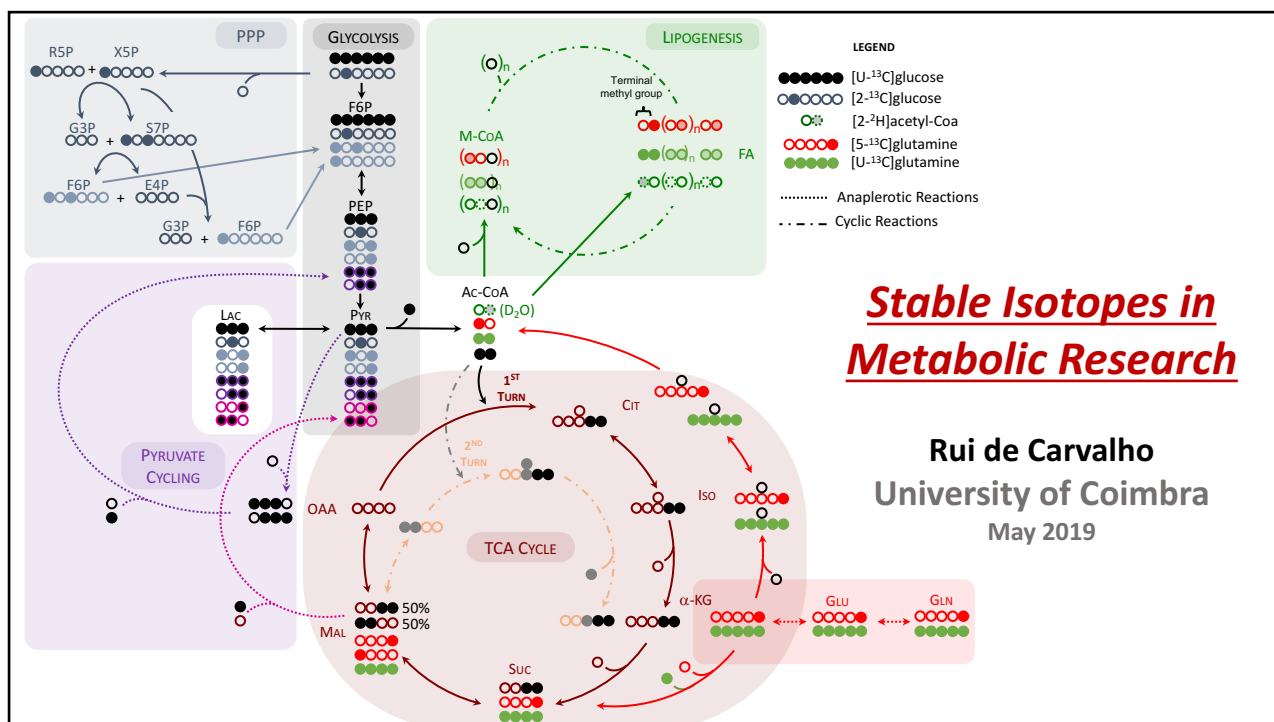




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RESEARCH ARTICLE

Metabolic remodeling triggered by salivation and diabetes in major salivary glands

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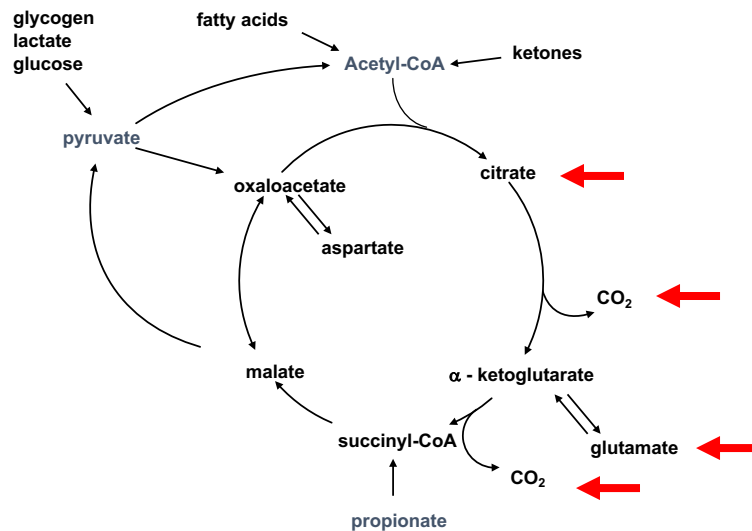
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The metabolic profile of major salivary glands was evaluated by ¹³C nuclear magnetic resonance isotopomer analysis (¹³C NMR-IA) following the infusion of [U-¹³C]glucose in order to define the true metabolic character of submandibular (SM) and parotid (PA) glands at rest and during salivary stimulation, and to determine the metabolic remodeling driven by diabetes. In healthy conditions, the SM gland is characterized at rest by a glycolytic metabolic profile and extensive pyruvate cycling. On the contrary, the PA gland, although also dominated by glycolysis, also possesses significant Krebs' cycle activity and does not sustain extensive pyruvate cycling. Under stimulation, both glands increase their glycolytic and Krebs' cycle fluxes, but the metabolic coupling between the two pathways is further compromised to account for the much increased biosynthetic anaplerotic fluxes. In diabetes, the responsiveness of the PA gland to a salivary stimulus is seriously hindered, mostly as a result of the incapacity to burst glycolytic activity and also an inability to improve the Krebs' cycle flux to compensate. The Krebs' cycle activity in the SM gland is also consistently compromised, but the glycolytic flux in this gland is more resilient. This diabetes-induced metabolic remodeling in SM and PA salivary glands illustrates the metabolic need to sustain adequate saliva production, and identifies glycolytic and oxidative pathways as key players in the metabolic dynamism of salivary glands.

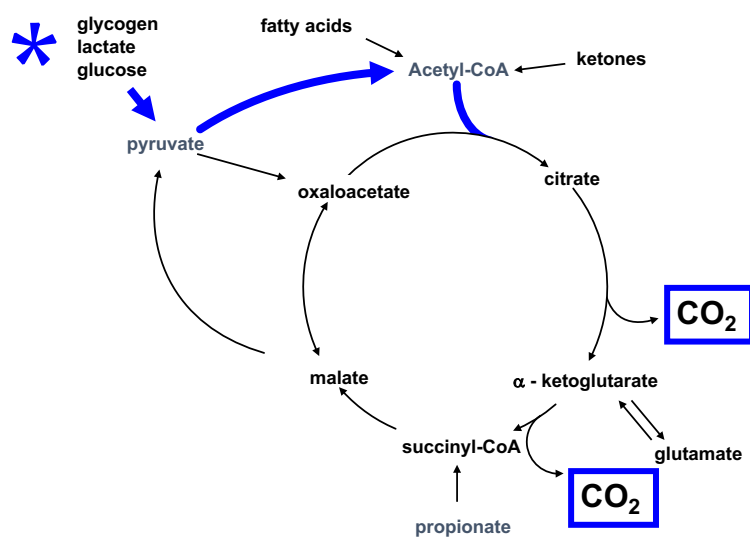
KEYWORDS
glycolysis, isotopomer analysis, Krebs' cycle, metabolism, salivary glands, salivary stimulation

Stable Isotopes in Salivary Gland Metabolic Research

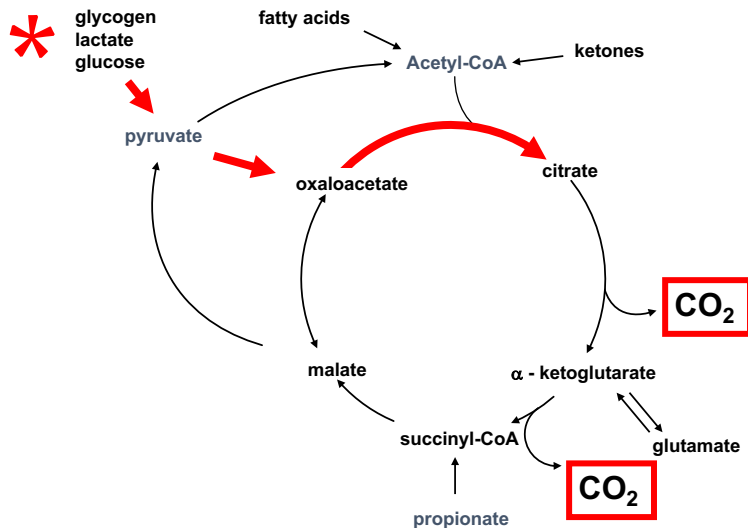
Tracers are Required to Measure Fluxes by Detecting Enrichment in Product Molecules



Analysis of Substrate Oxidation: CO_2 Release is Not Pathway - Specific



Analysis of Substrate Oxidation: CO₂ Release is Not Pathway - Specific



Properties of Carbon Isotopes

Nucleus	Abundance	Half-life	
¹¹ C	-	20 minutes	cyclotron & radiochemists
¹² C	98.9%	-	
¹³ C	1.1%	-	catalogue & lab sink
¹⁴ C	-	5730 years	catalogue, paperwork & radiation containment

¹¹C, ¹³C and ¹⁴C may all be used as tracers

Carbon-13: Natural Abundance

- ✓ Natural Abundance of ^{13}C $\sim 1.1\%$
- ✓ $\gamma^{13}\text{C} \sim 1/4 \gamma^1\text{H}$
- ✓ Low Sensitivity $\Rightarrow \uparrow[\text{sample}]$
- ✓ Consecutively Labeled ^{13}C Nuclei $\leq 0.011 \cdot 0.011$
($\sim 1:8300$)

Carbon-13: Enriched Metabolites

- ✓ Abundance of ^{13}C $> 1.1\%$
- ✓ Consecutively Labelled ^{13}C Nuclei $\Rightarrow {}^1J_{\text{CC}}$ Couplings
- ✓ Multiplets in Each Carbon Resonance



