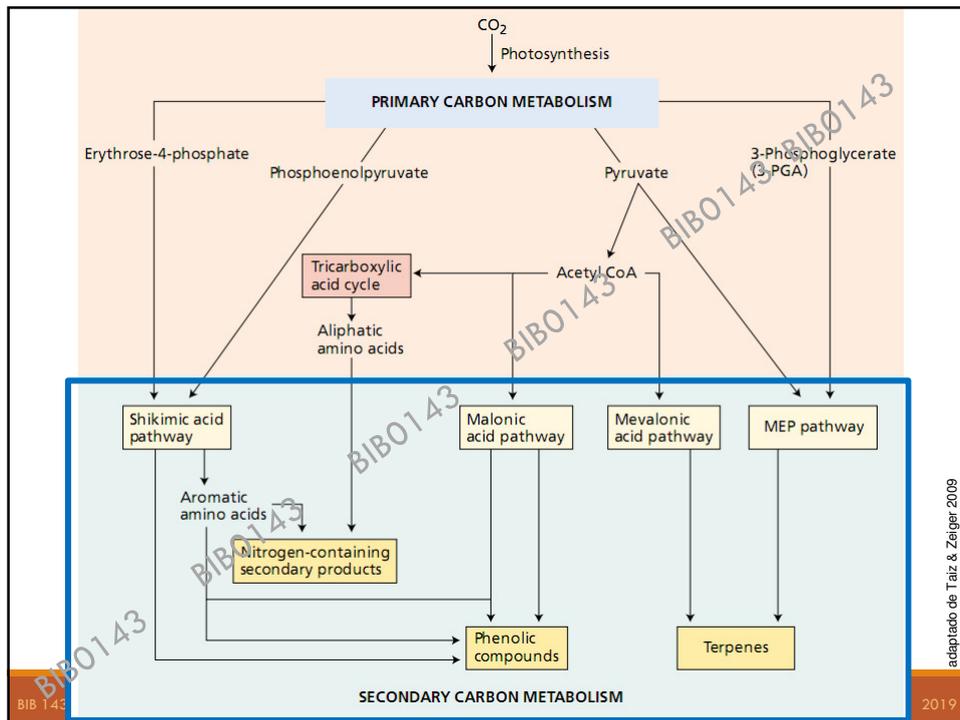


Interações planta-ambiente/herbívoros: aspectos econômicos e ecológicos

Déborah Yara A. C. dos Santos  
dyacsan@ib.usp.br





## METABÓLITOS SECUNDÁRIOS

**mentol**

CC1=C(C(C1)O)C(C)C

**cafeína**

CN1C=NC2=C1C(=O)N(C(=O)N2C)C

**ácido ursólico**

CC12C(C3C(C1)O)C(C(C2)O)C3

Promovem proteção contra herbívoros (insetos) como dissuasores alimentares, redutores de crescimento, toxinas, etc

Interferência bioquímica com outras plantas - alelopatia

Proporciona proteção contra patógenos (fungos) - fitoalexinas

- Podem servir como sinalizadores para nodulação em leguminosas e para polinizadores
- ....

**ácido jasmônico**

CC1=CC(=O)C=C1

**calatropina**

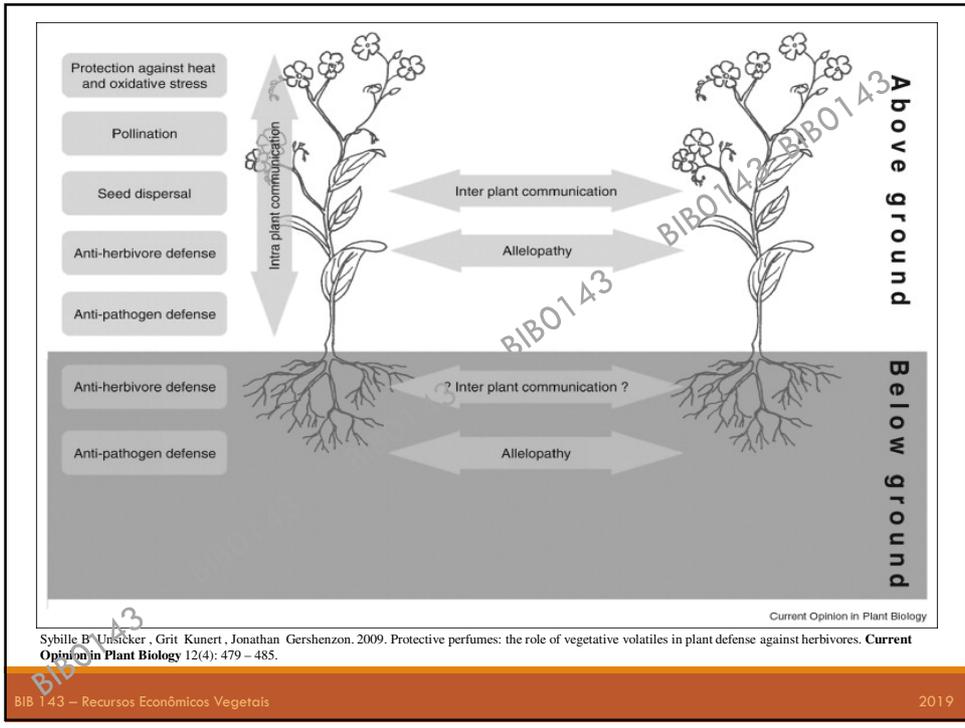
CC12C(C3C(C1)O)C(C(C2)O)C3

**cianidina-3-O-glucosídeo**

O=C1C=CC(=C2C(=C1)O)O2

**Metabólitos secundários são substâncias que NÃO participam dos processos de formação de protoplasto e geração de energia; são MEDIADORES em processos de interação das plantas com o ambiente; NÃO são UNIVERSAIS; apresentam ampla diversidade estrutural.**

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### INTERAÇÕES COM FATORES BIÓTICOS

#### POLINIZAÇÃO

The illustration shows a green, anthropomorphic flower with a smiling face and a yellow and black striped bee. The flower is holding a small brown object, possibly a seed or a piece of pollen, and the bee is interacting with it. A watermark 'BIB0143' is visible across the illustration. A URL is provided at the bottom right: <https://historiasnaturais.wordpress.com/page/2/>

Protection against heat and oxidative stress

Pollination

Seed dispersal

Anti-herbivore defense

Anti-pathogen defense

Anti-herbivore defense

Anti-pathogen defense

Sybille B. Unsicker, Grit Kunert, Jonathan Gershenzon. 2009. Protective perfumes: the role of vegetative volatiles in plant defense against herbivores. **Current Opinion in Plant Biology** 12(4): 479 – 485.

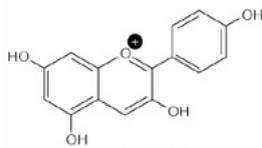
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## POLINIZAÇÃO

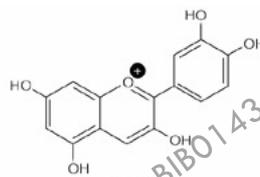


- ✓ relação benéfica aos dois envolvidos: animal – néctar e pólen; planta – transferência do pólen;
- ✓ a reprodução sexuada nas plantas envolve a polinização;
- ✓ a polinização, em especial a cruzada, amplia e garante a variabilidade genética dos vegetais;
- ✓ a relação entre o agente polinizador e a planta pode ser generalista ou especialista;
- ✓ os agentes polinizadores mais abundantes em angiospermas são os insetos.

