More Experimental Design, Poisson Distribution

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Lecture VI RBP5793

- More Experimental Design
- Poisson Distribution

  Useful Reference:

  A First Course in the Design and Analysis of Experiments
  Gary Oehlert, University of Minnesota. Available at
  http://users.stat.umn.edu/ gary/book/fcdae.pdf

From previous lecture we saw the concepts of

treatment, experimental unit, and response variable.

Also mentioned, the concepts of randomization

and replication. Two types of replication

Biological and Technical. What is the difference ?

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and replication. Two types of replication

Biological and Technical. What is the difference ?

Which is better and why ?

Biological Replication means we have more

Independent Experimental Units assigned to the various

treatments. In a balanced design, we have the same

number of experimental units per treatment. If not,

design is unbalanced. If we have just one factor

(*eg.* growth hormone) unbalanced designs may not be

a problem. If we have two (or more) factors

(*eg.* growth hormone and diet) unbalanced designs

are much more complicated to analyse.

Imagine we have an animal model to test

the combined effects of growth hormone and diet

on the weight gain of mice after a six week period.

Two levels for each factor. Four treatments in total.

4 Mice are assigned **at random** to each treatment.

16 mice in total.

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16 mice in total. Is this design balanced ?

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Experimental Design X

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During the six period one mouse dies. Is the design balanced.

Must analyze like an unbalanced design !

In the previous design with 2 mice per cage

what are the observational units ?

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what are the observational units ? Suppose we

used this design in another study which requires taking

a muscle sample from each mouse. We have 16 mice.

Now imagine we take **2** samples from each mouse.

32 samples in total. Same as biological replication ?

If not, why not ?

In previous example, not all samples were independent !

Two samples from the same mouse receiving the same

treatment are not independent observations ! For true

biological replication need to increase the number of

independent experimental units for each treatment.

In this case no increase in the numner of independent

experimental units, even though the total number of

measurements has increased. Example of **pseudoreplication**.

To see why pseudo replication is dangerous:

Consider the same quantity measured twice but

independently (imagine two different technicians).

Results are *X* and *Y* but with some error.

We use the mean $= (\frac{X+Y}{2})$ but since *X* and *Y*

have some error the result is not certain. We decide

to specify a confidence interval as well the result.

The confidence interval is related to variability.

Variance also related to variability. What is the variance ?

Variance is $(\frac{Var(X)+Var(Y)}{4}) + \frac{Cov(X,Y)}{2}$

If observations are independent then $Cov(X, Y) = 0$ and

Variance is $(\frac{Var(X)+Var(Y)}{4})$ only.

Now imagine we have *dependent observations*.

Now $Cov(X, Y) > 0$. This changes the variance.

For Independent Observations

variance $= (\frac{Var(X) + Var(Y)}{4})$

For Dependent Observations

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Which Variance is larger ?

Which Confidence Interval is larger ?

More independent observations leads to smaller confidence intervals. If observations are *dependent* reduction is not the same ! In example with two observations per mouse (pseudoreplication) observations between the same mouse are dependent. Not the same as independent biological replication ! May however be advantageous to have two samples per mouse, but have to modify the analysis.

After randomization and replication, there is a third

important concept in experimental design, **blocking**.

Imagine we wish to compare the effect of 3 different types of

of fungicide on leaf fungus growth in the tree species.

We find a forest with many trees of that species. We choose

some these trees at random. Each choosen tree is assigned 1

fungicide at random. The fungicide is applied to randomly

choosen leaves which are studied for fungus growth.

What are the experimental units ?

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Treatments ? What are possible problems with this design ?

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Maybe the growth of fungus depends on other

factors, such as moisture, sunlight, *etc.* which vary

a lot between trees and which are hard to measure.

However, within a given tree these factors do not vary

very much. How to solve ?

What are the experimental units ? Observational units ?

Treatments ? What are possible problems with this design ?

Maybe the growth of fungus depends on other

factors, such as moisture, sunlight, *etc.* which vary

a lot between trees and which are hard to measure.

However, within a given tree these factors do not vary

very much. How to solve ? We assign each fungicide at

random to *different* leaves in the same tree ! Why ?

In this new design, what are the treatments,

experimental units and observational units ?

When we treat leaves we first identify subsets of experimental

units which are similar with regards to external factors.

These subsets are *blocks*. Treatment comparisons within

the same block are less affected by external factors.

Differences in treatment are more visible within blocks.

Same treatment difference without blocking would require

larger samples.

Randomization, Replication and Blocking very old concepts.

Discussed in R. A. Fisher; *The Design of Experiments* (1935)

But still very relevant today !