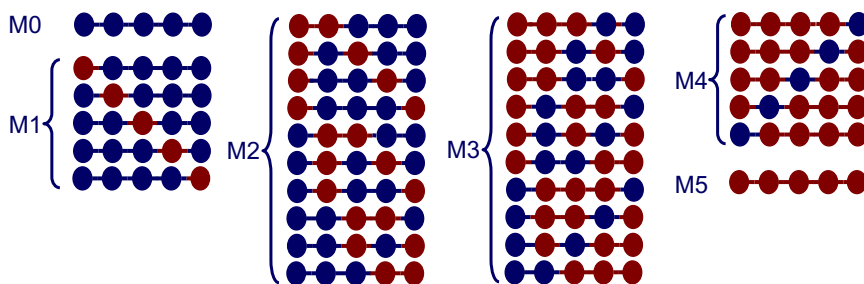
Isotopomer Information

Glutamate Isotopomers

✓ Isotopomer $\Leftrightarrow$ Isotope + Isomer

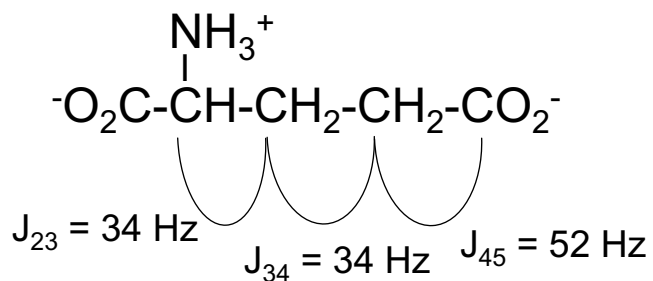
✓ Metabolite with n Carbons has 2^n Isotopomers

- glutamate $\rightarrow 2^5 = 32$ Isotopomers



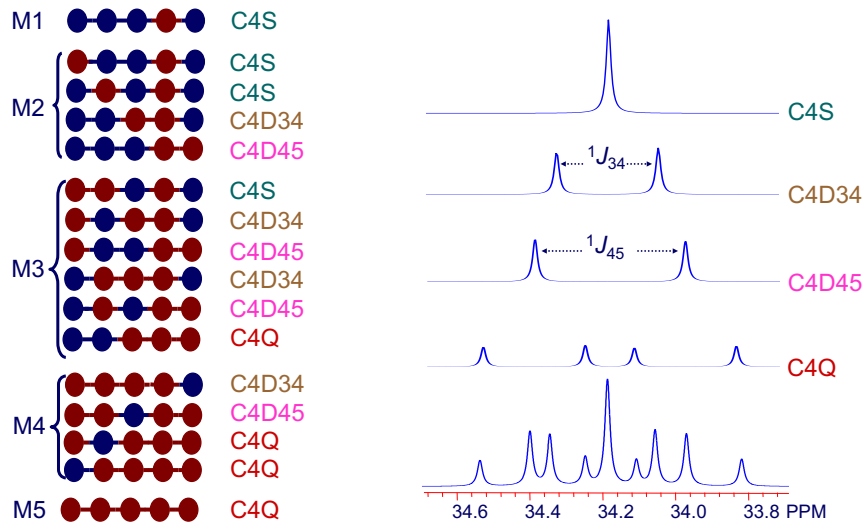
The Overwhelming Advantage of ^{13}C NMR for Analysis of Substrate Oxidation

Spin - Spin Coupling

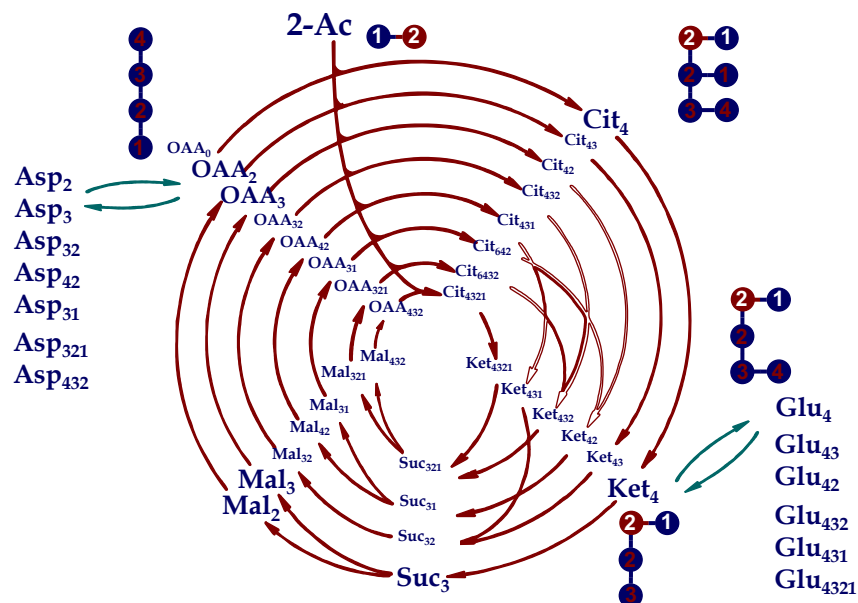


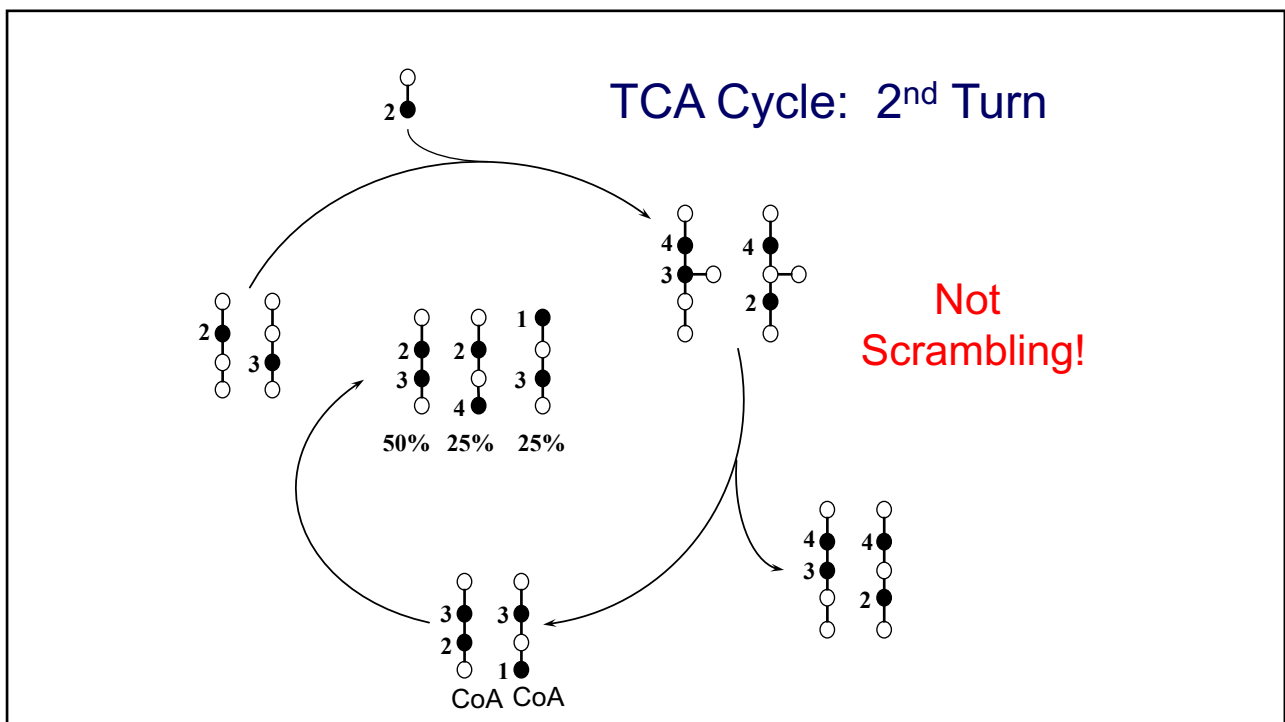
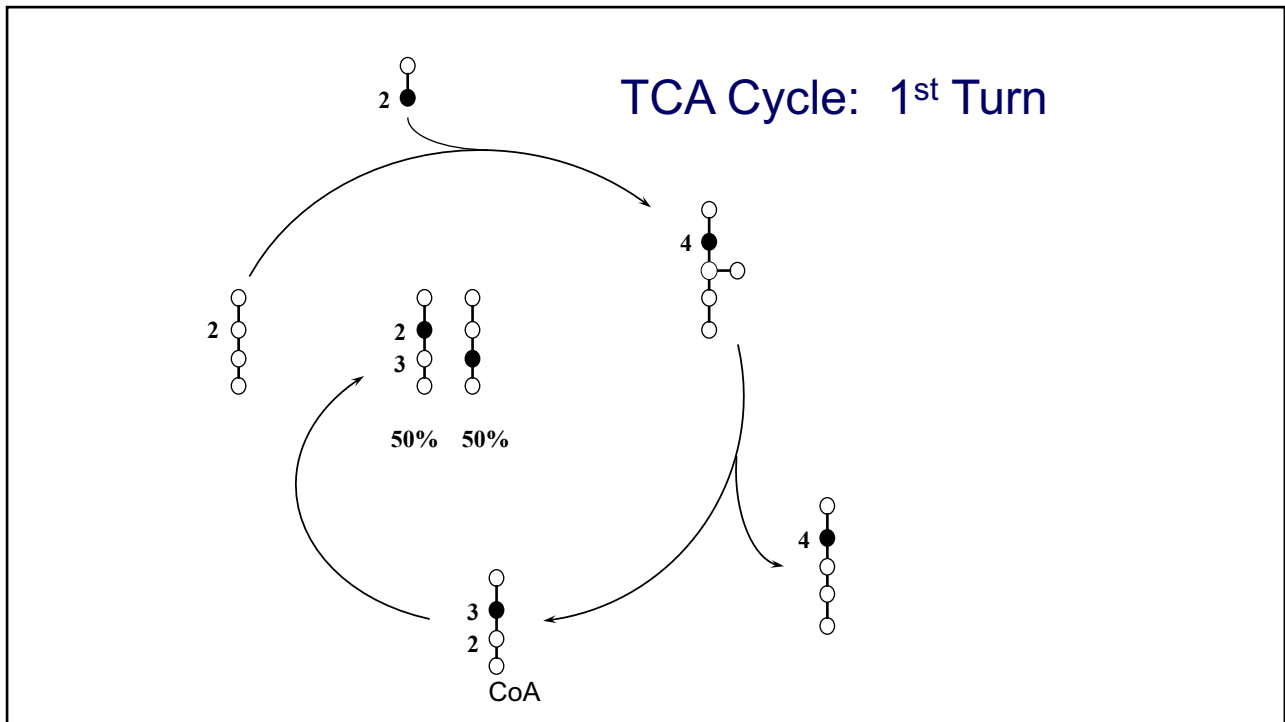
Distinction of Glutamate ^{13}C Isotopomers