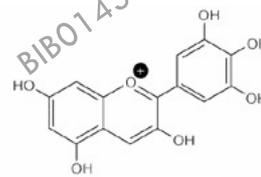
## Como os metabólitos secundários estão envolvidos no processo de polinização?



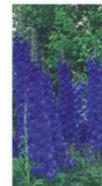
pelargonidina

*Pelargonium*  
(Ceranium)

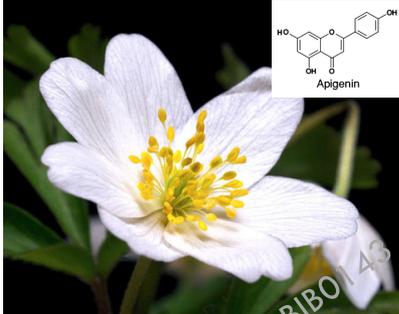
cianidina

*Rosa*  
(Rose)

delfinidina

*Delphinium*  
(Larkspur)

visão humana



visão abelha



Apigenin

Oc1ccc(O)c2c(c1)oc3c(O)c(O)c(O)c3o2

<http://www.dailymail.co.uk/science/article-473897/A-bees-eye-view-How-insects-flowers-differently-us.html>

BIB0143

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Ecological Economics 71 (2011) 80–88

Contents lists available at SciVerse ScienceDirect

ELSEVIER Ecological Economics journal homepage: www.elsevier.com/locate/econecol

Analysis

Valuing pollination services to agriculture

Rachael Winfree <sup>a,\*</sup>, Brian J. Gross <sup>b,1</sup>, Claire Kremen <sup>c</sup>

<sup>a</sup> Department of Entomology, 93 Lipman Dr., Rutgers University, New Brunswick, NJ 08901, USA  
<sup>b</sup> Food and Resource Economics, University of British Columbia, Vancouver, Canada, BC V6T1Z4  
<sup>c</sup> Department of Environmental Science, Policy and Management, University of California, Berkeley, Berkeley, CA 94720, USA



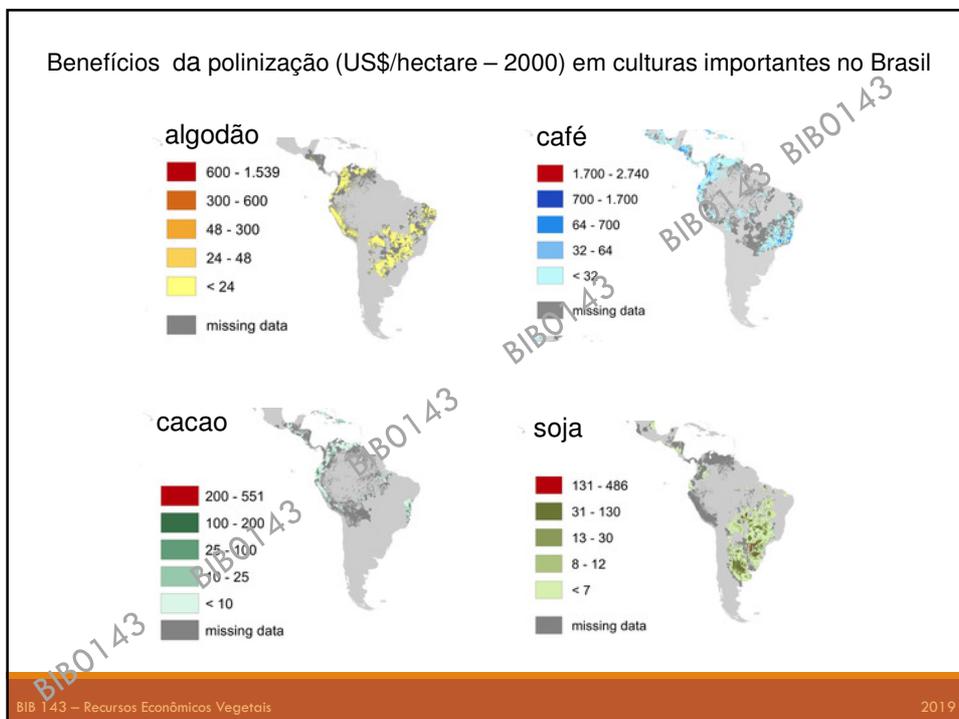
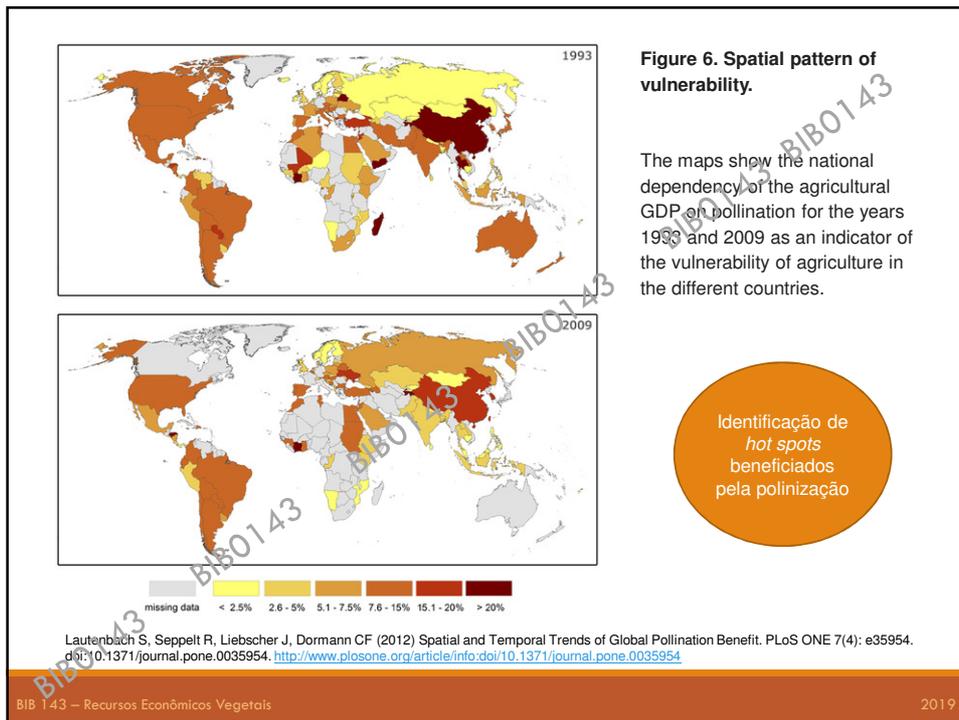
<https://en.wikipedia.org/wiki/Entomophily>

A polinização por animais em culturas é um importante **serviço ecossistêmico!**

- ✓ Cerca de 35% da produção global de alimento depende de agente polinizador

BIB0143

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Analysis

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<https://en.wikipedia.org/wiki/Entomophily>

A polinização por animais em culturas é um importante **serviço ecossistêmico!**

- ✓ Cerca de 35% da produção global de alimento depende de agente polinizador
  - ✓ Abelhas (Hymenoptera: Apiformes) - *Apis mellifera* 🐝
  - ✓ 17.000 espécies de abelhas nativas: café, melão, tomate, canola, girassol

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Protection against heat and oxidative stress

Pollination

Seed dispersal

**Anti-herbivore defense**

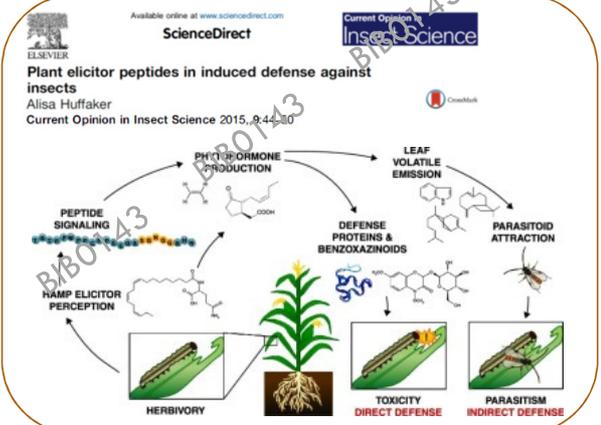
Anti-pathogen defense

Anti-herbivore defense

Anti-pathogen defense

## INTERAÇÕES COM FATORES BIÓTICOS

### AÇÃO ANTI-HERBIVORIA



Available online at [www.sciencedirect.com](http://www.sciencedirect.com)

ScienceDirect

Current Opinion in Insect Science

**Plant elicitor peptides in induced defense against insects**

Alisa Huffaker  
 Current Opinion in Insect Science 2015, 2:44–50

Sybille B. Unsicker, Grit Kunert, Jonathan Gershenzon. 2009. Protective perfumes: the role of vegetative volatiles in plant defense against herbivores. *Current Opinion in Plant Biology* 12(4): 479 – 485.