Replication, randomization, and blocking are essential components of any well planned and properly analyzed design. RNA-Seq designs and analyses are no exception

Statistical Design and Analysis of RNA Sequencing Data

Paul Auer & Rebecca Doerge Genetics **185** 405-416 (2010)

Even with randomization, replication and blocking we still

have many options and choices in experimental design.

Simplest choice is CRD ,Completely Randomized Design

(DIC, Delineamento Inteiramente Casualizado). Here we have

a fixed number of experimental units. Select sample sizes.

Assign units at random. If we have 8 units and we decide

we need 5 with treatment A and 3 with B how to implement

a random assignment ? (Each unit must have probaiblity 5/8

to get treatment A and 3/8 to get B).

Can be generalized to more than 2 treatments and

different numbers of units per treatment. Design can

balanced (balanceados) or the same number of units

per treatment or unbalanced (não balanceado).

When the treatment can be thought of as a combined

effect of more than one factor (*eg.* diet and drug)

we have a factorial design. Can be balanced or unbalanced.

If the factor A has three levels and B has 2

then any design in which there is at least

one unit assigned to each of the 6 possible

combinations is a full factorial design. Can be

balanced or unbalanced and can be extended.

Can imagine to be single factor with 6 levels

but not this analysis is not always useful.

Lynch, S. M. & J. J. Strain (1990) Nutrition Research 10, 449-460

6 treatments (3 milk levels and 2 copper levels)

were studied for their effect on rat liver function.
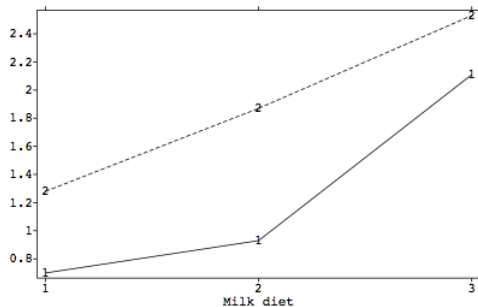
How does the milk level affect the outcome ?

How does the copper level affect the outcome ?

Is there an **interaction** between the two ?

(New feature for factorial designs).

Under the null hypothesis H_0 there is no effect.

and all 6 treatment means are the same. Under H_A

we can consider the main effects

(milk and copper affect outcome independently)

or the interaction effect

(the effect of milk is different depending on the copper level).

Can visualize this on an interaction plot.

Lines not perfectly parallel. Difference in response

at different milk levels may depend on the copper treatment

Need to check the difference of differences by a

statistical test and find the confidence interval. This

analysis is much easier in the two factor picture. Easier

to see how H_0 may be untrue than with an analysis

of 6 different treatments.

The most common design used with blocking is the

Randomized Complete Block Design RCBD

(Delineameno em Blocos Completos Casualizados DBCC)

Each block contains at least one experimental unit

for each treatment. Within each block the assignment

of experimental units to treatments is random.

Block definition is not based on statistics ! Requires some

specialist knowledge of the problem.

We have a cheese factory that works between 6:00 and 18:00

and we need to check that there is no difference in the quality

of the cheese produced from 6:00 to 10:00, 10:00 to 14:00

and 14:00 h to 18:00 h. Fresh milk is supplied everyday.

We know that cheese quality is affected by milk quality.

Is it best to compare cheese produced between

6:00 to 10:00 h on Monday with that produced on Tuesday

between 14:00 and 18:00 ?

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Is it best to compare cheese produced between

6:00 to 10:00 h on Monday with that produced on Tuesday

between 14:00 and 18:00 ? How to define blocks ?

Experimental Design XXX

Sometimes cannot use all treatments under consideration

in all blocks. If we have 3 types of eye drops (A,B,C) and

we wish to compare their effects on human subjects,

natural block is a person with only two eyes !

Blocks are Incomplete and cannot be completed using

a different choice of block. For each block only one

comparison possible. For making all possible comparisons

can use the Balanced Incomplete Block Design.

If we have 4 participants with (A,B) 4 with (B,C) and 4 with (A,C)

example of Balanced Incomplete Block Design.

Sometimes we have two factors with multiple levels and enough experimental units. But random allocation is not possible, for logistical reasons. This leads to the Split Plot Experimental Design. Now have two factors involved, split plot and whole plot factors. Also two levels of experimental unit, split plot and whole plot.

Imagine we have a field in which we wish to test

the effects of irrigation and plant type on yield.

We have two levels of irrigation and 3 plant types.

6 different treatments and 24 plots (experimental units).

However, it is difficult to assign adjacent plots to

different levels of irrigation (logistics). How to we proceed ?

Irrigation Levels I1 and I2, Plant types VA,VB and VC

For logistics must have same I level along each column

I1VA		I2VB		I2VC		I1VA
I1VC		I2VC		I2VA		I1VA
I1VB		I2VA		I2VA		I1VB
I1VA		I2VC		I2VB		I1VC
I1VB		I2VB		I2VB		I1VB
I1VC		I2VA		I2VC		I1VC

Example of a Split Plot Design.