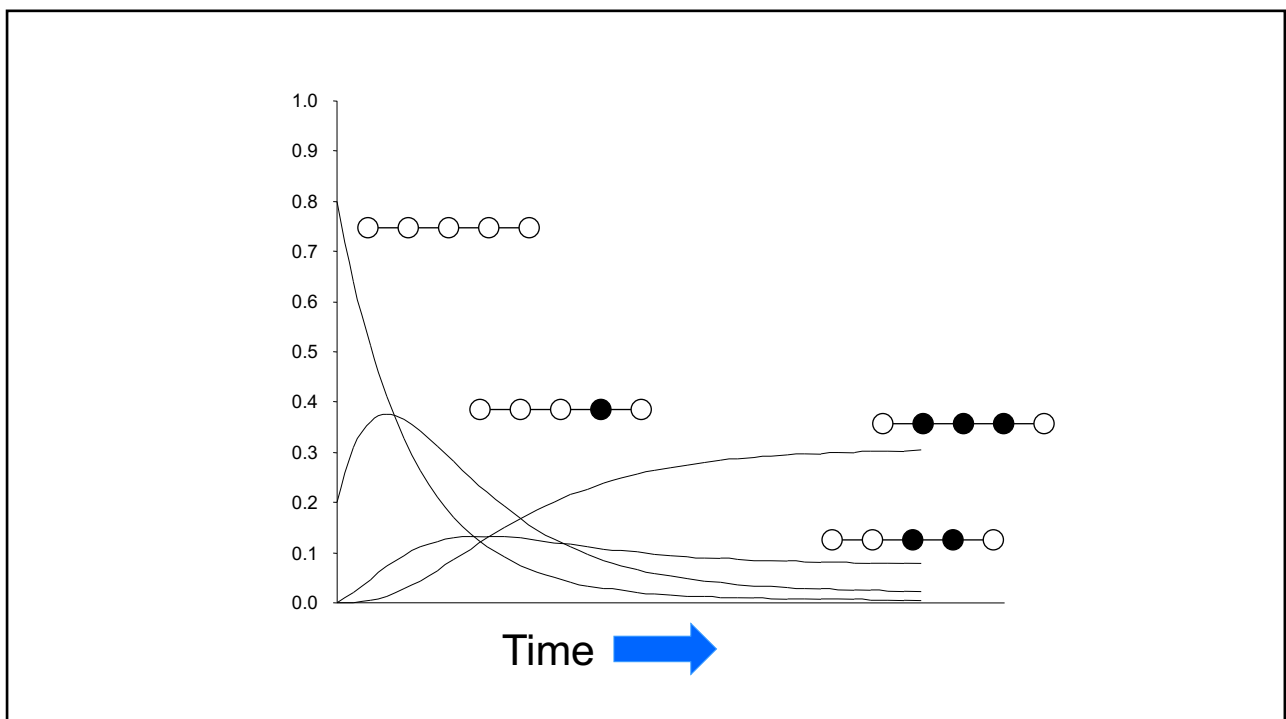
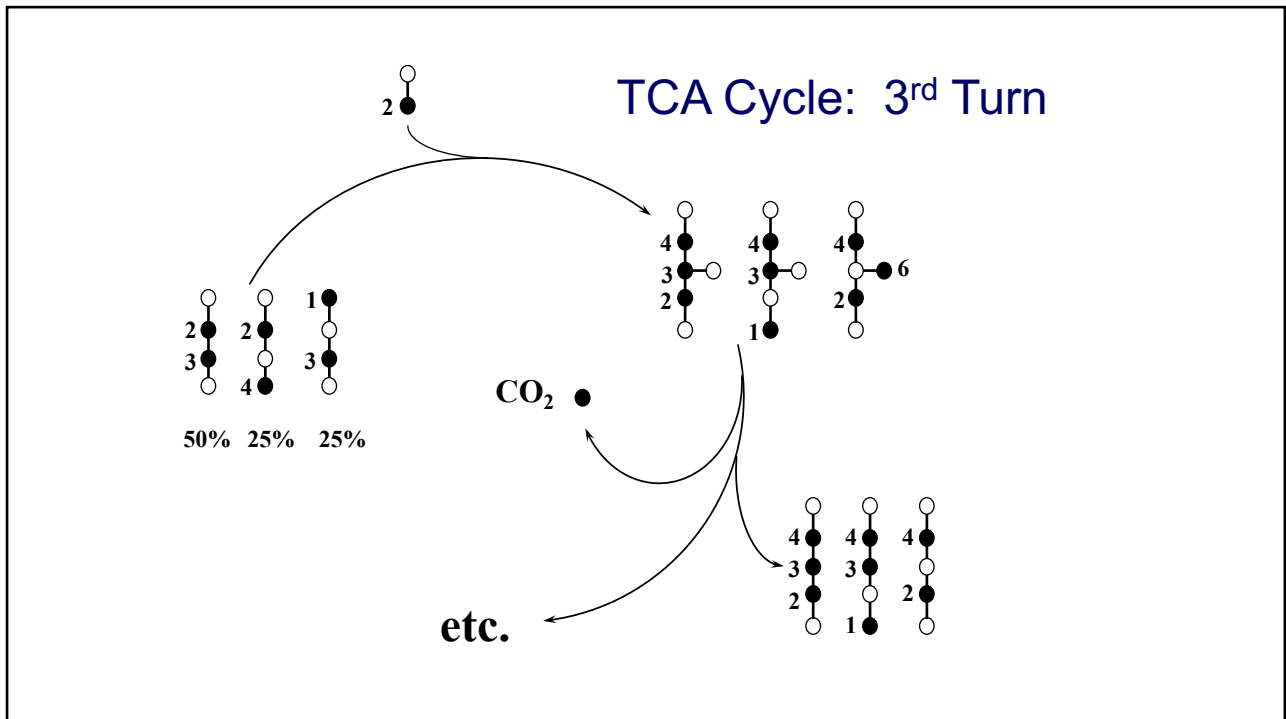
✓ Glutamate C4 Resonance $\Rightarrow$ Labeling in C4

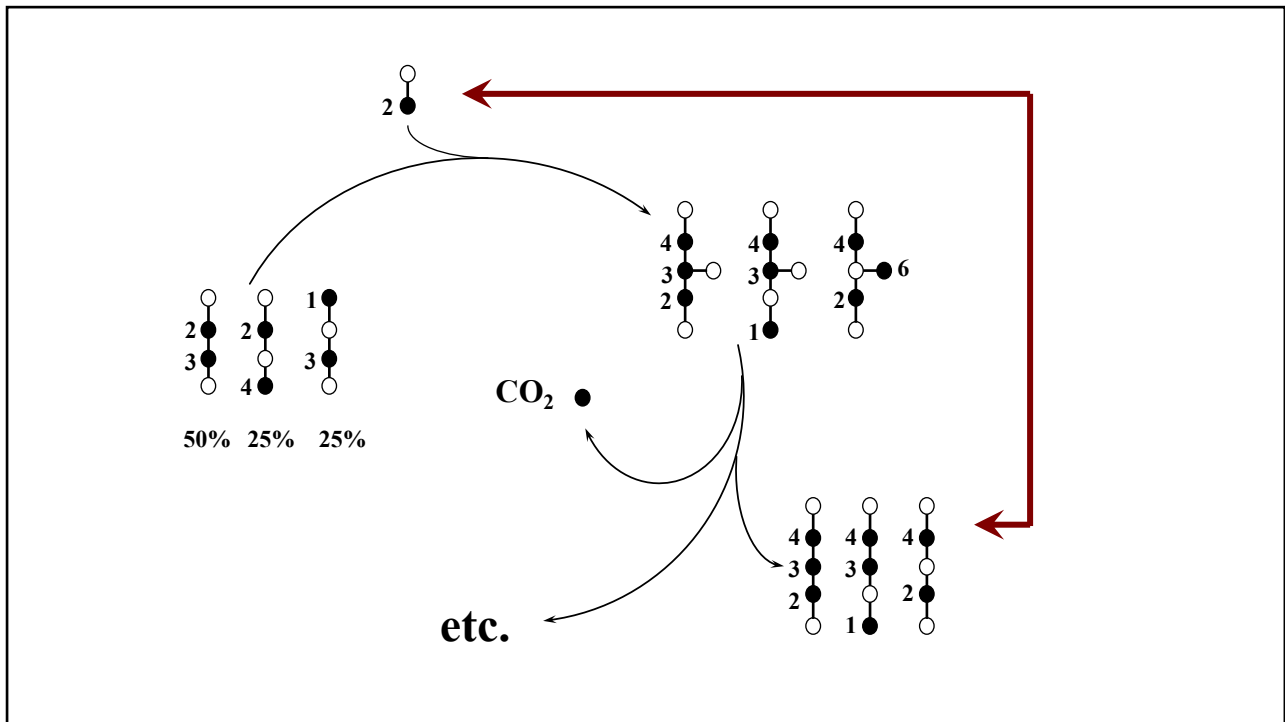


^{13}C Enrichment of Metabolites



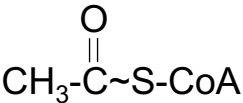


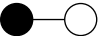
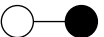
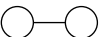




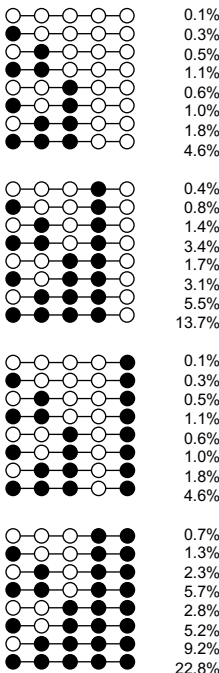
At steady state, a heart oxidizing:

[U- ¹³ C]LCFA	50%
[3- ¹³ C]LP	30%
[1,3- ¹³ C]KB	10%
¹² C-Glucose	10%



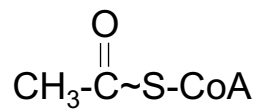
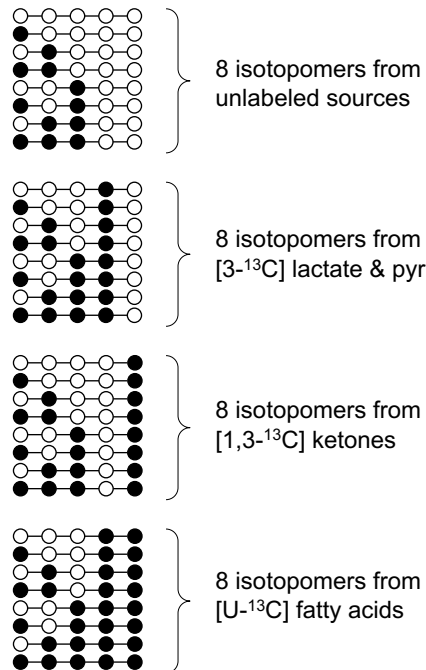
	0.5
	0.3
	0.1
	0.1



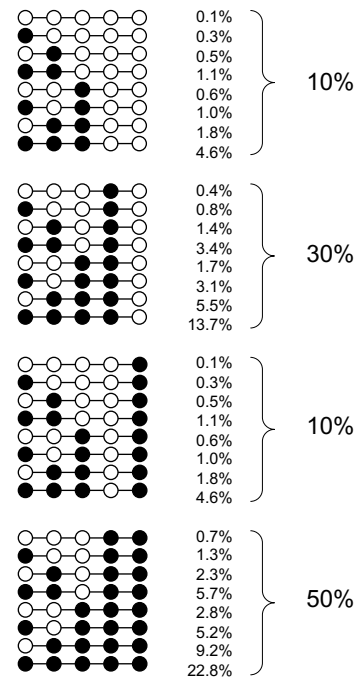
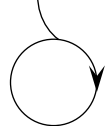


This metabolic situation
generates 32 isotopomers

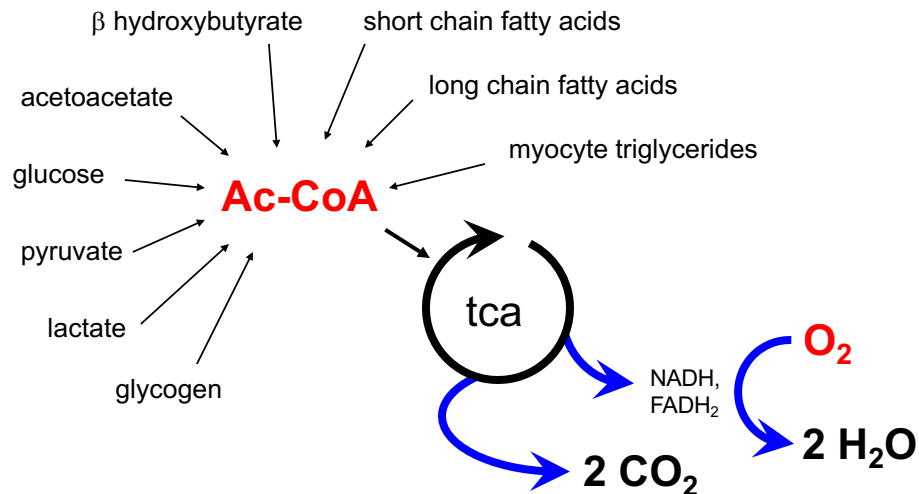
Since Ac-CoA label
corresponds to glutamate
C4 and C5 enrichment, it
is useful to cluster 4
groups of isotopomers
based on label in positions
4 & 5.



- 0.5 (LCFA)
- 0.3 (Lac/pyr)
- 0.1 (ketones)
- 0.1 (no label)



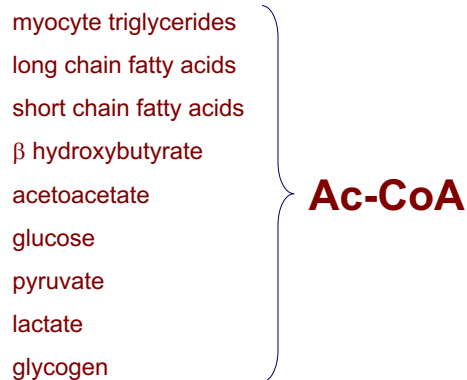
Cardiac metabolism transfers energy from many sources to the contractile apparatus

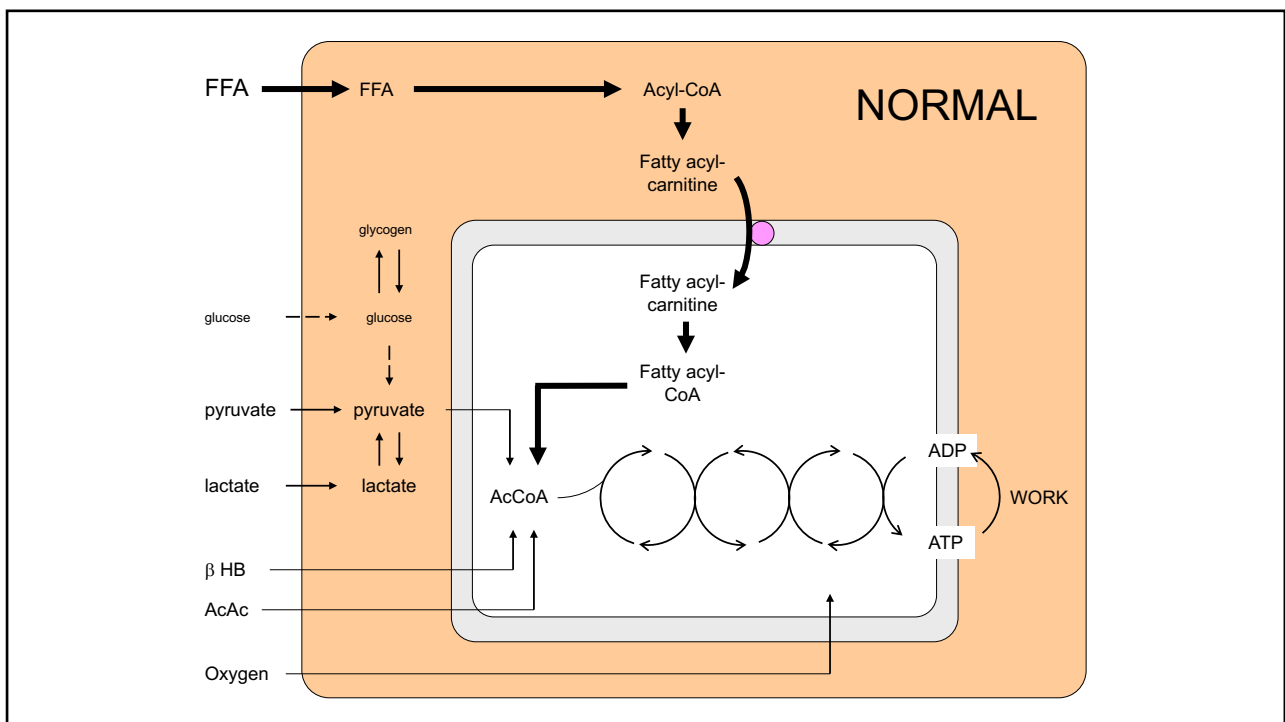
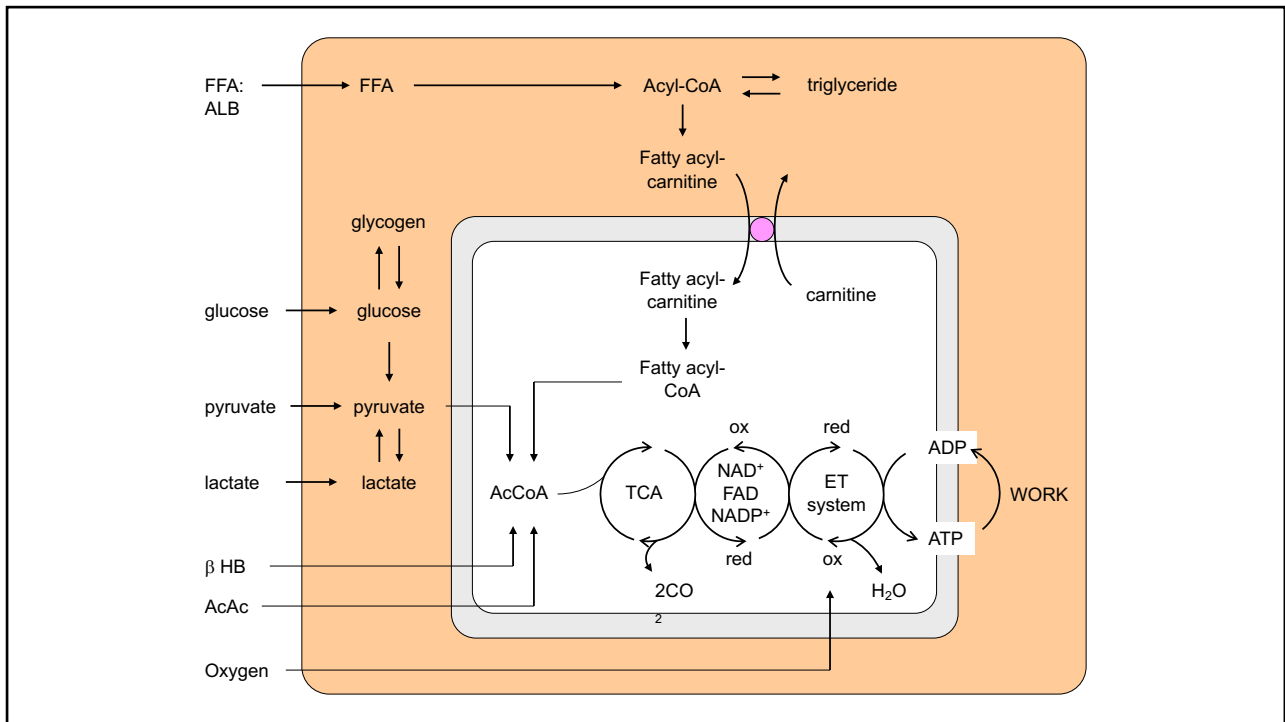