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## INTERAÇÕES COM FATORES BIÓTICOS

**VOCs**  
(volatile organic compounds)  
**GLVs**  
(green leaf volatiles)

Inter plant communication

Allelopathy

? Inter plant communication ?

Allelopathy

Intra plant communication

**Root-emitted volatile organic compounds can they mediate belowground plant-plant interactions?**

Benjamin M. Doherty · Pierre Delgado · Patrick J. Leffler · Plant Soil DOI 10.1007/s11104-016-2823-3

Fig. 2 Overview of the known and potential mechanisms mediating plant-plant interactions above and below the soil surface. Aboveground, both the variation in red/far-red and blue light levels in the plant's canopy and the volatile composition emitted by plants in the atmosphere enable plants to detect the presence of neighbours. Laboratory and field experiments have shown that VOCs are involved in within-plant and between-plant signalling aboveground. Belowground, plant-plant interactions are mediated by non-volatile exudates, the transport of compounds through common mycorrhizal networks (CMN), and indirect plant-plant interactions (alteration of soil chemistry, microbial populations and nutrient availability) have also been reported in the literature. The green arrows refer to plant-plant interactions mediated by root VOCs. 1, within-plant signalling (shoot-shoot or root-root); 2, intraspecific between-plant signalling (shoot-shoot or root-root); 3, interspecific between-plant signalling (shoot-shoot or root-root); 4, within-plant root-shoot signalling; +, attraction of parasitic plants; solid and labelled arrows, VOC emission. (Drawing: In Carolina Leivick)

Red/far-red and blue light levels

Parasitic Plants

CMN

Non-volatile exudates  
Indirect plant-plant interactions

Sybille B Unsicker, Grit Kunert, Jonathan Gershenzon, 2009. Protective perfumes: volatiles in plant defense against herbivores. *Current Opinion in Plant Biology* 12:4

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## O INIMIGO DO MEU INIMIGO É MEU ALIADO

3. Voláteis inibem a oviposição pelo adulto de *Manduca*

2. Herbivoria induz liberação de voláteis que atrai inimigos naturais da lagarta

1. Plantas de tabaco predadas pela lagarta

Sabelis et al. 2001. The enemy of my enemy is my ally. *Science* 291: 2104-5

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2019

## PLANT DEFENCES

Pesticide Outlook – June 2003

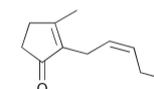
### CIS-JASMONE SWITCHES ON PLANT DEFENCE AGAINST INSECTS

Toby Bruce, John Pickett and Lesley Smart from Rothamsted Research at Harpenden in the UK describe a volatile plant activator which could have an important part to play in plant defence mechanisms

Substâncias voláteis induzidas em plantas injuriadas:

1. Repelente de insetos fitófagos.
2. Defesa indireta – atração de predador ou parasita do fitófago.
3. Sinais entre plantas – indução de mecanismos de defesa em plantas vizinhas antes do ataque dos herbívoros.

**cis-Jasmone**



Trigo (*Triticum aestivum*) x afídeo (*Sitobion avenae*) x parasitoide (*Aphidius ervi*)

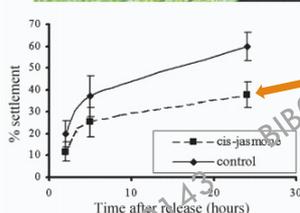


Figure 1 Settlement of *S. avenae* in simulator bioassay

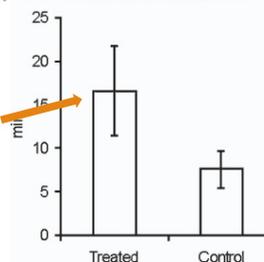
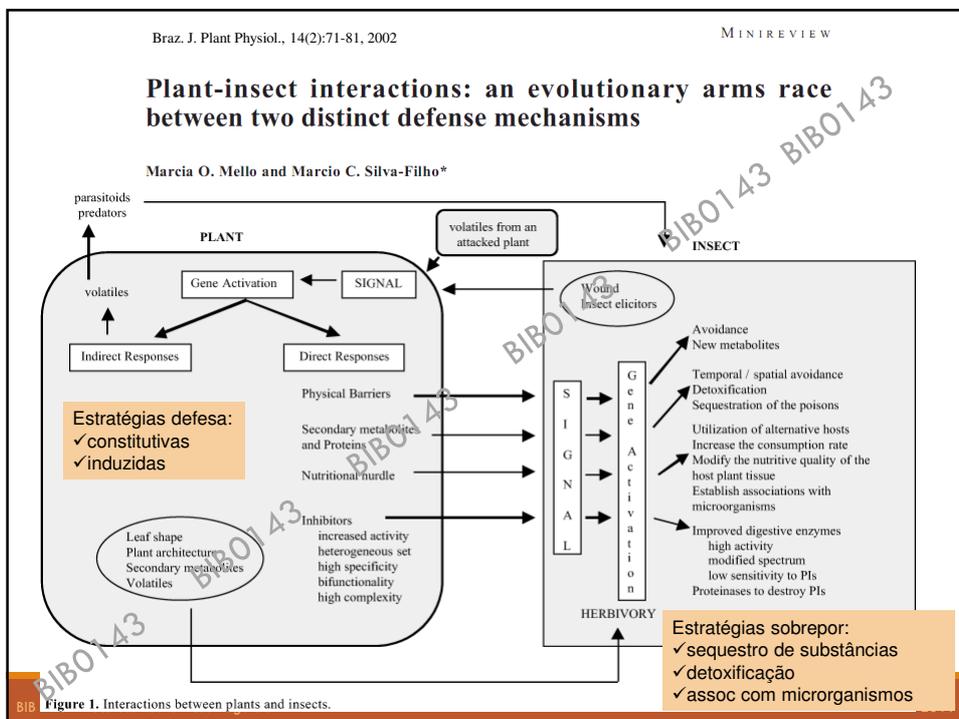
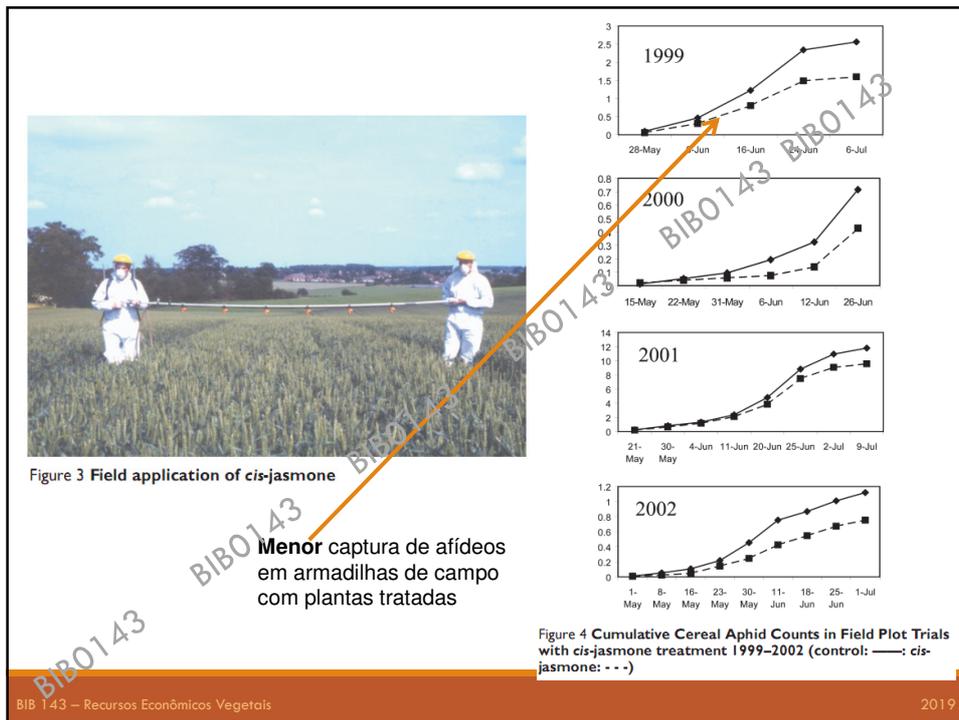