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I1VC		I2VC		I2VA		I1VA
I1VB		I2VA		I2VA		I1VB
I1VA		I2VC		I2VB		I1VC
I1VB		I2VB		I2VB		I1VB
I1VC		I2VA		I2VC		I1VC

Example of a Split Plot Design. Full factorial ?

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Example of a Split Plot Design. Full factorial ? Balanced ?

Irrigation Levels I1 and I2, Plant types VA,VB and VC

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I1VA		I2VC		I2VB		I1VC
I1VB		I2VB		I2VB		I1VB
I1VC		I2VA		I2VC		I1VC

Example of a Split Plot Design. Full factorial ? Balanced ?

Randomize columns for I1 and I2 and then randomize

each column separately for VA, VB and VC.

In this design, Column is the Whole Plot Experimental Unit.

A row within a given column is the Split Plot Experimental Unit.

I is the whole plot factor and V is the split plot factor.

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Good design for comparing the effects of changing V while

keeping I constant.

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Good design for comparing the effects of changing V while

keeping I constant.

How to modify design to keep same whole plot and split plot

experimental units but now use I as split plot factor and

V as whole plot factor ?

Same as before with a CRD

I1VA	I2VB	I1VA	I2VC
I1VC	I1VB	I2VB	I2VC
I2VC	I2VA	I1VA	I1VA
I1VA	I1VB	I2VB	I1VB
I2VB	I2VB	I1VC	I1VA
I2VC	I1VA	I1VC	I2VC

Randomly assign 24 plots, 4 each to 6 treatments

Full factorial ?

Same as before with a CRD

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I1VC		I1VB		I2VB		I2VC
I2VC		I2VA		I1VA		I1VA
I1VA		I1VB		I2VB		I1VB
I2VB		I2VB		I1VC		I1VA
I2VC		I1VA		I1VC		I2VC

Randomly assign 24 plots, 4 each to 6 treatments

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I2VC		I2VA		I1VA		I1VA
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I2VB		I2VB		I1VC		I1VA
I2VC		I1VA		I1VC		I2VC

Randomly assign 24 plots, 4 each to 6 treatments

Full factorial ? Balanced ?

Upto now we have thought of gene expression

in terms of the normal distribution. Observation does

not have to be a whole number and the normal

has mean and variance independent. The mean does

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Poisson Distribution I

Upto now we have thought of gene expression

in terms of the normal distribution. Observation does

not have to be a whole number and the normal

has mean and variance independent. The mean does

not define the value of the variance ! In RNA-Seq

the observations are counts and not continuous.

Need a statistical framework which is designed for counts.

Most straightforward distribution for describing counts

is the binomial distribution. Number of head is 4 or 7 . . .

but under no circumstances 3,6. Not Suitable for RNA-Seq !

Why ?

Poisson Distribution II

Most straightforward distribution for describing counts

is the binomial distribution. Number of head is 4 or 7 . . .

but under no circumstances 3,6. Not Suitable for RNA-Seq !

Why ? The observed value from a Binomial has

an upper limit. (Number of attempts). No such obvious

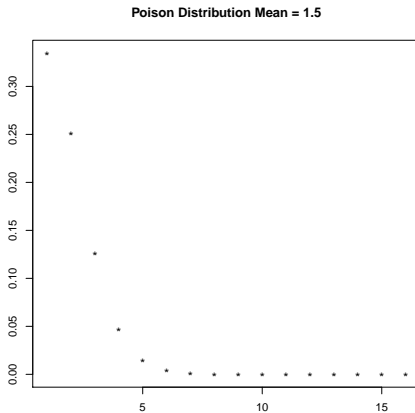upper limit in RNA-Seq data. Need another distribution.

The Poisson Distribution describes count data and has

no upper limit. Outcome can be 0,1,2,3 . . . but only positive

integers. Seems like a good starting point to describe

RNA-Seq. Equation for the Poisson

$$\frac{e^{-\lambda}\lambda^n}{n!}$$

n is the observed value and takes values $0, 1, 2, 3 \ldots$

No upper bound on the value of n. λ is the average value.

Poisson Distribution III

Poison Distribution Mean = 1.5

In previous slide we used $\lambda = 1, 2$

If we take one sample of size 200 from this

distribution what do we expect for the average value ?

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If we take one sample of size 200 from this

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If we take another sample (same size) from the same

distribution what do we expect for the average value ?

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If we take another sample (same size) from the same

distribution what do we expect for the average value ?

Can we expect a different value for a different sample ?

2000 Trials Mean = 1.5