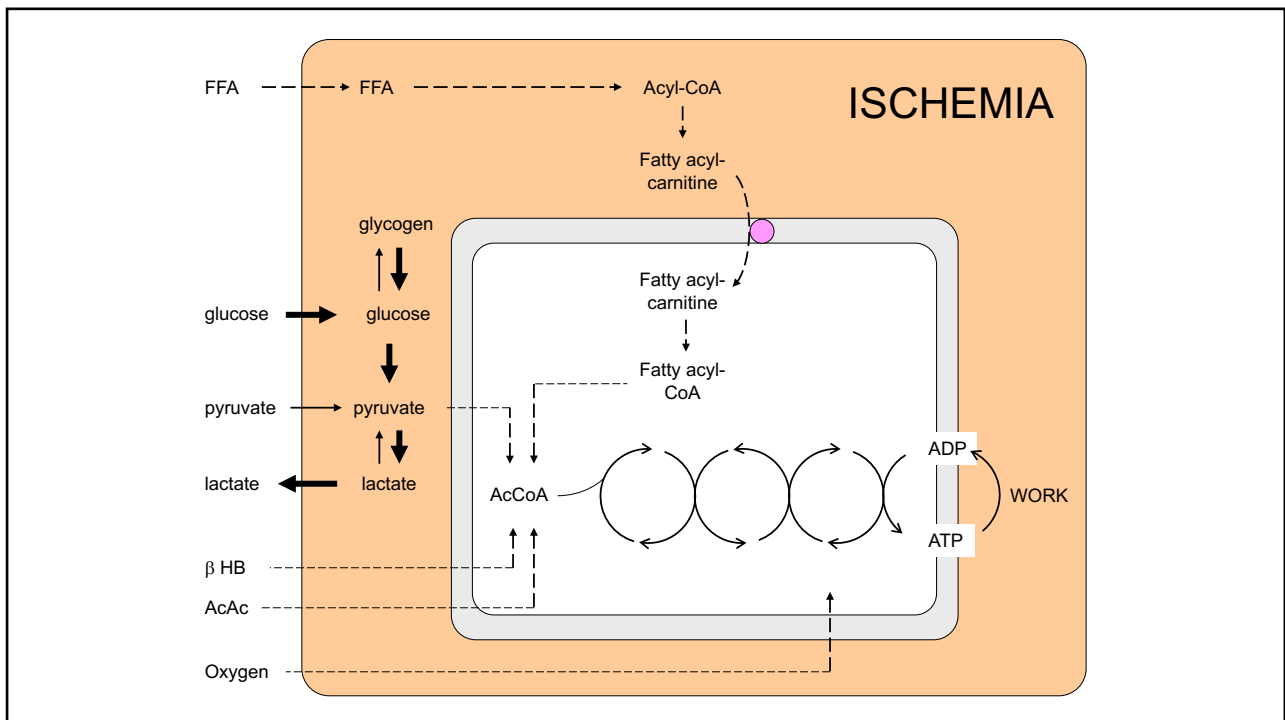
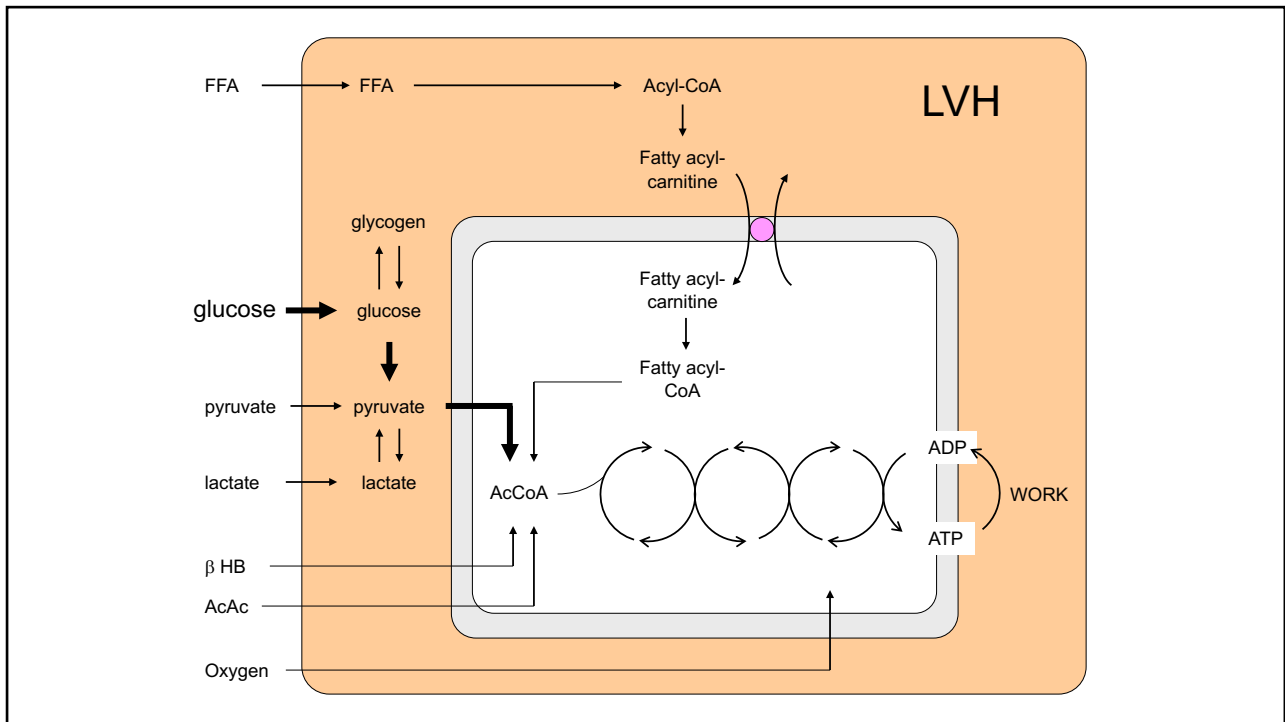
The enzymatic machinery to oxidize is available to oxidize each substrate.

By definition, sources of Ac-CoA in vivo are complementary:

$$F_{\text{trig}} + F_{\text{LCFA}} + F_{\text{glucose}} + \dots = 1$$







Conditions Associated with Altered FA Oxidation or Altered Gene Expression

High fatty acid oxidation

- Transition from fetal environment
- Fasting
- Diabetes
- Exercise

High carbohydrate oxidation

- Fetal environment
- Hypertrophy
- Myocardial ischemia
- Drugs

Why is it important to measure substrate oxidation in the heart?

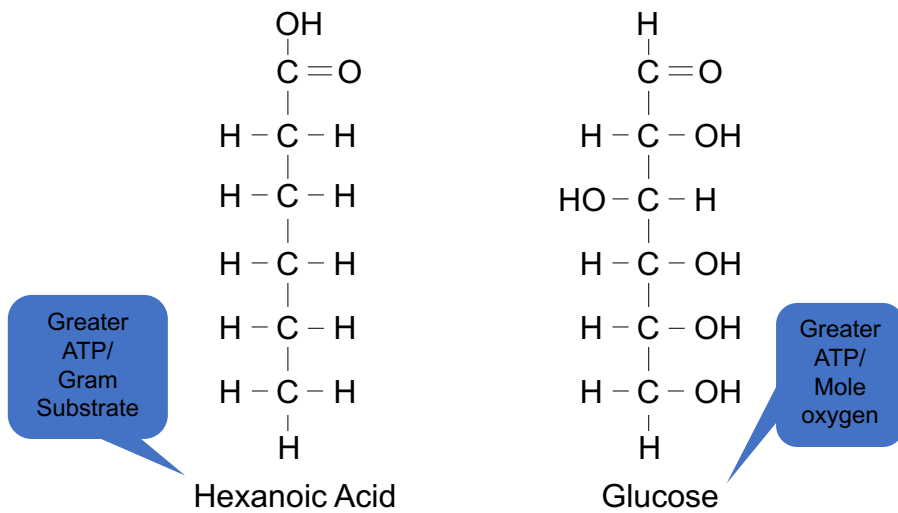
Many diseases or physiological states are associated with altered substrate oxidation or metabolism, which implies:

Understanding metabolism provides insight into pathogenesis

Measurement of metabolism yields diagnostic information

Therapeutic control of metabolism may improve clinical outcome

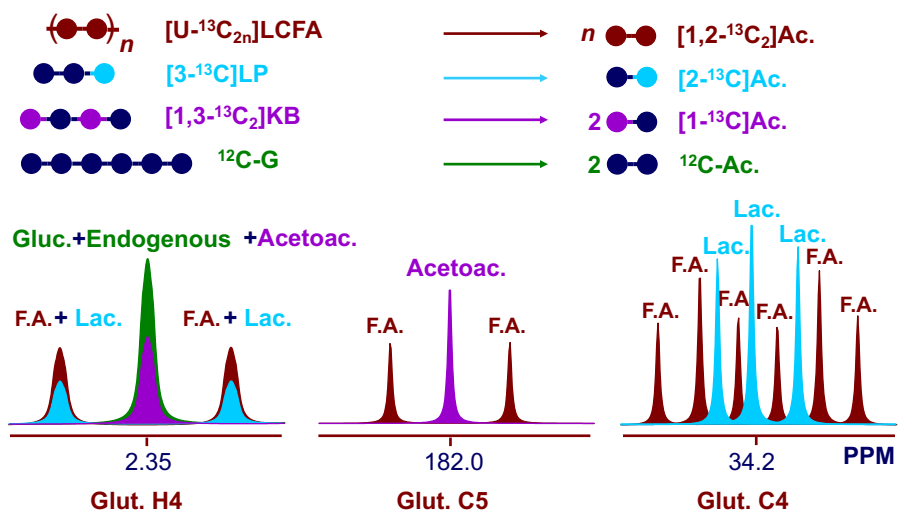
- Improve ATP yield / mole O₂ – critical for ischemic syndromes
- Reduce TG storage
- Reduce rhythm disturbances



FAs are more reduced (contain more chemical energy) than glucose

FA oxidation requires activation to acyl-CoA which consumes ATP

Substrate Competition by ^{13}C -NMR



Acetyl-CoA Enrichments (F_{ci})