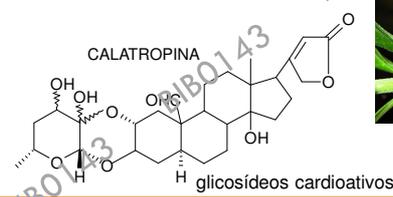
Figure 2 Time spent foraging by *Aphidius ervi* on cis-jasmone and control wheat seedlings.

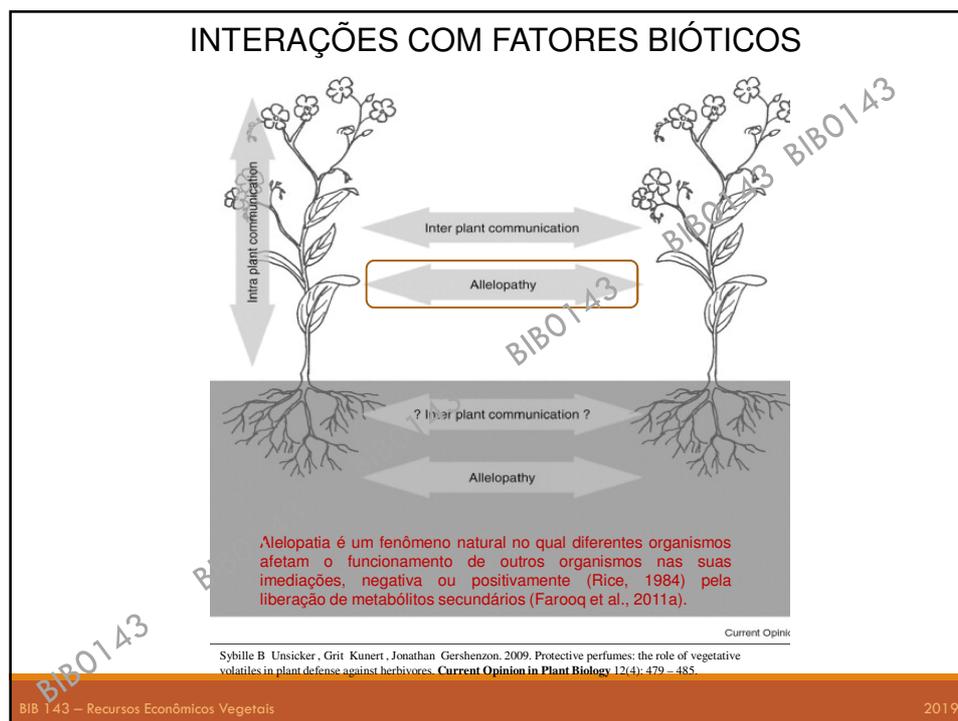
Plantas tratadas com sinalizador são **menos** atrativas ao herbívoro e **mais** ao seu parasita



Estratégias defesa nos insetos: sequestro de substâncias

monarca x *Asclepias* x pássaro





INTERNATIONAL JOURNAL OF AGRICULTURE & BIOLOGY *Int. J. Agric. Biol., Vol. 15, No. 6, 2013*  
 ISSN Print: 1560–8530; ISSN Online: 1814–9596  
 13S–011/2013/15–6–1367–1378  
<http://www.fspublishers.org>

**Review Article**

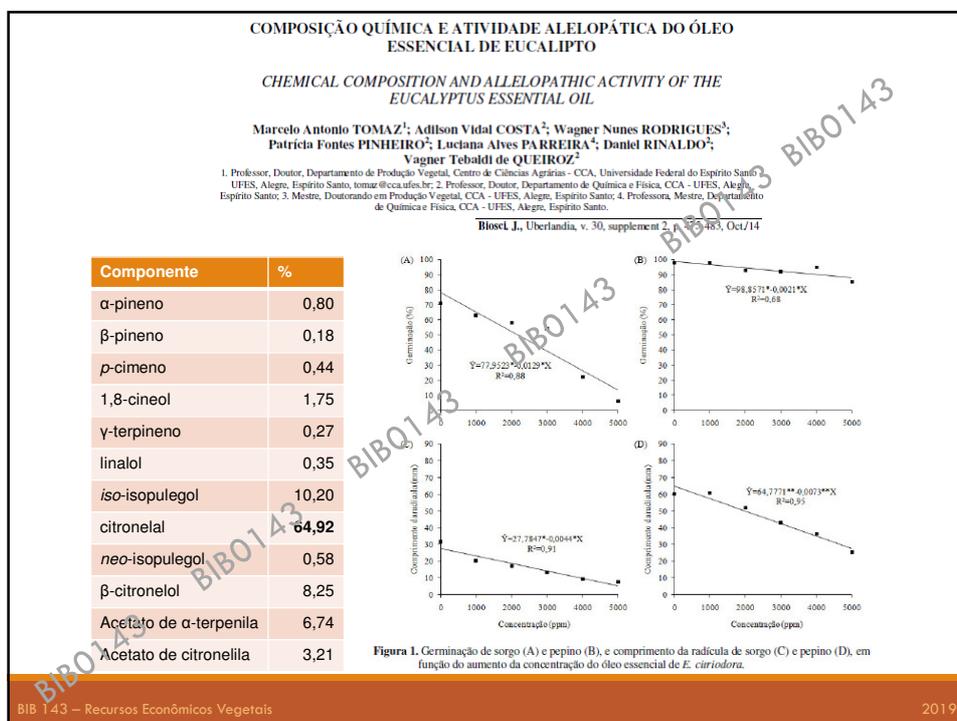
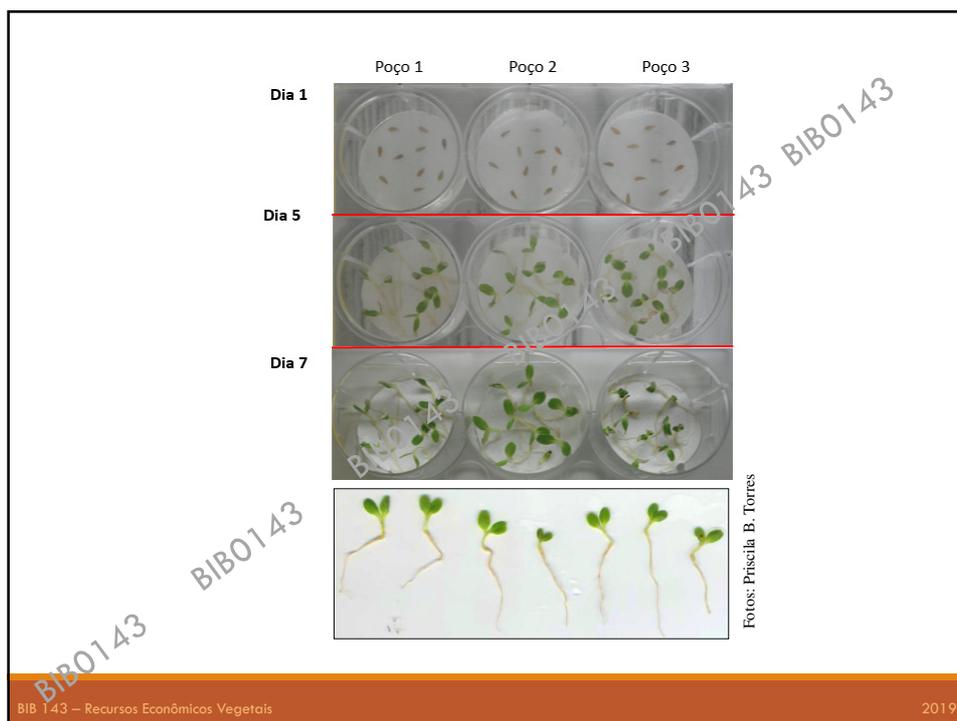
### Application of Allelopathy in Crop Production

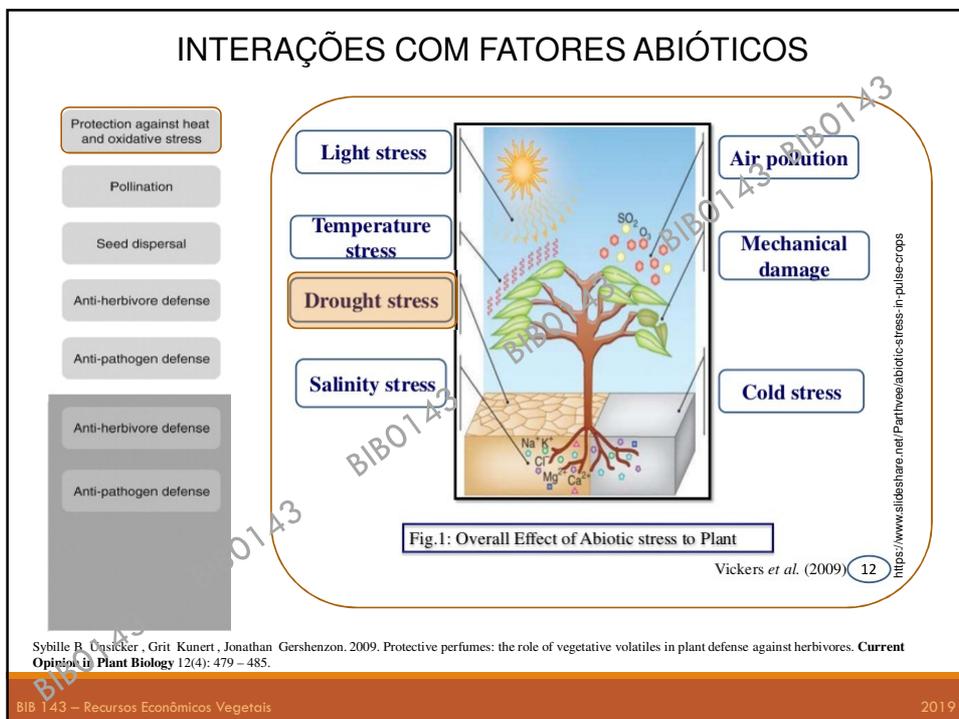
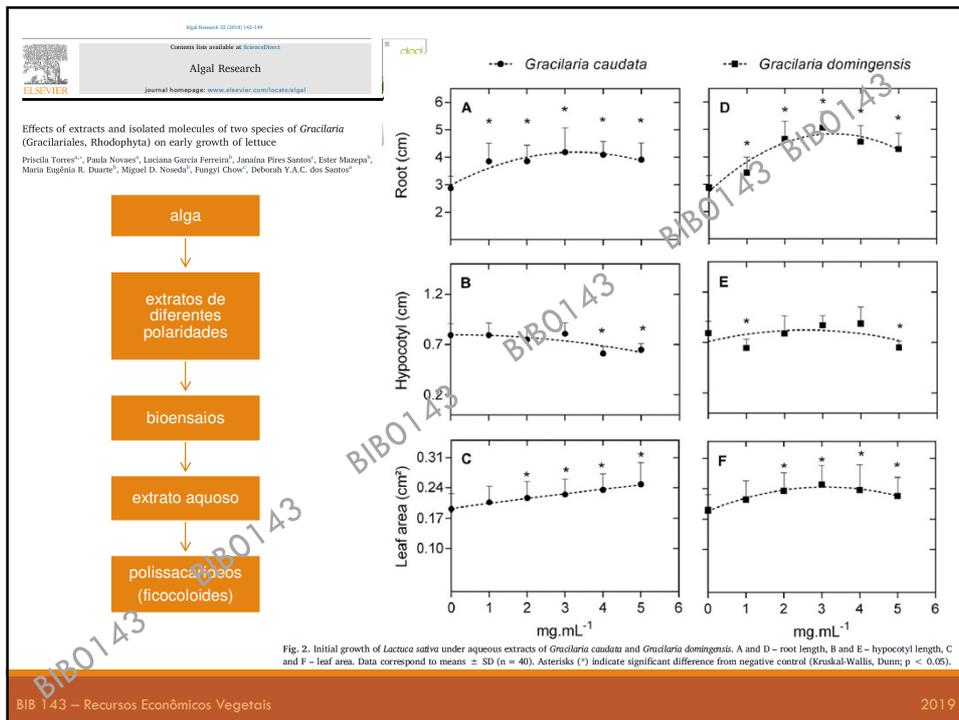
Muhammad Farooq<sup>1\*</sup>, Ali Ahsan Bajwa<sup>1</sup>, Sardar A. Cheema<sup>1</sup> and Zahid A. Cheema<sup>1</sup>  
<sup>1</sup>Allelopathy Laboratory, Department of Agronomy, University of Agriculture, Faisalabad-38040, Pakistan  
 \*For correspondence: farooqcp@gmail.com

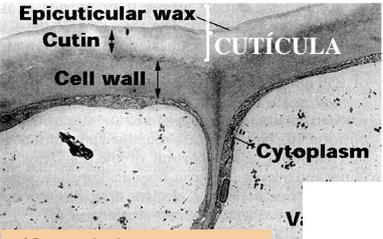
**Table 1: Weed control through allelopathic crop water extracts**