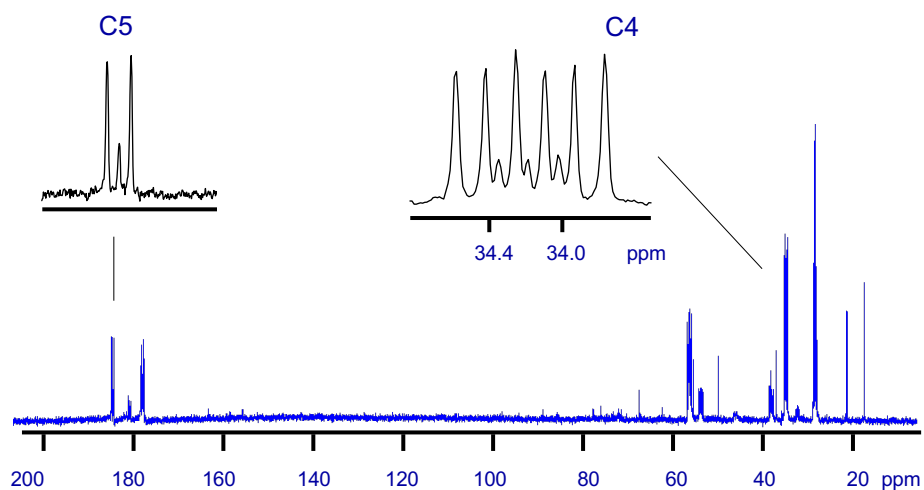
✓ $F_{C3} = \%[U-^{13}C_n]LCFA = C4Q \cdot (C4F/C3F)$

✓ $F_{C2} = \%[3-^{13}C]LP = C4D34 \cdot (C4F/C3F)$

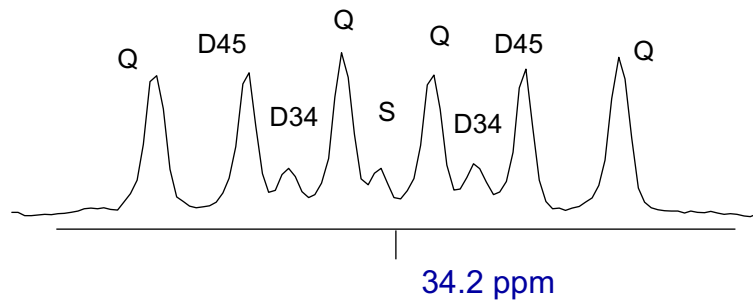
✓ $F_{C1} = \%[1,3-^{13}C_2]KB = (C5S/C5D) \cdot F_{C3}$

✓ $F_{C0} = \%Glucose + Endogenous = 1 - (F_{C3} + F_{C2} + F_{C1})$

Analysis of a ^{13}C NMR Spectrum from a Rat Heart Extract Supplied with $[U-^{13}C]$ LCFA, $[1,3-^{13}C]$ ketones and $[3-^{13}C]$ lactate/pyruvate

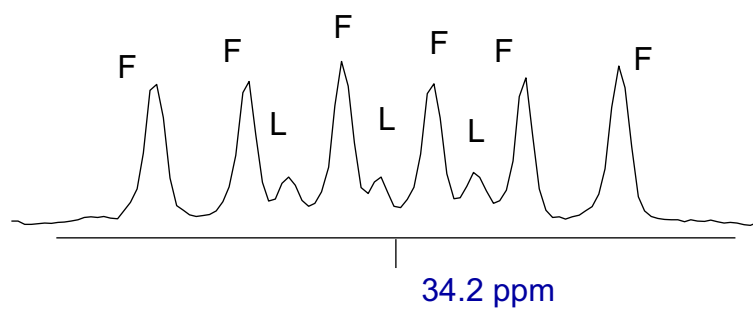


Analysis of C4



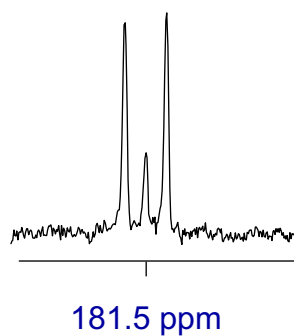
$$\frac{Q + D45}{S + D34} = \frac{62 + 29}{3 + 6} = \frac{\bullet\bullet}{\circ\bullet} = 10$$

Analysis of C4



$$\frac{\text{LCFA oxidation}}{\text{Lactate oxidation}} = 10$$

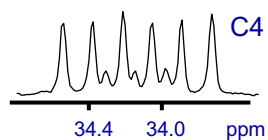
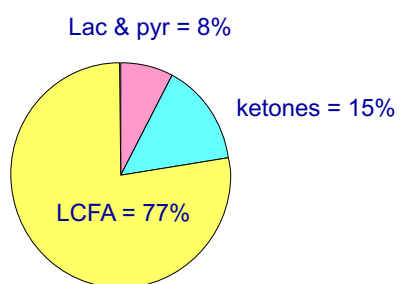
Analysis of C5



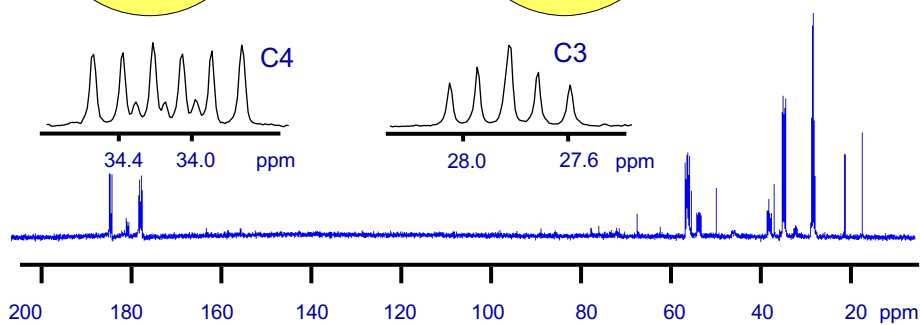
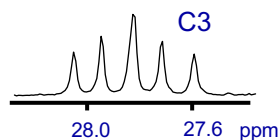
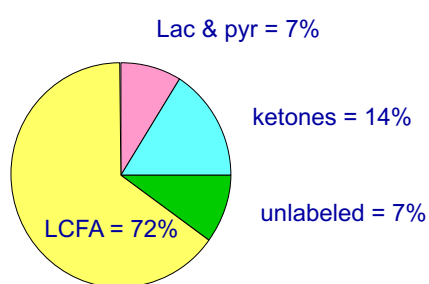
$$\frac{C5D45}{C5S} = \frac{84}{16} = \frac{\bullet\bullet}{\bullet\circ} = 5.2$$

$$\frac{LCFA}{AcAc} = 5.2$$

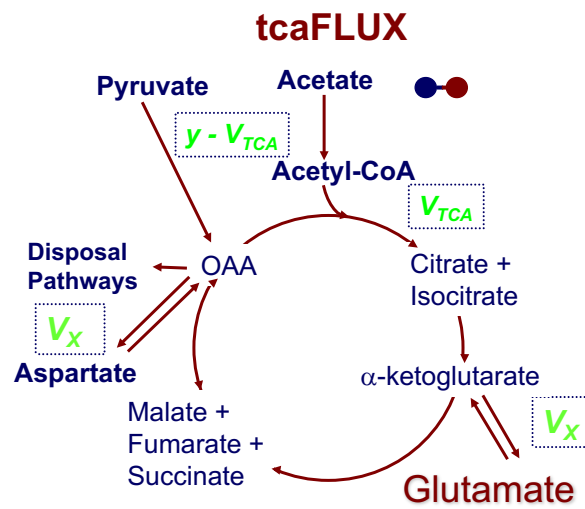
Analysis of C4 & C5:



Analysis of C3, C4 & C5:

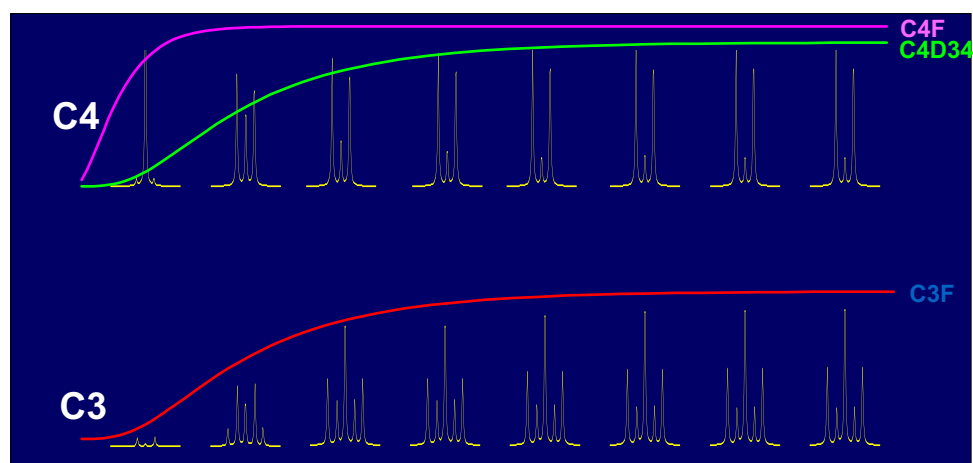


Kinetic Model of the Krebs Cycle

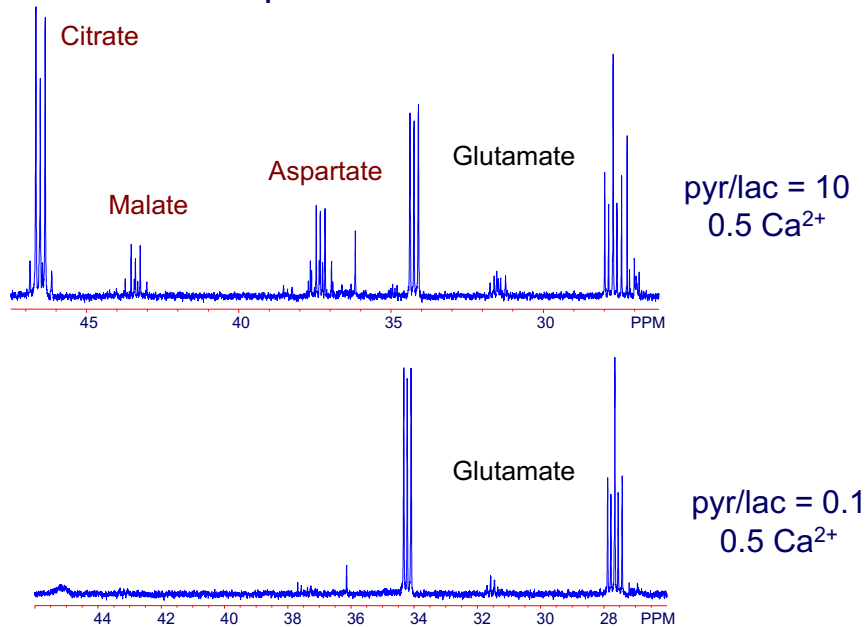


TCA cycle kinetics

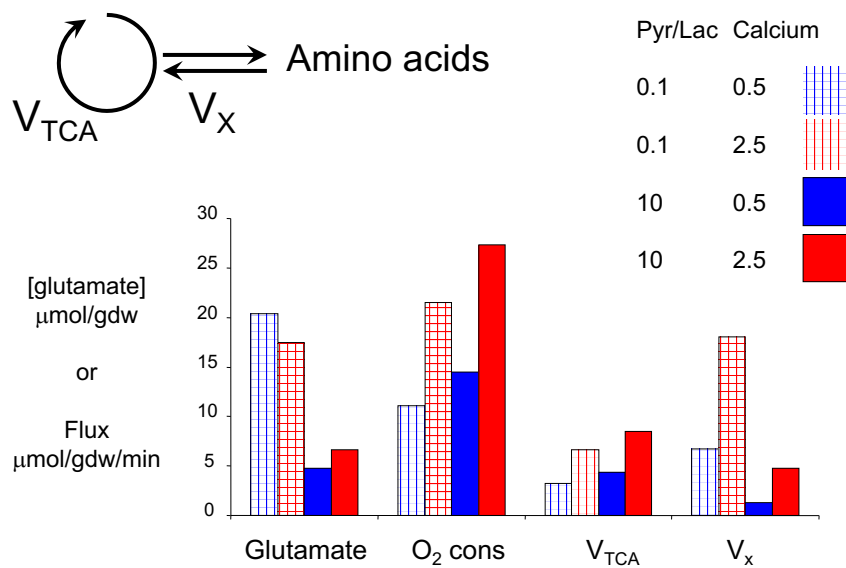
for a singly enriched substrate, $F_{C2} = 90\%$



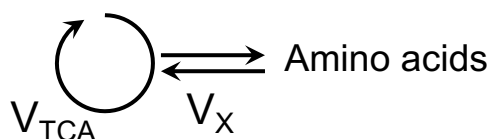
Metabolite pools are sensitive to cell redox



¹³C Isotopomer Analysis of TCA Cycle Kinetics: Redox & Calcium

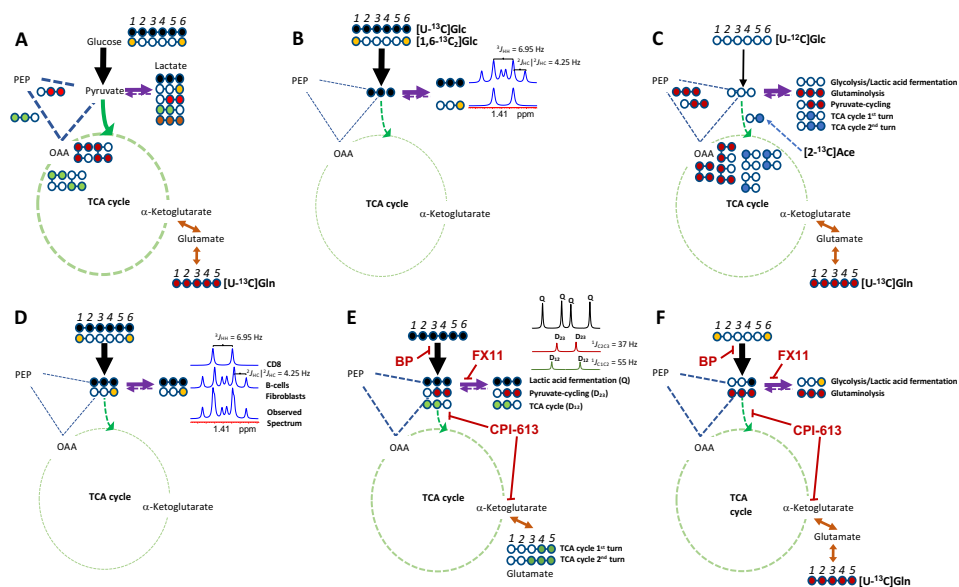


¹³C Isotopomer Analysis of TCA Cycle Kinetics: Redox & Calcium

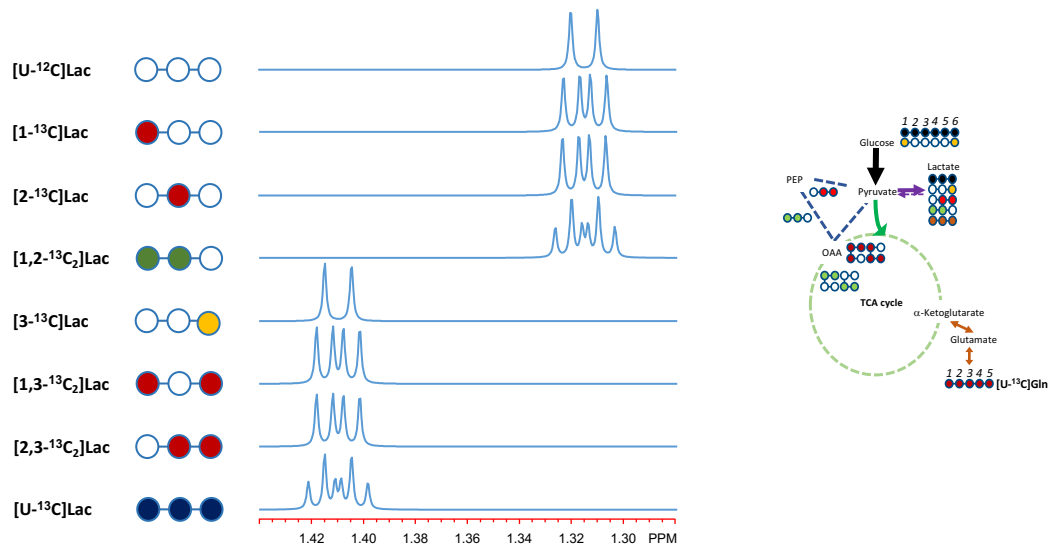


	Pyr/Lac	Calcium	V_X / V_{TCA}
	0.1	0.5	2.0
	0.1	2.5	2.7
	10	0.5	0.3
	10	2.5	0.6

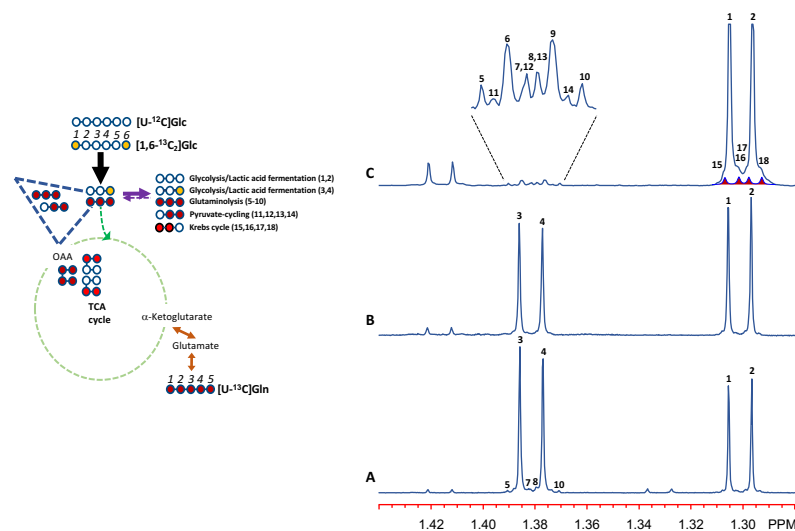
¹³C Isotopomer Analysis of Intermediary Metabolism