Allelopathic extract	Crop	Weeds controlled	Weed control		Yield in increase over control (%)	Reference
			Reduction in weed density (%)	Reduction in dry weight (%)		
Sorghum	Wheat	<i>Fumaria indica</i> , <i>Phalaris minor</i> , <i>Rumex dentatus</i> , <i>Chenopodium album</i>	21.6-44.2	35.4-49.0	11.0-20.0	Cheema and Khaliq (2000)
	Cotton	<i>Trianthema portulacastrum</i> , <i>Cynodon dactylon</i> , <i>Cyperus rotundus</i>	47.0	29.0-40.1	17.7-59.0	Cheema et al. (2002)
	Mungbean	<i>Cyperus rotundus</i> , <i>Chenopodium album</i> , <i>Convolvulus arvensis</i>	17.5-31.6	23.7-59.6	4.0-17.7	Cheema et al. (2001)
Rice		<i>Echinochloa colonum</i> , <i>Cyperus rotundus</i> , <i>Cyperus iria</i>	-	40.4	12.5	Wazir et al. (2011)
	Sunflower	<i>Avena fatua</i> , <i>Melilotus officinalis</i> , <i>Phalaris minor</i> , <i>Rumex obtusifolius</i>	10.6-33.6	2.2-16.5	1.6-10.7	Cheema et al. (2003), Naseem et al. (2010)
Sorghum + Sunflower	Wheat	<i>Avena fatua</i> , <i>Phalaris minor</i>	-	10.0-62.0	18.55-62.0	Jamil et al. (2009)
Sorghum + Brassica						
Sorghum + Tobacco						
Sorghum + Sesame						

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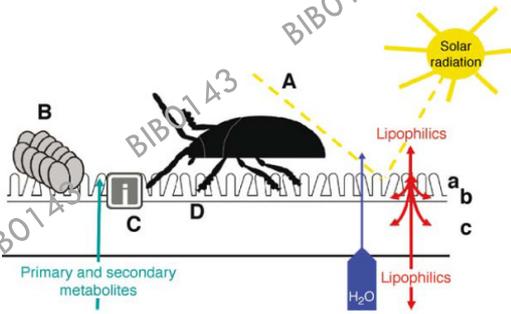






**Ceras cuticulares**

- ✓ Camada fotoprotetora
- ✓ Minimizar a adesão de resíduos nas superfícies das plantas
- ✓ Limitar perda de água não estomática
- ✓ Mediação da interação das plantas com ambiente e herbívoros/patógenos
- ✓ Manutenção do correto desenvolvimento de vários órgãos

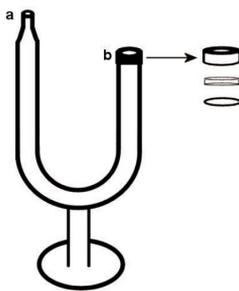


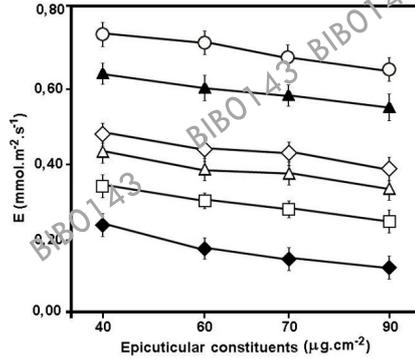
**Fig. 4. Processes on the plant surface.** (A) Reflectance, transmittance, and absorbance of solar radiation; (B) attachment of insect eggs (or spores of microorganisms, likewise); (C) physical and chemical cues used for host recognition by microorganisms and insects; (D) adhesion of insect legs and influences of surface characteristics on locomotion. Water and other metabolites diffuse in different degrees through the cuticular layer, formed by epicuticular wax crystals (a), the epicuticular wax film (b), and cutin and intracuticular waxes (c).

Müller, C. & Friederer, M. J. Chemical Ecology 31: 2621-51. (2005)

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**Hidrofobicidade** da cutícula depende da composição relativa das frações de hidrocarbonetos, álcoois e aldeídos (mais relacionada com as ceras);





Epicuticular constituents (µg.cm <sup>-2</sup> )	Ursolic acid (○)	hentriacontan-16-one (▲)	lupeol (◇)	lupeol + β-amyrin (△)	epifriedelinol (□)	n-alkanes (◆)
40	0.75	0.65	0.48	0.42	0.35	0.25
60	0.72	0.62	0.45	0.38	0.32	0.22
70	0.68	0.58	0.42	0.35	0.28	0.20
90	0.65	0.55	0.38	0.32	0.25	0.18

**alcanos** são mais eficientes que **terpenoides** como barreiras a perda de água

Oliveira & al. Epicuticular waxes from caatinga and cerrado species and their efficiency against water loss. Anais da Academia Brasileira de Ciências (2003) 75(4): 431-439

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A composição de cada fração depende da espécie e afeta diretamente a **condutância epidérmica** e a estrutura da cera na superfície;

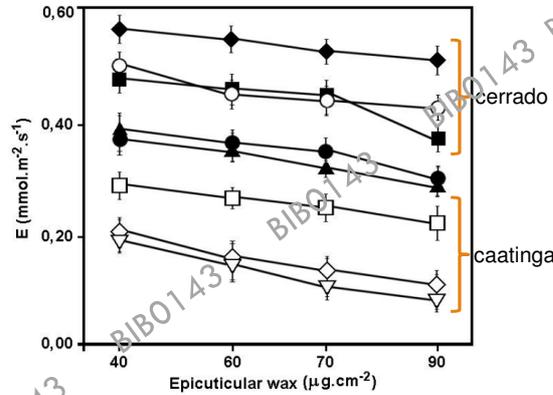


Fig. 2 – Evaporation rates ( $E$ ) in Whatman paper discs impregnated with foliar epicuticular waxes of species from caatinga (empty symbols) and cerrado (full symbols). (○) *Aspidosperma pyrifolium*, (◇) *Capparis yco*, (□) *Maytenus rigida*, (▽) *Ziziphus joazeiro*, (■) *Aristolochia esperanzae*, (▲) *Didymopanax vinosum*, (●) *Strychnos pseudoquina*, (◆) *Tocoyena formosa*. Values correspond to means  $\pm$  sd ( $n = 30$ ), obtained at 25°C and 65% relative humidity.

### Increased Accumulation of Cuticular Wax and Expression of Lipid Transfer Protein in Response to Periodic Drying Events in Leaves of Tree Tobacco<sup>11(W)</sup>

Kimberly D. Cameron, Mark A. Teece, and Lawrence B. Smart\*  
 Faculty of Environmental and Forest Biology (K.D.C., L.B.S.) and Faculty of Chemistry (M.A.T.), State University of New York, College of Environmental Science and Forestry, Syracuse, New York 13210  
 Plant Physiology, January 2006, Vol. 140, pp. 176–183

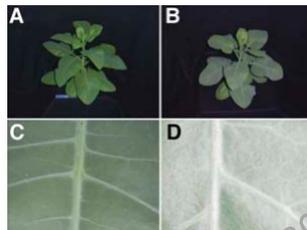


Figure 1. Tree tobacco plants exposed to periodic drying events. A, Well-watered plant. B, Plant exposed to three periodic drying events. C, Close-up of the adaxial surface of a fully expanded leaf from the plant in A. D, Close-up of the adaxial surface of a fully expanded leaf from the plant in B.

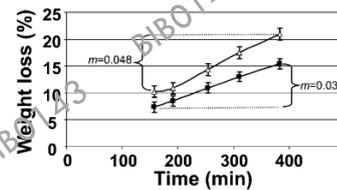
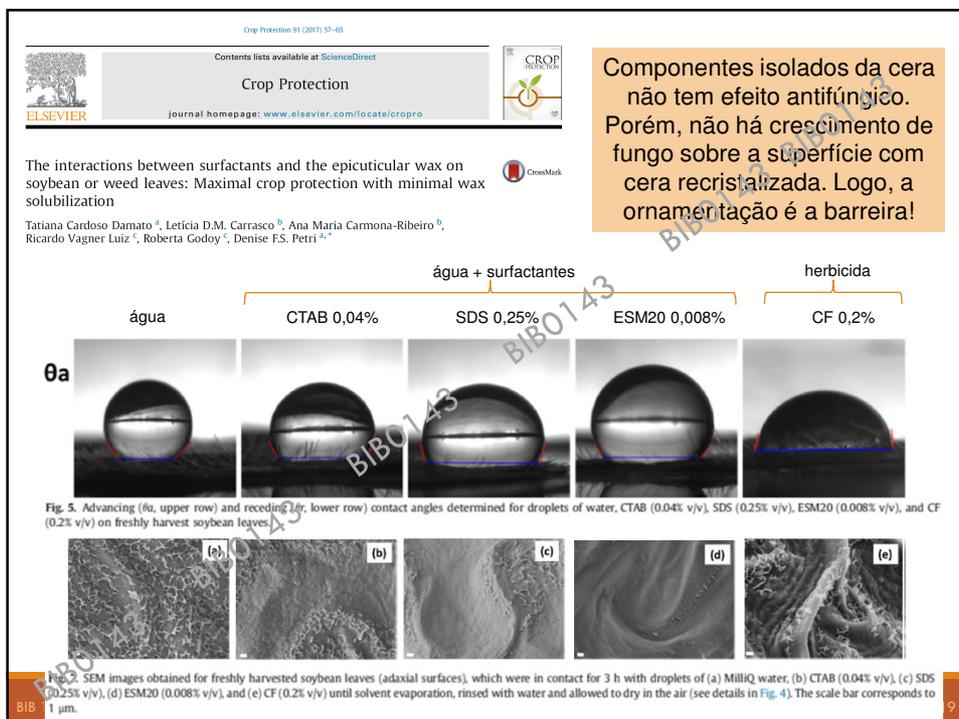
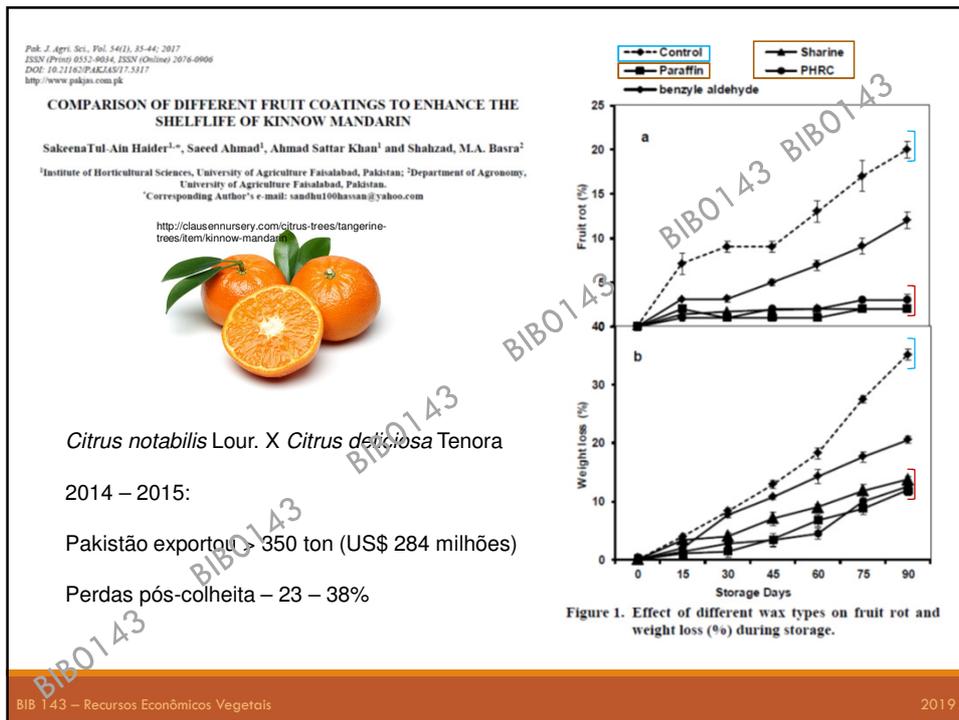


Figure 5. Rate of actual weight loss due to epidermal conductance from leaves excised from periodically dried (■) or well-watered (□) plants. Data represent one of three replicated experiments. Three leaves per plant from each of four well-watered and periodically dried plants were excised and immediately placed in a 30°C incubator. Leaves were weighed after excision at approximately 150 min, 180 min, and at hourly increments thereafter. The percent of water loss was determined relative to the original leaf weight. Error bars indicate the mean SD across all time points.  $m = 0.048$  and  $m = 0.035$  are the slopes of the lines.

Aumento na quantidade de cera nas plantas submetidas a seca

Aumento na tolerância a seca relacionado ao aumento de cera total



### Efeito Lotus

Quanto menor o ângulo de contato da gota com a superfície, maior a possibilidade dessa superfície ficar molhada; relacionado com a impermeabilidade

<http://www.olharnano.com/artigos/4001/63001/O-efeito-L%C3%B3tus->

Super-hidrofílico    Hidrofílico    Intermediário

Hidrofóbico    Super-hidrofóbico

<https://www.youtube.com/watch?v=sCjmyQIHYM>

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Chemosphere 76 (2009) 1445–1450

Contents lists available at ScienceDirect

**Chemosphere**

journal homepage: [www.elsevier.com/locate/chemosphere](http://www.elsevier.com/locate/chemosphere)

**Anthocyanins and tannins in ozone-fumigated guava trees**

Fernanda Mendes de Rezende<sup>a</sup>, Cláudia Maria Furlan<sup>b,\*</sup>

**Table 1**  
Mean percentages (± standard deviation) of anthocyanins and tannins on *Psidium guajava* 'Paluma' after 30 days of exposure to different fumigation treatments: CF – charcoal-filtered air; NF – ambient non-filtered air; NF + O<sub>3</sub> – ambient non-filtered air plus 40 ppb of O<sub>3</sub>; n = 10; Values followed by # correspond to statistically different means when comparing to CF (p < 0.05).

Treatment	Anthocyanins	Tannins	
		Total	Condensed
CF	0.970 ± 0.4	0.136 ± 0.016	0.035 ± 0.006
NF	1.295 ± 0.7	0.160 ± 0.017 <sup>#</sup>	0.051 ± 0.009 <sup>#</sup>
NF + O <sub>3</sub>	1.607 ± 0.6 <sup>#</sup>	0.162 ± 0.014 <sup>#</sup>	0.046 ± 0.006 <sup>#</sup>