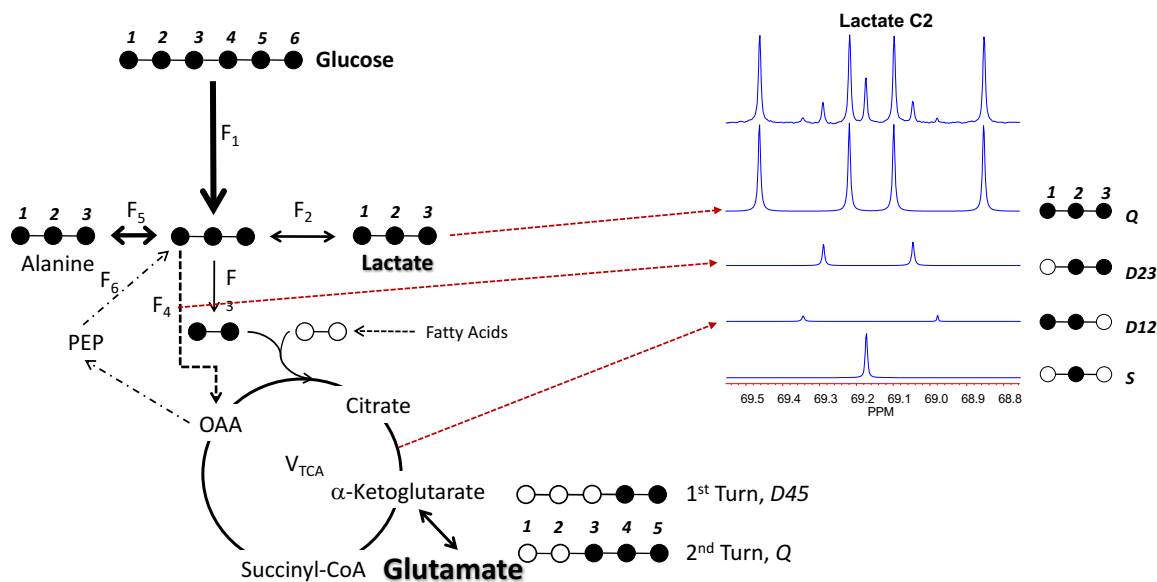
^{13}C Isotopomer Analysis of Lactate



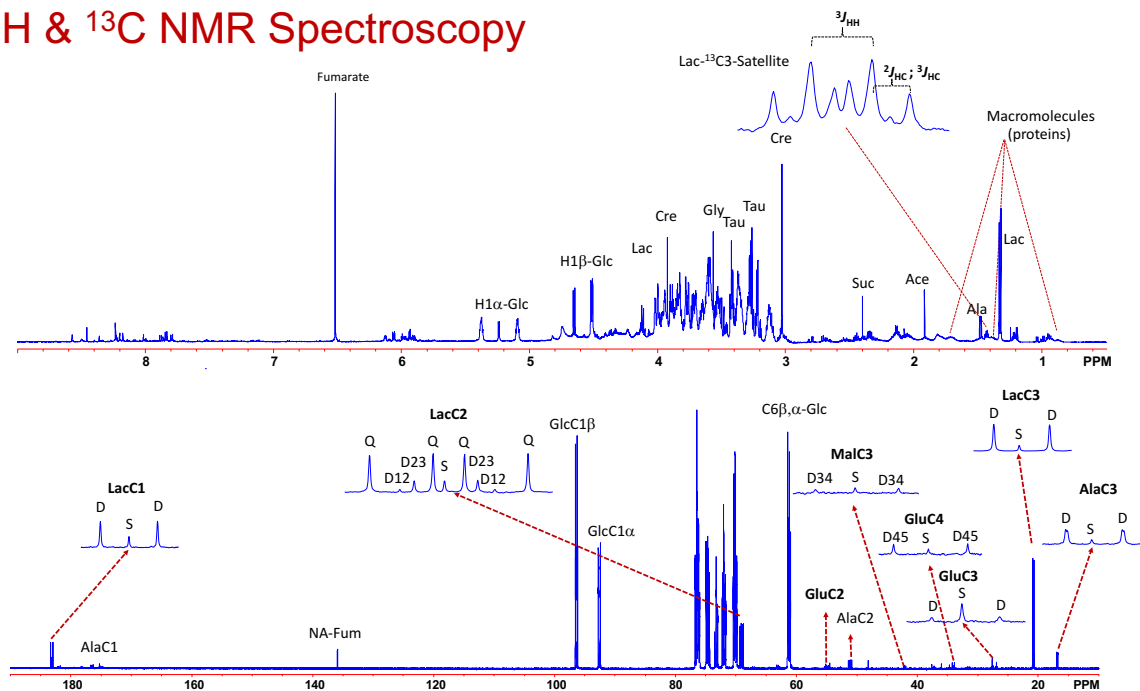
^{13}C Isotopomer Analysis: Glycolysis vs Glutaminolysis



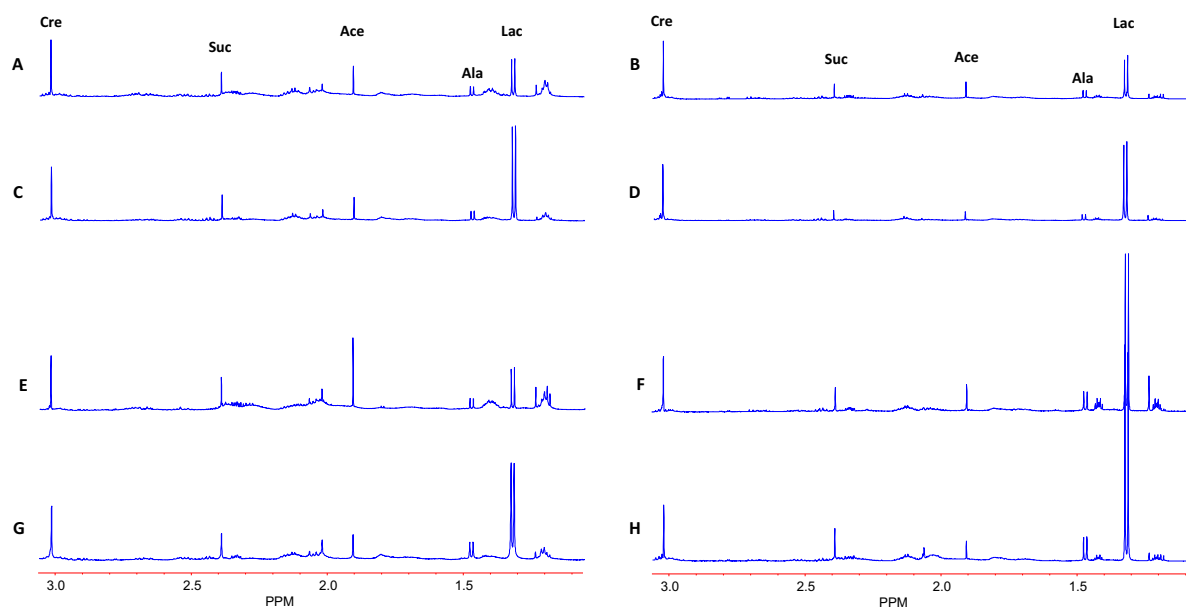
^{13}C Isotopomer Analysis: Salivary Glands



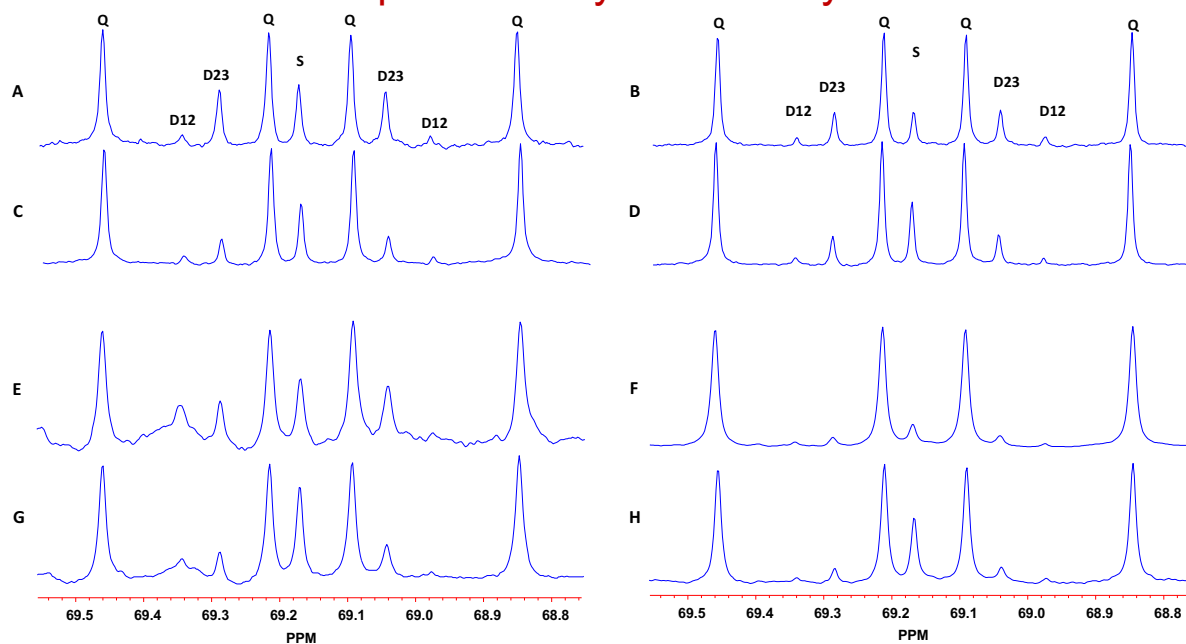
^1H & ^{13}C NMR Spectroscopy



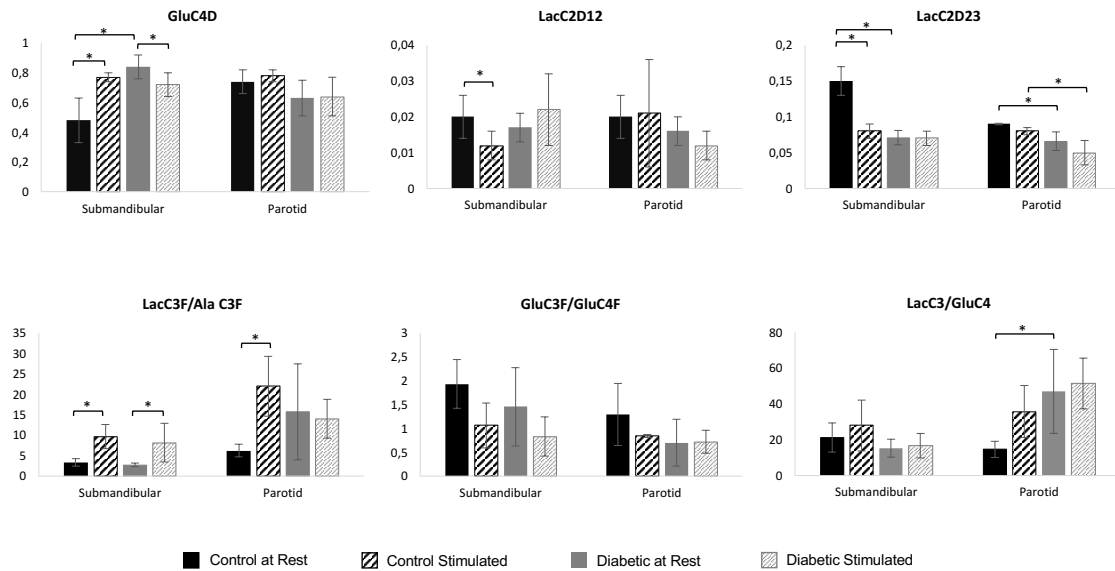
^1H NMR Spectroscopy: Levels of Intermediary Metabolites



^{13}C Isotopomer Analysis: Salivary Glands



^{13}C Isotopomer Analysis: Salivary Glands



^{13}C Isotopomer Analysis: Salivary Glands