**Correlação entre porcentagem de injúrias foliares e teor de antocianinas.**

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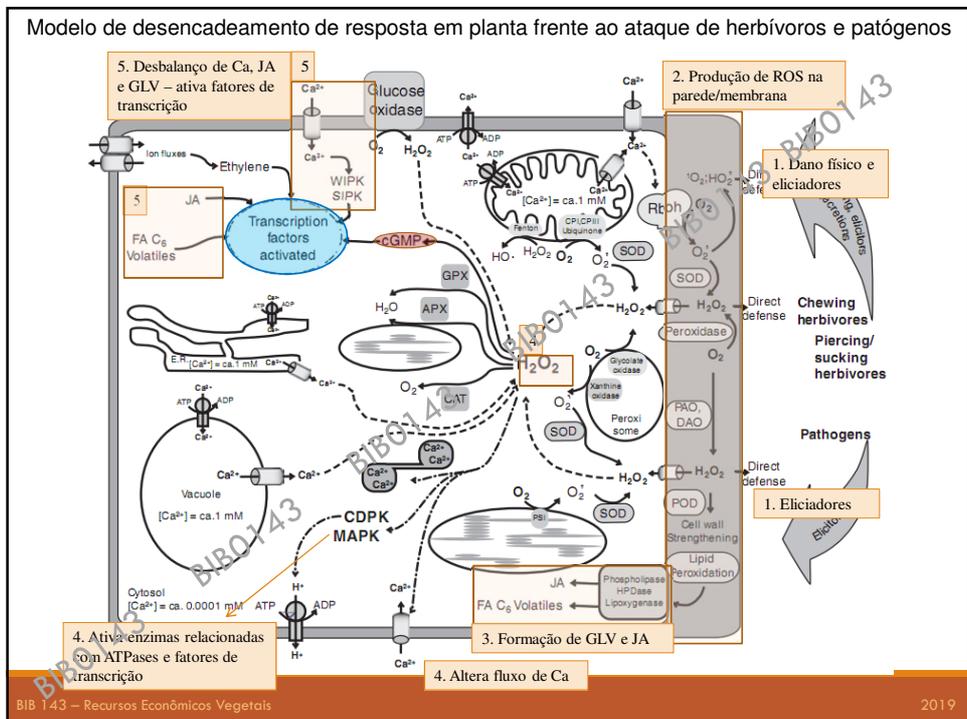
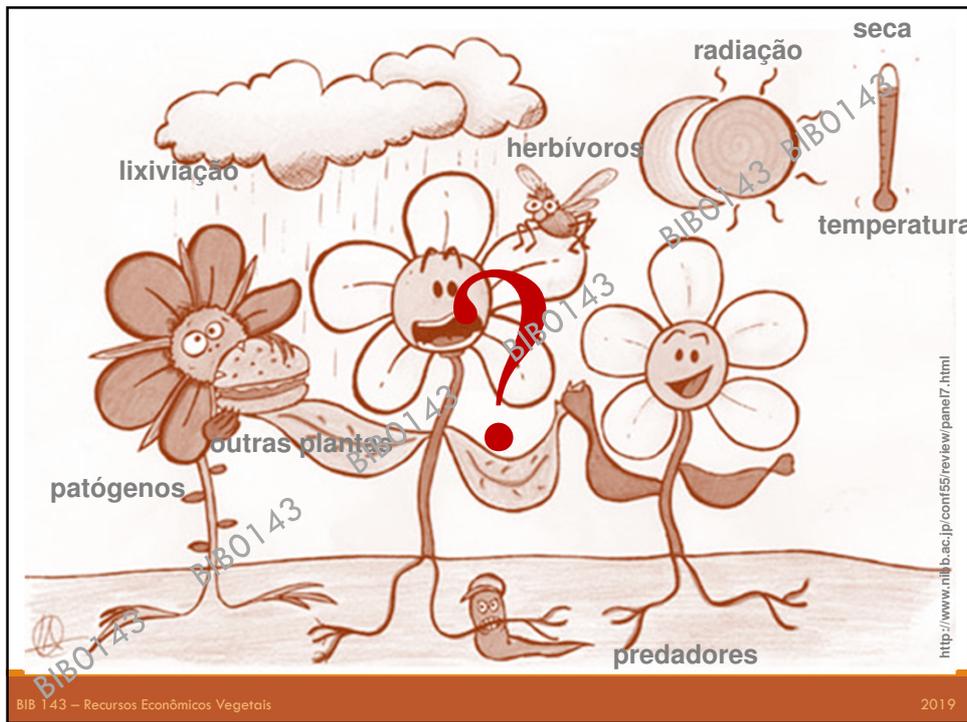
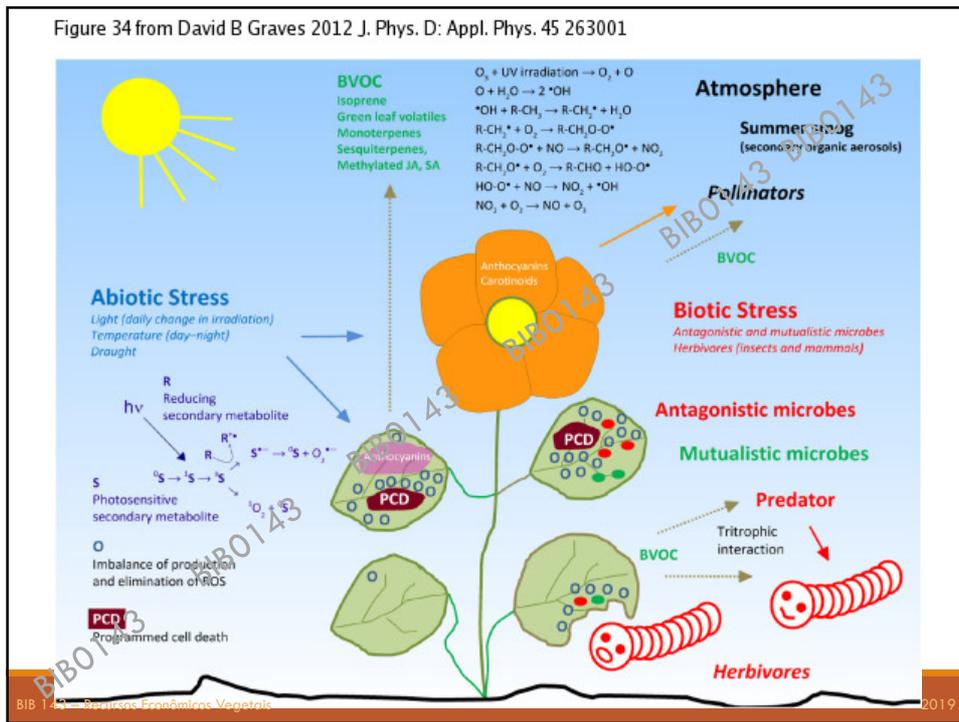


Figure 34 from David B Graves 2012 J. Phys. D: Appl. Phys. 45 263001



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Signaling events in plants: Stress factors in combination change the picture

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**A. Control condition**

Nutrients Energy → Biomass → Reproduction

↑ Defense

**B. Stress condition**

Nutrients Energy → Biomass → Reproduction

↑ Defense

Fig. 1. Energy channelling is shifted from biomass production to defense programs. Under stress conditions nutrients and energy are redirected from biomass production towards defensive processes. Additionally reproduction is accelerated under adverse environmental conditions.

Stresses	Signals	Transduction	Responses	Adaptation
Heat	ABA	MAPK	Translation	Defense Growth Reproduction
Drought	JA	CPK	Ion transport	
Ozone	SA	SnRK3	Metabolism	
Salinity	Ethylene	SnRK2	Transcription	
Light	ROS	SnRK1	MicroRNAs	
Herbivores	Ca <sup>2+</sup>	TOR-1	Chromatin	
Necrotrophs	Lipids			
	Sugars			
	Amino acids			
Biotrophs				

Fig. 2. Crucial events in the signal transduction pathway activated by several biotic and abiotic stress factors. The schematic diagram shows how different stress factors activate different signals, which themselves trigger signal transduction cascades. Different signalling events result in appropriate plant responses leading to adaptation processes including defense, growth and reproduction. ABA (abscisic acid); ROS (reactive oxygen species); JA (jasmonic acid); SA (salicylic acid); Ca<sup>2+</sup> (Calcium); MAPK (mitogen-activated protein kinase); CPK (Ca<sup>2+</sup>-dependent protein kinase); SnRK (sucrose non-fermenting-1-related protein kinase); TOR-1 (target of rapamycin).

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